



XENOPHORA

TAXONOMY

Supplément à Xenophora, Bulletin de l'Association Française de Conchyliologie



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XENOPHORA TAXONOMY is a supplement to **XENOPHORA**, the quarterly bulletin published by AFC. It is shipped to all AFC members with the regular publication.

Cover:

Trochidae

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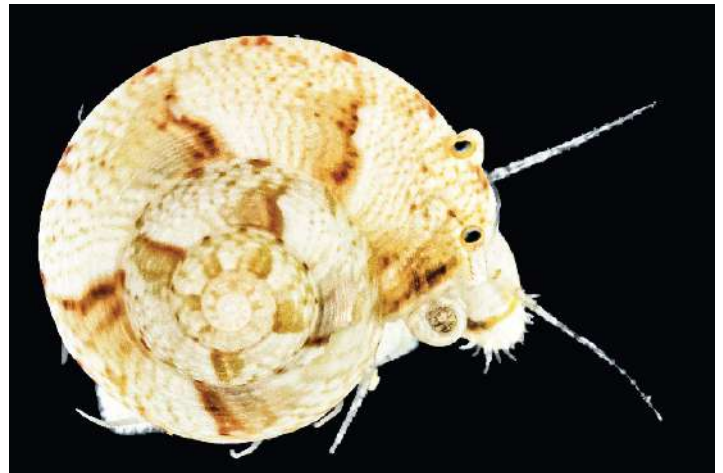
(Souverbie, 1875)

MNHN La planète revisitée

Koumac 2019 KD552 PM652

Photographie : Philippe MAESTRATI (MNHN)

Identification : David HERBERT, juin 2023



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Impression : Groupe IGR NEUVILLE IMPRESSION/IMPRIMERIE SEIC-

☎ 03 85 92 05 29

Espace Neodev - Rue Sainte Elisabeth - 71300 MONTCEAU LES MINES

DESCRIPTION OF *RAPHITOMA ASPERA* SPEC. NOV. (MOLLUSCA, GASTROPODA, RAPHITOMIDAE)
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Keywords

Conoidea, Greece, Mediterranean Sea, new species, *Raphitoma*

Abstract

A new species of the Neogastropoda family Raphitomidae Bellardi, 1875 is described here from the Hellenic waters of the Aegean Sea, and based on 10 specimens. The new species is diagnosed by protoconch and teleoconch characters. It belongs to those species with smooth surface of the teleoconch and bears paucispiral protoconch indicative a lecithotrophic larval development.

Introduction

The family Raphitomidae Bellardi, 1875 is the most diverse of the Conoidea J. Fleming, 1882 (Kantor & Taylor 2002; Bouchet *et al.* 2011). The limits of its type genus *Raphitoma* Bellardi, 1847 in the Mediterranean Sea and the North-eastern Atlantic Ocean have been recently redefined by Fassio *et al.* (2019) to harbour the fossil type species *R. hystrix* Bellardi, 1847 and its surviving descendant *R. pseudohystrix* (Sykes, 1906), some spiny species such as *R. cordieri* (Payraudeau, 1826) and *R. horrida* (Monterosato, 1884), and those that have been once grouped within the genus *Philbertia* Monterosato, 1884 – now accepted as *Raphitoma* Bellardi, 1847. It is represented in the Mediterranean by some 50 species (Prkić *et al.*, 2020), and appears as a morphologically homogeneous group of rather uncommon infralittoral species easily recognizable by their sculpture and colour pattern of the shell (Kontadakis & Manousis, 2022). The under way revision of the Hellenic

members of the genus *Raphitoma* (Manousis *et al.*, 2017; Manousis *et al.*, 2018; Kontadakis *et al.*, 2019; Kontadakis & Manousis, 2022) revealed 29 species for the area (Manousis, 2023). The issue of poecilogony in raphitomids has been also tackled; French researchers commented on the issue as follows: “Une équipe de chercheurs grecs ... s’est lancée à son tour dans cette étude très complexe avec à la base le problème de poecilogonie. Vu la difficulté de faire des différences claires par l’analyse moléculaire, il semblerait que de nombreuses espèces jumelles soient présentes dans notre Méditerranée qui aujourd’hui est devenue le centre mondial de la biodiversité de cette famille”; English translation: “In recent years, a team of Greek researchers ... embarked in turn on this very complex study based on the issue of poecilogony. Given the difficulty of making clear distinctions by molecular analysis, it would seem that many twin species are present in our Mediterranean, which today has become the world centre of the biodiversity of this family” (Pelorce & Horst, 2020). Indeed, after a preliminary publication on the possibility of poecilogony among Mediterranean members of the genus *Raphitoma* (Manousis *et al.*, 2017) the issue was re-approached statistically by means of Orthogonal Regression Analysis (ORA) on published data for congruence with the proposed poecilogony hypothesis, and showed congruence (Manousis *et al.*, 2019). Eventually, in a work aimed at confirming poecilogony in the genus *Raphitoma*, Russini *et al.* (2020) used an integrative approach to species delimitation and applied morphological observations as well as DNA markers analyses to show, though still inconclusively, that poecilogony must be the case. While studding the Hellenic

raphitomids we had the opportunity to examine specimens that could belong to a new species. These samples were collected along the coasts of the Aegean Sea and show features typical of the genus *Raphitoma* and at the same time others that evidently distinguish them from all known Mediterranean congeners. After comparing them with freshly collected similar and sympatric members of the family, we are of the opinion that they represent an undescribed species that is presented below.

Methods

The collection of specimens was conducted from October 2008 to May 2023 in various locations of Greece according to Manousis and Galinou-Mitsoudi (2014). For each specimen collected, the following data have been recorded: location, depth, type of habitat/substrate, size (length, unless otherwise stated), and date. For the species nomenclature updates (June 2023) besides the Marine Biodiversity and Ecosystem Functioning EU Network of Excellence and World Register of Marine Species, the Taxonomic on-line Database on European Marine Mollusca were also used. In addition, the Hellenic Network on Aquatic Invasive Species and the Marine Mediterranean Invasive Alien Species database were used for the alien species status in the Hellenic and Mediterranean Seas. Protoconch whorls were counted according to Verduin (1976), the measurement of the protoconch dimensions was performed at side view with its transition to the teleoconch at the far left, and the measurement of the protoconch maximum diameter was performed at top view according to Gofas and Oliver (2010). The slenderness (h/w) of the shell was estimated including the outer lip of the aperture in the shell's width. The specimens are deposited in the private collections of C. Kontadakis, T. Manousis, G. Mbazios, P. Ovalis, G. Polyzoulis and C. Zeimbekis. Scientists are welcome to have access to the biological material for observation after arrangement. The holotype of *Raphitoma aspera* Kontadakis, Mbazios, Ovalis, Polyzoulis, Zeimbekis, 2023 is deposited at MNHN-IM-2000-38780.

Results

Abbreviations:

CCK – Collection C. Kontadakis; **CCZ** – Collection C. Zeimbekis; **CGMb** – Collection G. Mbazios; **CGP** – Collection G. Polyzoulis; **CPO** – Collection P. Ovalis; **CSG** – Central Saronic Gulf; **CTM** – Collection T. Manousis; **HCS** – Hard Coralligenous Substrate; **Isl.** – Island; **SGFN** – Small Gear Fishing Nets

Raphitoma aspera Kontadakis, Mbazios, Ovalis, Polyzoulis, Zeimbekis, 2023, n. sp., MNHN-IM-2000-38780 (Figs. 1a-d, 2a-d)

Systematics:

Superfamily **Conoidea** J. Fleming, 1822

Family **Raphitomidae** Bellardi, 1875

Genus ***Raphitoma*** Bellardi, 1847

Type Species: *Raphitoma hystrix* Bellardi, 1847⁺ (type by subsequent designation)

Description of holotype: (Fig. 1a) Shell small for the genus. 9.20 mm high and 4.00 mm wide (h/w = 2.30), turritiform, hyaline and translucent. Paucispiral protoconch light brown in colour with whitish nucleus, and regularly cancellated and convex 1.5 whorls. Protoconch I (embryonic shell) 330 µm wide, protoconch II (larval shell) 430 µm wide, exhibiting on its last part a weak keel continuous with the first spiral cord of the teleoconch. The turreted teleoconch consists of 5.5 plano-convex whorls separated by a conspicuous suture. Moderately wide subsutural ramp. The body whorl occupies some 65% of the total height, is conical in shape, and bears 14 orthocline and broad axial ribs with interspaces as wide as the ribs, 13 equal in width and equidistant (but the 1st adapical) spiral cords much thinner than the ribs, 5 of which are situated over the suture and the rest 9 below it to the distinct tail of the shell. The spiral cords in their intersections with the axial ribs are smooth, elongated and curved. The relatively low number of ribs and spirals endow the shell with a coarsely orthogonally cancellated “honeycomb” appearance. The first 2 adapical cords increase rapidly in size and form an oblique and conspicuous shoulder immediately below the suture. The aperture is elongated lanceolate, occupies some 44% of the shells height and exhibits a smooth and slightly sinuous columella in its lower part, angled at its upper part. The anterior siphonal canal is long and wide while the posterior one is deep and narrow. The outer lip exhibits externally the endings of the spiral cords, while internally 7 conspicuous and regularly positioned teeth. Shell surface bright, rather smooth, and with faint incremental scars. Background colour universally dark honey-brown with rare irregularly placed whitish spots. Soft parts unknown.

Type Material examined:

Holotype (Fig. 1a) MNHN-IM-2000-38780: shell, CGMb, h: 9.20 mm, 80-120 m, SGFN, HSC, Lavrio, Attica, March 2007

Paratype 1 (Fig. 1b): shell, CCZ, h: 7.50 mm, 60-80 m, SGFN, HSC, Paros Isl., March 2007

Paratype 2 (Fig. 1c): shell, CGP, h: 8.00 mm, 60-80 m, SGFN, HSC, Alonissos Isl., August 2017

Paratype 3 (Fig. 1d): shell, CPO, h: 8.50 mm, 55 m, SGFN, HSC, CSG, May 2018

Paratype 4 (Fig. 2a): shell, CGMb, h: 7.70 mm, 80-100 m, SGFN, HSC, CSG, May 2018

Paratype 5 (Fig. 2b): shell, CCZ, h: 8.70 mm, 60-80 m, SGFN, HSC, Paros Isl., March 2007

Paratype 6 (Fig. 2c): shell, CGMb, h: 8.00 mm, 80-100 m, SGFN, HSC, Lavrio, Attica, March 2007

Paratype 7 (Fig. 2d): shell, CTM, h: 6.85 mm, 60-80 m, SGFN, HSC, Skyros Isl., August 2017

Other material examined:

One shell, CCK, h: 6.10 mm, 100 m, SGFN, HSC, CSG, January 2014; One shell, CTM, h: 8.85 mm, 60-80 m, SGFN, HSC, Skyros Isl., August 2017

Variability:

Protoconch I observed maximum diameters fall within 2 groups, of some 320 µm and some 410 µm, respectively; Protoconch II maximum diameter 330-470 µm; Number of axial ribs 14-15; Number of spiral cords 13-16; Number of apertural teeth 7-9;

Shell colour light honey-yellow to light honey-brown; White spots infrequent and mainly at the level of the suture.

Type Localities: Alonissos Isl., Central Saronic Gulf, Lavrio, Paros Isl., Skyros Isl..

Distribution: Known only from the type localities.

Habitat: The specimens were collected from hard coralligenous substrates.

Etymology: This species is named “*aspera*” because of its rough sculptural decoration (L. *asper*, rough) (Jaeger, 1972).

Remarks

Raphitoma aspera n. sp. (Figs. 1a-d, 2a-d) can easily be distinguished from its sympatric and non sympatric congenates mainly by its paucispiral protoconch, turritiform and hyaline shell, plano-convex whorls, proportionally high body whorl, small number of axial ribs and spiral cords, elongated crosses over the ribs, coarse cancellated appearance. More specifically, it differs from other paucispiral protoconch species:

R. alida Pusateri & Giannuzzi-Savelli, 2016 in its smaller size, smaller protoconch, plano-convex whorls, equal strength of axial and spiral sculpture, not spinulose tubercles at the intersections, no white spots on the subsutural ramp (Pusateri et al., 2016, Figs. 11–14).

R. farolita F. Nordsieck, 1977 (Fig. 2e) in its not ovate-pupoid contour, not convex whorls, complete lack of microgranules, wide subsutural ramp, lack of large whitish blotches (Giannuzzi Savelli et al., 2018a, Fig. 30).

R. griseomaculata Pusateri & Giannuzzi-Savelli, 2018 (Fig. 2f) in its larger protoconch, its not slender or fusiform shell, the fewer axial ribs and spiral cords, the wider subsutural ramp, its fewer inner denticles of the outer lip of the aperture, and the colour (Giannuzzi Savelli et al., 2018b, Figs. 23–26).

R. papillosa (Pallary, 1904) (Fig. 2g) in its plano-convex whorls, the fewer axial ribs and spiral cords, the wider subsutural ramp, and the colour pattern (Giannuzzi Savelli et al., 2018b, Fig. 29).

R. philberti (Michaud, 1829) (Fig. 2h) in its not slender and fusiform shell, the fewer axial ribs and spiral cords, the wider subsutural ramp, and the colour pattern (Giannuzzi Savelli et al., 2018b, Fig. 28).

R. smriglioi Pusateri & Giannuzzi-Savelli, 2013 (Fig. 2i) in its not slender and fusiform shell, the fewer axial ribs and spiral cords, the wider subsutural ramp, and the colour pattern (Giannuzzi Savelli et al., 2018b, Fig. 27).

R. spadiana Pusateri & Giannuzzi-Savelli, 2012 (Fig. 2j) in its not convex whorls, its not irregular cancellation, its fewer axial ribs and spiral cords, its not tuberculate crosses, and its not fusiform contour (Pusateri et al., 2012, Figs. 8–13).

We should also emphasize that, unlike *R. aspera* n. sp. which exhibits a uniform colouration, *R. spadiana* and similar

looking species such as *R. contigua*, *R. lineolata*, and *R. smriglioi* are characterized by spiral decoration of a lighter shade on a brownish background colour in the interspaces. Due to the possibility of *Raphitoma aspera* n. sp. following poecilogony in its mode of larval development (Manousis et al., 2019; Russini et al., 2020) we also compared it with other planktotrophic protoconch congenates exhibiting similar sculptural decoration, to find out that *Raphitoma aspera* n. sp. differs in its proportionally much higher body whorl, its plano-convex whorls, its general contour and the colour pattern from: ***R. andrehoarui*** Pelorce & Horst, 2020 (Fig. 2k) (Manousis et al., 2018, from synonym); ***R. cordieri*** (Payraudeau, 1826) (Russini et al., 2020, Fig. 6E); ***R. georgiae*** Kontadakis, 2022 (Fig. 2l) (Kontadakis & Manousis, 2022); ***R. hispidella*** Giannuzzi-Savelli & Pusateri, 2019 (Fig. 2m) (Manousis et al., 2018) (Giannuzzi-Savelli et al., 2019, Figs. 11–17); ***R. melitis*** Kontadakis & Mbazios, 2019 (Fig. 2n) (Manousis et al., 2018); ***R. pumila*** (Monterosato, 1890) (Giannuzzi-Savelli et al., 2019, Figs. 1–6, Fig. 7); ***R. radula*** (Monterosato, 1884) (Pusateri et al., 2016, Fig. 10, Figs. 16–22); ***R. stanici*** Prkić, Giannuzzi-Savelli & Pusateri, 2020 (Prkić et al., 2020, Figs. 10A–F, 11A–E, 12A–B, 16A).

Lastly, we wish to emphasize that the observed two groups of protoconch I maximum diameter size of *R. aspera* n. sp., in which the smaller size is that of the larger by a factor of 0.7937, points out to the direction of sequential poecilogony as described by Manousis et al. (2019), and that the new species seems to be a rather rare one as only a very limited number of shells has been collected during fifteen years of intensive material collection and search.

Acknowledgements

The authors are grateful to the anonymous referees for their useful contribution to improve the publication and Virginie Heros of MNHN-Paris for handling the holotype deposition process.

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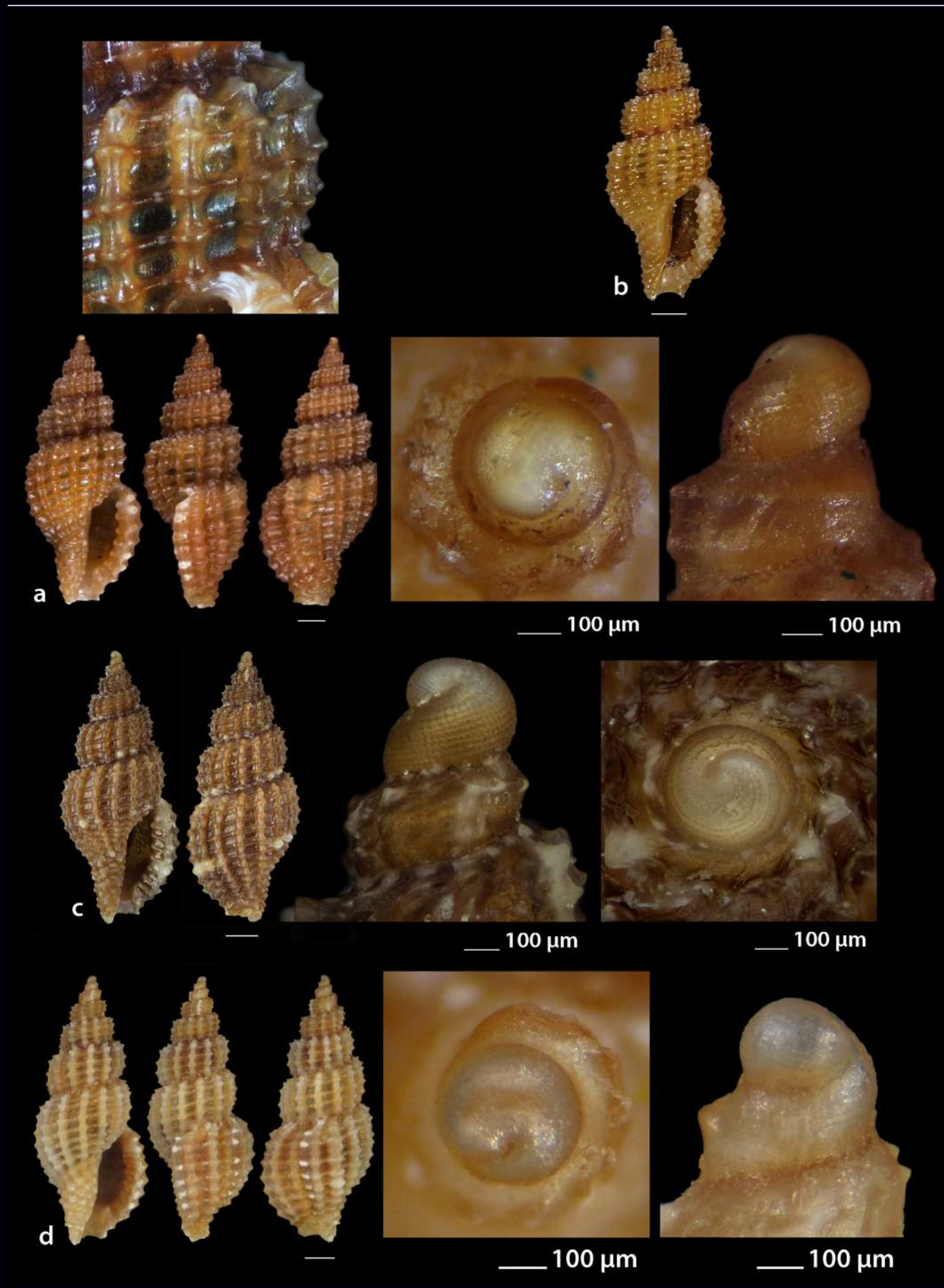


Figure 1: *Raphitoma aspera* n. sp.. a. Holotype, MNHN-IM-2000-38780; b. Paratype 1; c. Paratype 2; d. Paratype 3. Barr = 1 mm, unless otherwise indicated.

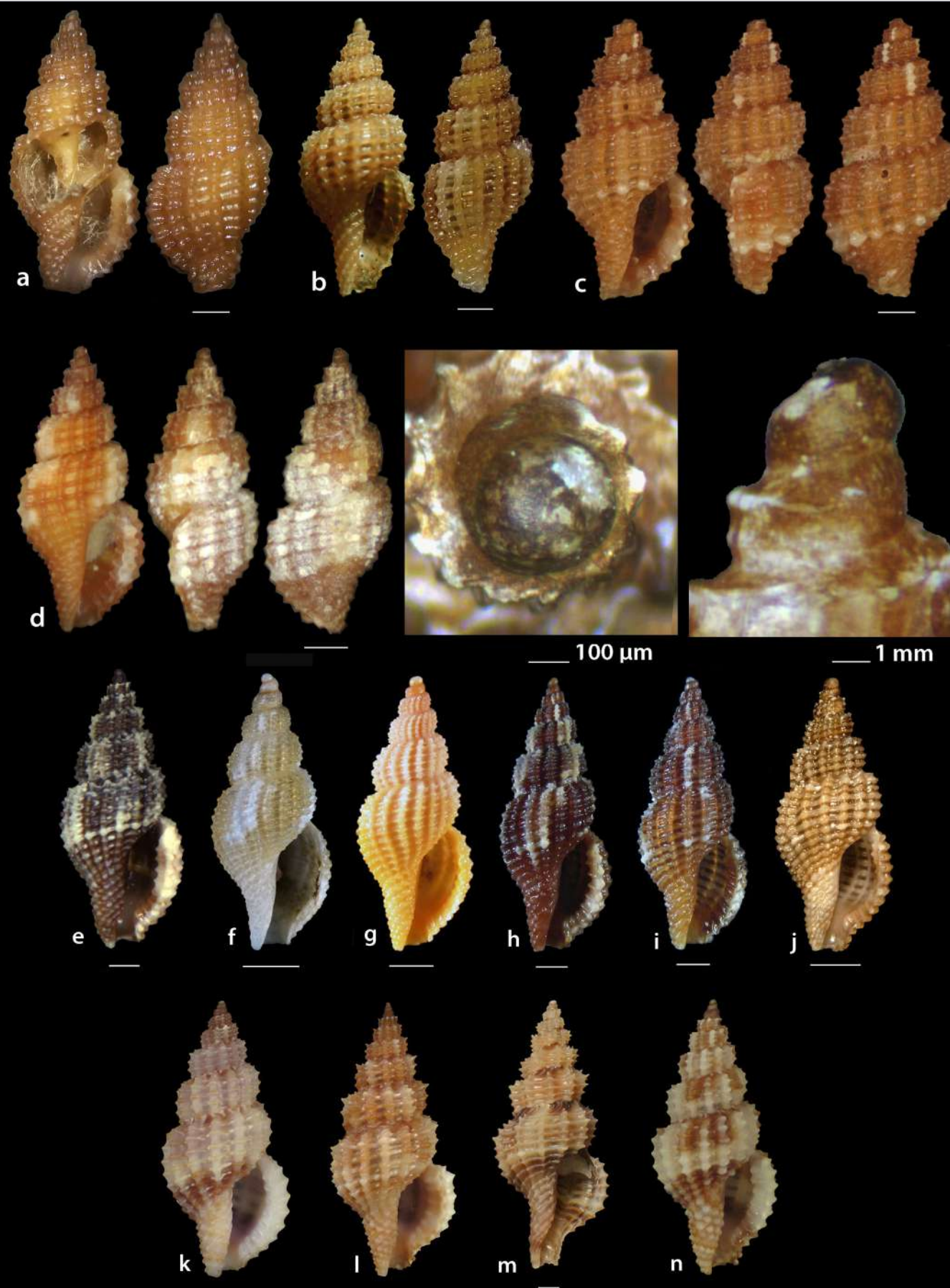


Figure 2: a-d. *Raphitoma aspera* n. sp.. a. Paratype 4; b. Paratype 5; c. Paratype 6; d. Paratype 7; e. *R. farolita* F. Nordsieck, 1977; f. *R. griseomaculata* Pusateri & Giannuzzi-Savelli, 2018; g. *R. papillosa* (Pallary, 1904); h. *R. philberti* (Michaud, 1829); i. *R. smriglioi* Pusateri & Giannuzzi-Savelli, 2013; j. *R. spadiana* Pusateri & Giannuzzi-Savelli, 2012; k. *R. andrehoarui* Pelorce & Horst, 2020; l. *R. georgiae* Kontadakis, 2022; m. *R. hispidella* Giannuzzi-Savelli & Pusateri, 2019; n. *R. melitis* Kontadakis & Mbazios, 2019. Barr = 1 mm, unless otherwise indicated..

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ABSTRACT

Dauciconus (Dauciconus) jacquescolombi Monnier & Limpalaër, 2016, an endemic species of the Lesser Antilles waters (La Martinique, Guadeloupe & Saint Lucia) was synonymized by authors with *Dauciconus (Dauciconus) riosi* (Petuch, 1986) from Brazil.

The same authors also synonymized the French Guyana species *Dauciconus (Dauciconus) massemini* Monnier & Limpalaër, 2016 with the more widespread Caribbean species *Dauciconus (Dauciconus) daucus* (Hwass in Bruguière, 1792), under the form name “*vikingorum*”.

Darioconus rosiae Monnier, Batifoix & Limpalaër, 2018 from Southwest Madagascar is synonymized with *Darioconus behelokensis* (Lauer, 1989) by other authors. These three species have a status listed as “unaccepted” in the online World Register of Marine Species (WoRMS) which is widely used to curate collections in both institutional and private spheres.

We herein repeat the taxonomic arguments that we presented in the original descriptions of these species that justify our opinion of a separation at species level for these taxa so that these taxa can be “accepted” in WoRMS as valid.

RÉSUMÉ

Dauciconus (Dauciconus) jacquescolombi Monnier & Limpalaër, 2016, une espèce endémique des eaux des Petites Antilles (La Martinique, Guadeloupe & Sainte Lucie) a été synonymisée par certains auteurs avec *Dauciconus (Dauciconus) riosi* (Petuch, 1986) du Brésil.

Les mêmes auteurs ont aussi synonymisé l'espèce de Guyane française : *Dauciconus (Dauciconus) massemini* Monnier & Limpalaër, 2016 avec l'espèce Caribéenne plus répandue : *Dauciconus (Dauciconus) daucus* (Hwass à Bruguière, 1792) sous la forme « *vikingorum* ».

Darioconus rosiae Monnier, Batifoix & Limpalaër, 2018 du sud-ouest de Madagascar est regroupé avec *Darioconus behelokensis* (Lauer, 1989) par d'autres auteurs. Ces trois espèces ont à ce jour un statut répertorié comme « non

accepté » dans le Registre mondial, en ligne, des espèces marines (WoRMS), largement utilisé pour conserver les collections des institutions et des particuliers.

Nous réitérons ici les arguments taxonomiques que nous avons développés lors des descriptions originales de ces espèces et qui justifient à notre avis une séparation au niveau des espèces de ces taxa. Ainsi ces taxa peuvent être “acceptés” comme valides dans le WoRMS.

MATERIAL AND METHODS

Taxonomy follows Monnier, Limpalaër, Robin & Roux (2018). We describe shell morphology using the terminology established in Röckel *et al.* (1995) and modified in Monnier, Limpalaër, Robin & Roux (2018). Adult shells were measured with digital calipers, most of the time the measurements were rounded to 0.01 millimeter.

REPOSITORIES

MNHN: Muséum National d'Histoire Naturelle, Paris, France
MNRJ: Museu Nacional Rio de Janeiro, Brazil
MZSP: Museu de Zoologia da Universidade de São Paulo, Brazil
CMNH: Cleveland Museum of Natural History, Cleveland, Ohio, USA
EM: Eric Monnier reference collection, Paris, France
LL: Loïc Limpalaër reference collection, Haudivillers, France
JC: Jacques Colomb reference collection, Marseille, France
DM: David Massemin reference collection, Papeete, Tahiti, French Polynesia

RESULTS & REMARKS

- *Dauciconus (Dauciconus) jacquescolombi*

Michael Rabiller and Georges Richard published an important paper (Xenophora Taxonomy 24 – avril 2019) that reported the results of the KARUBENTHOS 2 expedition on offshore Conidae from Guadeloupe Island.

In this work, *Dauciconus jacquescolombi* and *Dauciconus massemini* are synonymized with other species.

“*Conus jacquescolombi*” is listed as a synonym of “*Conus riosi*” Petuch, 1986. The justification for this taxonomic decision,

translated from French, is that “... on the grounds of a few differences that we do not find on the material we could examine... we will maintain provisionally the name of *Conus riosi* for the material collected at Desirade Island”.

Rabiller and Richard also state that “As there are many pictures in the cited papers, the reader will manage to make his own opinion”.

In fact, there is no refutation there of our arguments. We therefore need to repeat them briefly here:

D. (D.) jacquescolombi from Martinique, Guadeloupe and Saint Lucia is a rare species in collections. Most specimens are from old private collections. The Desirade shell discussed by Rabiller and Richard is one of the most recently collected and confirms the presence of the species in the Guadeloupean waters.

In our publication of 2016, we gathered 26 specimens of *D. (D.) jacquescolombi* from various private and public collections and made careful comparisons with *D. (D.) riosi*.

Shells of *D. (D.) jacquescolombi* are in the average less elongated (relative diameter (RD) of 0.64) than those of *D. (D.) riosi* (average relative diameter of 0.61). They are also more conical (position of maximal diameter (PMD) of 0.95 against 0.92).

Moreover, the protoconch and apex of the first one is invariably pink to red and the apex of the second one is always yellow to orange. We checked this character again on all the material at disposal today that comprises several more specimens from Brazil. We do indeed find again the same apex colour difference and no exceptions appear. The periostraca are also different: smooth in *D. (D.) jacquescolombi* and tufted in *D. (D.) riosi*. Other details are given in the original description.

Two Brazilian species have been described in the last two years: *Dauciconus luizcoutoi* Crabos, Pomponet Oliveira, Pereira de Queiroz & Passos de Almeida, 2022 and *Dauciconus rosapex* Crabos, Pomponet Oliveira, Pereira de Queiroz & Passos de Almeida, 2023. Both have a pink protoconch. Only a handful of shells of these two species are presently known and we have no specimen at hand. Yet the shells have patterns that are very different from both *D. riosi* or *D. jacquescolombi* and also have a pale pink interior of the aperture. They cannot be confused with the Antillean *D. (D.) jacquescolombi*.

Moreover, the colour of the apex proved to be a useful tool to separate the two cryptic species of the genus *Dauciconus*: *Dauciconus (Dauciconus) daucus* (Hwass in Bruguière, 1792) and *Dauciconus (Dauciconus) quasidaucus* (Touitou, Puillandre, Bouchet & Clovel, 2020).

Between 2016 and today no examples of *D. (D.) riosi* or *D. (D.) jacquescolombi* have been collected in the islands and coasts separating Saint Lucia and the waters off Bahia and Espirito Santo States in Brazil, although the northern coasts of Brazil have yielded a fair number of new Cone species during this period.

The synonymy established by Rabiller and Richard is therefore unsupported by the available evidence. We

consider *Dauciconus (Dauciconus) jacquescolombi* Monnier & Limpalaër, 2016 to be a valid species on the grounds of conchological differences noted and a distinct geographical range separation that indirectly indicates that *Dauciconus (Dauciconus) jacquescolombi* and *D. (D.) riosi* are evolutionary independent units.

It is therefore important to recognize the correct identity and specific validity of the species from the French Antilles as an original component of the fauna of the Lesser Antilles. This is also important for conservation purposes.

• *Dauciconus (Dauciconus) massemini*

D. (D.) massemini was described from French Guyana. Several specimens in the type lot had their DNA sampled. Later, a single living specimen was found by the MNHN expedition KARUBENTHOS 2 near Les Saintes, Guadeloupe. The DNA of this specimen fits with the specimens collected in French Guyana (Clovel & Touitou, 2020). This shell represents an important and surprising range extension as the ecological conditions in both localities are very different.

This rare species is characterized by a rather constant olive cream coloration with spiral lines of small dark brown dots, a white mid-body band and a few white blotches. The apex is whitish. The shells of *D. (D.) daucus* (Hwass in Bruguière, 1792) and *D. (D.) quasidaucus* (Touitou, Puillandre, Bouchet & Clovel, 2020) are quite different having a more solid pattern and brighter colour – most often orange – than *D. (D.) massemini*. The protoconchs of *D. (D.) daucus* and *D. (D.) quasidaucus* are respectively pink and yellow. There is little doubt that the two taxa represent different species.

However, Rabiller and Richard state that (translated from French): “On the one hand... the specimens attached at first sight to the latter taxon (*D. (D.) massemini*) (sampled after the mission ACOUPA of MNHN) are in fact some *Conus daucus*. On the other hand and in the same way, the specimens from KARUBENTHOS 2 (sampled too) provisionally attached to *C. vikingorum* are also *C. daucus*. Always having doubts on the distinction between *C. vikingorum*-*C. massemini*, it seems reasonable to us to assign the older name (that of 1993 – *C. vikingorum*) to what remains, in every case, a rare form”.

Two arguments are important in this statement:

1. Molecular phylogenies show that *D. (D.) massemini* is the same as *D. (D.) daucus*.
2. *D. (D.) vikingorum* is also a *D. (D.) daucus*.

On the first argument, molecular phylogenies and trees are not given by Rabiller and Richard. It is therefore impossible to directly check their statement. In fact, they could not publish the phylogenetic trees they used. Fortunately, Touitou *et al.* (2020) published a molecular phylogeny tree (fig 1. p. 41). This tree shows that *D. (D.) daucus* and *D. (D.) quasidaucus* are placed in two different branches. *D. (D.) massemini* is sister to *D. (D.) norai* and then to *D. (D.) boui* and *D. (D.) quasidaucus*. The molecular evidence contradicts Rabiller and Richard's conclusion. *D. (D.) massemini* is distinct from *D. (D.) daucus*.

The second argument is that *D. (D.) massemini* is the same as *D. (D.) vikingorum*. DNA samples from the typical population of

D. (D.) vikingorum have not been collected. So, they cannot be compared. However, the shells do not have the same shapes and colour ranges as we showed in plates 3 and 8 of our 2016 publication and again in plate 2 of this paper. The geographical range of both species is given in a map on page 22 of the original description. It shows a wide separation between both species. The indirect evidence is therefore that two distinct species are involved with separated populations evolving independently. We confirm our opinion that *D. (D.) massemini* is a valid species.

- ***Darioconus rosiae*** Monnier, Batifoix & Limpalaër, 2018

The source of supposed synonymy is the citation listed (Tenorio, M. J. "Illustrated catalog of the living cone shells, 2018 Update supplement". 1-15.).

The citation only provides an opinion by the author that the described taxon: *Darioconus rosiae* Monnier, Batifoix & Limpalaër, 2018 is "...a local form of *Darioconus behelokensis*" without further justification.

Luigi Bozzetti (2021) expressed the same opinion in a note on *Phasmoconus (Phasmoconus) chindeensis* with the words: "Speaking of Madagascar Cones, two recently described taxa: *Conus rosiae* Monnier, Batifoix (sic) & Limpalaër, 2018 and *Conus michelcharlesi* Monnier, Limpalaër & Prugnaud, 2020, have turned out to be synonyms respectively of *Conus behelokensis* Lauer, 1989 and *Conus gracianus* da Motta, 1982". No argument is given as support of this opinion. Consequently, it cannot be discussed or investigated. We will not deal with it and concentrate our efforts on Tenorio's synonymy.

Jean Louis Batifoix (co-author of the description of *D. rosiae*) and the first author made many trips to fishermen villages near Toliara in the Southwest of Madagascar to understand the *Darioconus* populations in the area. Seeing hundreds of these shells, discussing with fishermen and finding juvenile shells with intact protoconchs they were able to map out where the species and their variations occur.

D. behelokensis (Lauer 1989) is found in a very restricted range near the fishermen village of Beheloka at about 40 km. south of the Onilahy river mouth and 60 km South of Toliara.

The pattern and colors of this species are almost constant: the ground color is whitish mostly covered with various amounts of grayish blue and mauve to pale violet shades. The pattern varies very little from orange to chestnut brown with many very fine darker spiral dotted lines regularly spaced on the last whorl. The tents are edged with dark brown.

D. rosiae appears to live in various localities within the great coral reef in the North of Toliara from Manombo to Morombe: Ambatomilo, Beangolo or Andravony. *D. rosiae* has a white ground color sometimes with shades of blue grey or pink. The pattern and color of the shells are much more variable. The shells color varies greatly from light orange, mustard, bright brick-red to very dark brown, and can also vary widely in size. Spiral lines are less numerous, sometimes missing, and less regularly spaced than in *D. behelokensis*. The tents are edged with dark brown too and can vary widely in size (more

in *D. rosiae* than in *D. behelokensis*). *D. rosiae* has a larger average size, is heavier and has a more conical shape than *D. behelokensis*.

Moreover, there are about 100 km between the extreme north location of *D. behelokensis* and the extreme south location of *D. rosiae* with two very large and mighty rivers (Onilahy and Fiherenana Rivers) separating the geographical ranges of the two species. At the estuaries there are no lagoons and therefore no suitable habitat for *Darioconus* species. These features appear to settle a barrier that separates two populations: the smaller in the south is *D. behelokensis*, the northern and more widespread is *D. rosiae*. The wider range seems to be linked with the extent of a continuous lagoon and coral barrier reef.

The argument that *Darioconus rosiae* is a "local form" is in contradiction with the wide geographical range documented in the original description. In fact, *D. behelokensis* is much more "local" than its northern counterpart. Tenorio's argument cannot be retained as a reason for synonymy.

Both species have paucispiral protoconch which is an indication of direct development. This reduces the possibility of genetic exchange between populations separated by wide estuaries. They most likely evolved separately and therefore meet the requirements of distinct valid species.

ACKNOWLEDGEMENTS

We are sincerely grateful to Bill Fenzan for his suggestions and for the reading of the manuscript, Jean-Louis Batifoix for his help and Alain Robin for the photos.

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Plate 1



1 *Dauciconus* (D.) *jacquescolombi* 54.16 mm paratype 15 Fort de France, Martinique, 25-45 m, Coll. EM; 2 *Dauciconus* (D.) *jacquescolombi* 42.80 mm, Dominique, 30 m, Coll. EM; 3 *Dauciconus* (D.) *jacquescolombi* 43.92 mm, Paratype 3, Coll. JC; 4 *Dauciconus* (D.) *jacquescolombi* 51.34 mm, Fort de France, Martinique, 25-45 m, Coll. EM; 5 *Dauciconus* (D.) *riosi* 42.00 mm Guarapari, Espiritu Santo, Brasil, 50-60 m, Coll. EM; 6 *Dauciconus* (D.) *riosi* 57.87 mm, Guarapari, Espiritu Santo, Brasil, 50-60 m, Coll. EM; 7 *Dauciconus* (D.) *riosi* 57.03 mm, Anchieta, Espiritu Santo, Brasil, 60m, Coll. EM; 8 *Dauciconus* *luizcoutoi* 58.1 mm Holotype MZSP 157116 9 *Dauciconus* *rosapex* 63.1 mm, Holotype MNRJ 24489

Plate 2



1 *Dauciconus (D.) massemini* 35.75 mm, Iracoubo, French Guyana, 102-104 m, Holotype MNHN IM 2013-56509; 2 *Dauciconus (D.) massemini* 51.50 mm, French Guyana, paratype 12, private coll; 3 *Dauciconus (D.) massemini* 12.0 mm, French Guyana, paratype 8, coll. DM; 4 *Dauciconus (D.) daucus* 39.82 mm, Anse d'Arlet, Martinique, Coll. EM; 5 *Dauciconus (D.) daucus* 52.20 mm, Malmok, Aruba, 10 m, Coll. EM; 6 *Dauciconus (D.) daucus* 40.67 mm, Martinique, Coll. EM; 7 *Dauciconus (D.) daucus* 32.10 mm, Portobelo, Panama, 80 m, Coll. EM; 8 *Dauciconus (D.) quasidaucus* 53.17 mm, Martinique, Coll. EM; 9 *Dauciconus (D.) vikingorum* 37 mm, Puerto Colombia, Colombia, 35 m Holotype CMNH _ (1); 10 *Dauciconus (D.) vikingorum* 33.7 m Manaure, Guajira, Colombia 45-55m , Coll. EM

Plate 3



1 *Darioconus rosiae* 58.3 mm , Ambatomilo, S.W. Madagascar, Holotype MNHN_IM-2000-33712; 2 *Darioconus rosiae* 58.13 mm , Salary, S.W. Madagascar, Paratype 1, Coll. EM; 3 *Darioconus rosiae* 50.77 mm , Beangolo, S.W. Madagascar, Paratype 2, Coll. EM; 4 *Darioconus rosiae* 62.33 mm , Beangolo, S.W. Madagascar, Paratype 5, Coll. EM; 5 *Darioconus rosiae* 51.02 mm , Salary, S.W. Madagascar, Paratype 8, Coll. EM; 6 *Darioconus rosiae* 59.03 mm , Ambatomilo, S.W. Madagascar, paratype 10, Coll. EM; 7 *Darioconus behelokensis* 48.3 mm , Beheloka, S.W. Madagascar, Holotype MNHN_IM-2000-2581; 8 *Darioconus behelokensis* 50.78 mm Beheloka, S.W. Madagascar, Coll. EM; 9 *Darioconus behelokensis* 53.96 mm Beheloka, S.W. Madagascar, Coll. EM



Phasianellidae
Phasianella solida
(Born, 1778)

MNHN La planète revisitée
Kourmac 2019KM712 LC2322

Photographie: LAURENT CHARLES

Identification: DAVID HERBERT Juin 2023

SPECIES OF THE SUPERFAMILIES TROCHOIDEA (GASTROPODA: ARENEIDAE, CALLIOSTOMATIDAE, COLLONIIDAE, LIOTIIDAE, SKENEIDAE, SOLARIELLIDAE, TROCHIDAE, TURBINIDAE) AND SEGUENZIOIDEA (GASTROPODA: CALLIOTROPIDAE, CATAEGIDAE, CHILODONTAIDAE, SEGUENZIIDAE) DREDGED BY THE R/V PELICAN IN THE GULF OF MEXICO

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KEYWORDS Gulf of Mexico, Trochoidea, Seguenzioidea, geographic distribution, range extensions, habitat.

ABSTRACT This paper reports thirty-three species of the superfamily Trochoidea and eight species of the superfamily Seguenzioidea dredged by the R/V *Pelican* in the Gulf of Mexico; the specimens were dredged off the coasts of Louisiana, Mississippi, Alabama, Florida and Bahía de Campeche, Mexico, between 1996 and 2019. Each species is accompanied by accurate coordinates and depth and, in many cases, type of bottom, the water depth varying from 36 m to 1773 m. A range extension for a number of the species is reported.

INTRODUCTION

This is a continuing report on molluscan species dredged in campaigns organized by the Biology Department at the University of Louisiana at Lafayette on the R/V *Pelican* in the Gulf of Mexico (GOM) between 1996 and 2019. The vessel was operated by the Louisiana Universities Marine Consortium (LUMCON). The history of these campaigns, as well as species belonging to other families, has been reported elsewhere (see García, 2021, 2022a, 2022b, 2023a, 2023b, 2023c).

The superfamily Trochoidea has sustained many changes in the last decades. In their seminal study on this group Hickman & McLean (1990) recognized only three families in the Trochoidea: Turbinidae, Trochidae and Skeneidae; however, they state that “Few superfamilies of marine gastropods rival the Trochacea [herein Trochoidea] in numbers of genera, species, and individuals, and few superfamilies have been more resistant to a stable suprageneric classification” (1990:3). Decades later, Bouchet & Rocroi had a different systematic interpretation of superfamilial placements (2005:244-245), considering some families as belonging to Turbinoidea Rafinesque, 1815.

We use in this paper the latest taxonomy as prescribed by WoRMS (2023), which places five of the families treated here in Trochoidea. Although many of the genera in this superfamily are herbivorous, others (e.g., *Calliostoma*) are carnivorous. Thirty-three species of Trochoidea belonging to eight families are recorded: Arenidae (4), Calliostomatidae (14), Collonidae (1), Liotiidae (1), Skeneidae (1), Solariellidae (5), Trochidae (3) and Turbinidae (4).

The “enigmatic superfamily Seguenziacea [herein Siguenzioidea]” (Quinn, 1983: 725) is composed of small, deep-water species that, because of their habitats and size, are rarely collected and poorly understood. A number of its genera, including *Seguenzia*, were previously placed in Trochoidea, but further anatomical studies made by Quinn (Ibid) have separated this group from the latter. Contents of the stomach of a *Seguenzia* sp. indicate that they are detritivores (Ibid.). We, again, follow

the taxonomy prescribed by WoRMS (2023). Eight species of Siguenzioidea belonging to four families were dredged by the *Pelican*: Calliotropidae (3), Cataegidae (1), Chilodontidae (2), and Seguenziidae (2).

Accurate coordinates are given for all stations, and type of bottom is supplied when available; however, when the species was collected at more than ten stations the locality data are generalized. Mexican records are all from Bahía de Campeche.

Material and methods

All the material used in this paper was dredge by the R/V *Pelican*, and is in the collection of the author.

Three types of dredges were used: a “Boxcore”, or grab, dredge that makes a single bottom catch; a rectangular dredge measuring approximately 1.5 x 1.5 x 0.5 m, used for hard bottom; and the Benthic Skimmer, a very specialized dredge designed for soft sediment in deep water (see García, 2007). The Benthic Skimmer has the general shape of an upper case “I”, with a frontal gape of 3 m, a length of 3.66 m, and a back width of 1.8 m. The central, narrower portion allows for most of the mud to exit the dredge, while biological specimens are safely collected at the back end. After extracting the larger species, contents of the dredge were sifted, first through a quarter inch mesh, and afterwards through a 1/16-inch mesh. The residual sediment was inspected under magnification.

SYSEMATICS

Commented list of collected species

Superfamily Trochoidea
Family Areneidae Gray, 1857

Arene bairdii (Dall, 1889) (Fig. 1). Dredged alive in hard, calcareous bottom; more frequently dredged as empty specimens in soft bottom. ALABAMA: at 3 stations, in 72-108 m. FLORIDA: at 1 station, in 74-81 m. LOUISIANA: at 7 stations, in 60-110 m. MEXICO: Bahía de Campeche, at 4 stations, in 53-94 m.

Arene tricarinata (Stearns, 1872) (Fig. 2). All specimens were dredged empty in soft bottom. ALABAMA: 29°19.62'N, 87°46.19'W, in 97-108 m; 29° 21'N, 87°42'W, in 140 m. FLORIDA: 27°36.5' N, 83°29' W, in 40 m. LOUISIANA: 28°4.57'N, 90°59.99'W, in 87.9 m; 28°06.78'N, 90°55.58'W, in 101-99 m, in mud. MEXICO: 22°16.28'N, 91°30.42'W, in 108-107 m, in mud.

Arene variabilis (Dall, 1889) (Fig. 3). All of the specimens were dredged empty in soft sediment. ALABAMA: at 3 stations, in

100-165 m. LOUISIANA: at 5 stations, in 34-99 m. MEXICO: at 5 stations, in 73-124 m.

Arene venustula Aguayo & Rehder, 1936 (Fig. 4). GOM records were first reported by the *Pelican* campaigns. LOUISIANA: 27°49.24'N 92°53.71'W, in 50-65 m, in rubble, alive. MEXICO: 22°10.42'N, 91°09.55'W, in 46-48 m, in rubble.

Family Calliostomatidae Thiele, 1924 (1847)

Calliostoma barbouri Clench & Aguayo, 1946 (Figs. 5a, 5b). The Bahía de Campeche find was a new record for the Mexican coast. FLORIDA: 28°10.28'N, 84°01.95'W, in 41-41.5 m; 29°43.08'N, 85°53.16'W, in 39-38.7 m. LOUISIANA: 28°38.16'N, 89°33.19'W, in 60-70 m; 27°59.189'N, 91°39.466'W, in 77-79 m, in rubble; 28°02.508'N, 92°26.872'W, in 64 m, in rubble. MEXICO: 21°48.10'N, 92°06.17'W, in 36-46 m.

Calliostoma cf. hirtum Quinn, 1992 (Fig. 6). This single, juvenile specimen is very similar to Quinn's taxon. Quinn's species has been recorded only from the northeastern coast of the Dominican Republic (Quinn, 1992; Rosenberg, 2009). FLORIDA: 24°35.29'N, 83°41.86'W, in 496-509 m.

Calliostoma dentatum Quinn, 1992 (Fig. 7). Previously recorded only from Texas and Louisiana (Rosenberg, 2009; Rosenberg et al., 2009), these were new record for the Mexican coast. MEXICO: 20°51.16'N, 92°26.28'W, in 93-94 m, in granular sediment; 20°51.49'N, 92°21.44'W, in 63-65 m, in mud.

Calliostoma euglyptum (A. Adams, 1854) (Fig. 8). The scarcity of this species in the *Pelican* dredgings is presumably due to its habitat of water shallower than most sampling stations. MEXICO: 22°10.80'N, 91°09.00'W, in 46-48 m, in rubble.

Calliostoma fascians Schwengel & McGinty, 1942 (Figs. 9-10). Specimens from Bahía de Campeche tend to be more carinated than from elsewhere in the GOM; they are similar to the type figure drawn in Schwengel & McGinty, 1942 (p. 15, pl.6, fig.2). ALABAMA: 29°21'N, 87°42'W, in 140 m; 29°31'N, 87°22.75'W, in 165 m; 29°24.43'N, 87°58.63'W, in 74-72 m, in mud; 29°19.62'N, 87°46.19'W, in 97-108 m. LOUISIANA: 28°4.57'N, 90°59.99'W, in 87.9 m. MEXICO: 20°50.22'N, 92°18.91'W, in 53 m, in mud; 22°16.28'N, 91°30.42'W, in 108-107 m, in mud; 20°51.16'N, 92°26.28'W, in 93-94 m, in granular sediment; 20°00.35'N, 92°26.10'W, 73-77m, in mud; 20°51.16'N, 92°26.28'W, in 93-94 m, in granular sediment; 20°52.40'N, 92°24.83'W, in 77-81 m, in mud.

Calliostoma jujubinum (Gmelin, 1791) (Fig. 11). This common species was dredged only twice, presumably because it normally inhabits areas shallower than those sampled by the *Pelican*. FLORIDA: 24°43.33'N, 83°13.48'W, in 61-61.2 m. MEXICO: 22°11.33'N, 91°08.71'W, in 49-53 m, in rubble.

Calliostoma marionae Dall, 1906 (Fig. 12). Two empty specimens were dredged at the Louisiana station, which indicates that the species may not be as rare as it seems in the northern GOM. ALABAMA: 29°15.00'N, 88°20.77'W, in 80 m, in rubble with gorgonia. LOUISIANA: 28°05.49'N, 91°01.96'W, in 57.6-65.9 m, in rubble.

Calliostoma oregon Clench & Turner, 1960 (Fig. 13). The species

is found around the entire GOM but it is seldom collected, presumably because of its deep-water habitat. LOUISIANA: 27°55.879'N; 91°12.839'W, in 203-204 m. MEXICO: 22°36.05'N, 90°27.80'W, in 158- 206 m, in sand/ shells.

Calliostoma cf. oregon Clench & Turner, 1960 (Fig. 14). This may be a very young *C. oregon*; it is illustrated here for the record. MEXICO: 22°11.46'N, 91°08.71'W, in 50-55m, in rubble with sediment.

Calliostoma orion Dall, 1889 (Fig. 15). The most commonly encountered living *Calliostoma* in Bahía de Campeche, usually associated with sponges. MEXICO: 22°16.07'N, 90°42.89'W, in 56-57 m, in rubble; 22°16.08'N, 90°42.89'W, in 54-56 m, in rubble with sponges, alive; 22°17.90'N, 90°35.63'W, in 60-59, in rubble, alive; 22°16.45'N, 90°39.83'W, in 53-55 m, in rubble with sponges and gorgonia, alive; 21°06.467'N, 92°08.466'W, in 49-46 m, inside tube sponge, alive.

Calliostoma pulchrum (C. B. Adams, 1850) (Fig. 16). FLORIDA: 27°36.89'N, 83°21.19'W, in 37 m, in rubble, sand and sponges; 28°15.754'N, 83°44.996'W, in 33.9-33.9 m, alive; 29°26.24'N, 87°34.49'W, in 65-81 m, in rubble; 29°17.75'N, 85°47.18'W, in 60-68 m; 29°26.32'N, 87°34.38'W, in 75-70 m, alive.

Calliostoma roseolum Dall, 1880 (Fig. 17) ALABAMA: 29°21'N, 87°42'W, in 140 m; 29°14'N, 88°15'W, in 122 m; 29°19.62'N, 87°46.19'W, in 97-108 m. FLORIDA: 25°01.354'N, 83°43.559'W, in 82-85 m, in rubble. LOUISIANA: 27°59.141'N, 91°38.832'W, in 91-65 m, in calcareous rubble with some sponges; 28°38.16'N, 89°33.19'W, in 60-70 m; 28°05.009'N, 91°11.365'W, in 110-109 m, in mud with pieces of old wood. MEXICO: 21°51.32'N, 92°03.68'W, in 66-68 m, in mud/shells.

Calliostoma scalenum Quinn, 1992 (Fig. 18). This species was the most commonly dredged *Calliostoma* in Louisiana. FLORIDA: at 3 stations, in 41-68 m. LOUISIANA: at 14 stations in 53 to 91 m, in calcareous rubble on offshore banks.

Calliostoma yucatecanum Dall, 1881 (Fig. 19). In spite of its toponymy, we did not dredge this species during our campaign in Bahía de Campeche, Yucatán Peninsula; however, it has been commonly collected off Contoy Light, northeastern Yucatán. Our easternmost station in Campeche was at 90°21'W; presumably the branch of the Yucatán Current that veers NW towards Louisiana has brought the veligers elsewhere but the southwestern GOM. The specimens dredged off the Louisiana coast are scarce, but are larger and more colorful than those from off Contoy. LOUISIANA: 28°44.76'N, 90°14.15'W, in 27-28 m, in rubble/sponges; 28°05.041'N, 91°01.648'W, in 70-75 m, in rubble; 28°41.486'N, 89°49.912'W, in 90-100 m; 28°39.377'N, 90°19.892'W in 26-26m, alive.

Family Colloniidae Cossman, 1917

Cantrainea macleani Warén & Bouchet, 1993 (Fig. 20). The pictured specimen is a topotype. LOUISIANA: 27°46.904'N 91°30.286'W, in 546-555 m, "Bush Hill" site, in hydrocarbon cold seeps; 27°43.4'N, 91°16.74'W, in 640-680 m.

Family Liotiidae Gray, 1850

"*Cirsotrema*" species (Fig. 21). This is an undescribed species belonging to a genus similar to *Cirsotrema*. It is rather common in calcareous rubble in the northwestern GOM. LOUISIANA: 27°48.7'N, 93°02.88'W, in 55-65 m; 28°05.897'N, 91°02.169'W, in 56 m, in calcareous rubble; 27° 49.24N, 92° 53.71W, in 50-65 m; 28° 0.561'N, 91° 02.205'W, in 58 m; 28°06.066'N, 91°02.418'W, in 57-65 m, in calcareous rubble.

Family Skeneidae W. Clark, 1851

Haplocochlias francesae (Pilsbry & McGinty, 1945) (Fig. 22). Formerly placed in *Parviturbo*, a single specimen was dredged by the *Pelican*. FLORIDA: 26°52.36'N, 83°27.22'W, in 54.1-54.3 m.

Family Solariellidae Powell, 1951

Lamellitrochus lamellosus (Verrill & Smith, 1880) (Fig. 23). ALABAMA: 29°14'N., 88°16'W, in 100 m; 29°21'N, 87°42'W, in 140 m. FLORIDA: 27°48.62'N, 84°45.80'W, in 205- 210.5 m; 24°21.792'N, 83°53.965'W, in 1211 m; several specimens collected alive. LOUISIANA: 27°43.4'N, 91°16.74'W, in 640-680 m using a Boxcore dredge; 27°53.65'N, 91°21.48'W, in 219-221 m, in mud. MEXICO: 22°47.17'N, 90°15.97'W, in 350 m, in sediment.

Solariella cristata Quinn 1992 (Fig. 24). The earlier record for this species in the GOM was the Florida Keys (Rosenberg, 2009); this is the first record for the GOM proper. LOUISIANA: 28°37.86'N, 89°33.25'W, in 68-104 m, in gray mud; 28°38.06'N, 89°33.13'W, in 66 m; 27°53.568'N, 91°21.481'W, in gray mud.

Solariella lacunella (Dall, 1881) (Fig. 25). The records for Louisiana and Mexico are new range extensions. FLORIDA: 27°45.27'N, 84°27.64'W, in 104-108 m; 27°42.71'N, 84°13.09'W, in 68- 68.5 m; 27°45.27'N, 84°27.64'W, in 104-108 m; 29°39.90'N, 86°12.67'W, in 60 m, alive; 29°26.29'N, 87°35.11'W, in 74-81 m, in rubble; 27°51.79'N, 84°59.82'W, in 308-323 m. LOUISIANA: 28° 6.21'N, 91° 2.23'W, in 99.3 m. MEXICO: 20°52.40'N, 92°24.83'W, in 77-81 m, in mud; 20°51.16'N, 92°26.28'W, in 93-94 m, in granular sediment; 20°51.16'N, 92°26.28'W, in 93-94 m, in granular sediment; 22°20.00'N, 90°49.43'W, in 84-89 m.

Solariella multirestis Quinn, 1979 (Fig. 26). Records for this species were formerly confined to the island of St. Vincent, in the Lesser Antilles. *S. multirestis* was collected at five stations in Louisiana, but nowhere else in the GOM. LOUISIANA: 28°38.16'N, 89°33.19'W, in 60-70 m; 27°53.560'N, 91°21.644'W, in 150 m, in mud/shell fragments; 27°53.56'N, 91°21.6'W, in 95-95m; 27°55.14'N, 92°22.71'W, in 168-160 m; 27°53.65'N, 91°21.48'W, in 219-221 m, in mud.

Suavotrochus lubricus (Dall, 1881) (Fig. 27). Formerly placed in the genus *Solariella*. FLORIDA: 27°51.79'N, 84°59.82'W, in 308-323 m; 24°35.29'N, 83°41.86'W, in 496-509 m. LOUISIANA: 27°53.65'N, 91°21.48'W, in 219-221 m, in mud. MEXICO: 22°47.17'N, 90°15.97'W, in 350 m, in sediment.

Family Trochidae Rafinesque, 1815

Gaza superba (Dall, 1881) (Fig. 29). This is a common species in

the appropriate habitat. LOUISIANA: 27°35.22'N, 91°52.31'W, in 600-960 m, in hard mud. MISSISSIPPI: 29°04.110'N, 88°20.647'W, in 475 m, in mud.

"*Margarites*" sp. (Fig. 30). A tentative generic placement for this single specimen. FLORIDA: 27°51.79'N, 84°59.82'W, in 308-323 m.

Family Turbinidae Rafinesque, 1815

Lithopoma phoebium (Röding, 1798) (Fig. 31). This common species normally inhabits shallower waters. Louisiana specimens are characterized by their red pigmentation; no white specimens were collected there. FLORIDA: 24°49.06'N, 83°42.80'W, in 80.7- 81.5m. LOUISIANA: at 13 stations, in 57-80.5 m, in calcareous rubble. MEXICO: 22°05.89'N, 91°22.89'W, in 40-42 m, in grass/sand/rubble; 21°49.30'N, 92°05.59'W, in 38-50 m.

Lithopoma tectum (Lightfoot, 1786) (Fig. 32). Much less common in Louisiana than *L. phoebium*, the few specimens collected tend to have a pinkish hue. LOUISIANA: 28°02.55'N, 92°26.77'W, in 64-67 m, in sand/rubble; 28°03.71'N, 92°27.77'W, in 59-62 m, in medium-size rubble; 27°58.947'N, 91°39.382'W, in 66-79 m, in rubble; 28°05.548'N, 91°01.040W, in 58 m, in rubble.

Turbo cailletii Fischer, 1886 (Figs. 33a, 33b). Mostly collected empty, it is a common species found on top of calcareous banks off the Louisiana coast. FLORIDA: 24°44.77'N, 83°43.71'W, in 70.6- 72.9 m; 24°37.08'N, 83°35.06'W, in 68.4-89.3 m; 24°38.27'N, 83°36.36'W, in 65.6-65.8 m; 26°25.77'N, 83°43.03'W, in 72.9-78.0 m, in granular sand & *Halimeda* algae. LOUISIANA: at 16 stations, in 53-110 m, in calcareous rubble. MEXICO: 21°48.10'N, 92°06.17'W, in 36-46 m, alive.

Turbo castanea Gmelin, 1791 (Fig. 34). A common species collected alive on top of the calcareous surface of the Louisiana offshore banks. LOUISIANA: at 22 stations, in 53-74 m, in calcareous rubble. MEXICO: 22°16.08'N, 90°42.89'W, in 54-56 m, in rubble with sponges.

Superfamily Seguenzioidea Verrill, 1884

Family Calliotropidae Hickman & J. H. McLean, 1990.

Calliotropis cf. globosa Quinn, 1991 (Fig. 35). The author describes this species as "white, nacreous under thin outer porcelainous layer" (Quinn, 1991b); the two live specimens dredged by the *Pelican*, collected in 2006, were of an iridescent golden color as shown in image; in 2023 they are nacreous white. The maximum reported size was 9.7 mm (Rosenberg, 2009); the largest specimen measures 11 mm. *C. globosa* has not been reported from the GOM (Rosenberg et al., 2009); the nearest locality is the state of Quintana Roo, on the Caribbean coast of Yucatán. FLORIDA: 28°44.43'N, 87°30.53'W, in 1773-1699 m, in mud, alive.

Calliotropis lissocona (Dall, 1881) (Fig. 36). Although its type locality is the mouth of the Mississippi River, the species was not found in Louisiana during the *Pelican* campaigns. FLORIDA: 27°51.79'N, 84°59.82'W, in 308- 323 m.

Calliotropis tiara (R. B. Watson, 1879) (Fig. 37). Formerly placed

in *Solariella*, the *Pelican* record was new for the Mexican coast. MEXICO: 22°47.17'N, 90°15.97'W, in 350 m, in sediment.

Family Cataegidae J. H. McLean & Quinn, 1987.

Kanola meroglypta McLean & Quinn, 1987 (Fig. 38). Formerly placed in the genus *Cataegis*, this species is usually associated with hydrocarbon seeps. LOUISIANA: 28°32.088'N, 89°22.396'W, in 688-776m, alive; 28°23.935'N, 89°22.508'W, in 675-765m, in mud, alive; 28°06.52'N, 89°36.99'W, in 850- 610 m, in mud; 27°46.904'N, 91°30.286'W, in 546-555 m; "Bush Hill" site, hydrocarbon cold seeps; 28°25.306'N; 89°33.001'W, in 767-801m, alive.

Family Chilodontaidae Wenz, 1938.

Euchelus guttarosea Dall, 1889 (Fig. 28). The species had been reported in the GOM only from the Florida Keys (Rosenberg, 2009). LOUISIANA: 27°53.65'N, 91°21.48'W, in 219-221 m, in mud.

Mirachelus clinocnemus Quinn, 1979 (Fig. 39). Formerly known in the GOM only from the Florida Keys (Rosenberg, 2009). ALABAMA: 29°19.62'N, 87°46.19'W, in 97-108 m. LOUISIANA: 27°55.14'N, 92°22.71'W, in 168-160 m.

Mirachelus corbis (Dall, 1889) (Fig. 40). This was a new record for the northern GOM. LOUISIANA: 28°4.57'N, 90°59.99'W, in 87.9 m; 27° 59.141'N, 91° 38.832'W, in 91-65 m, in calcareous rubble with some sponges; 28°03.439'N, 92°26.978'W, in 71-74 m, in calcareous rubble. MISSISSIPPI: 28°37.86'N, 89°33.25'W, in 68-104 m.

Family Seguenziidae Verrill, 1884.

Basilissopsis sp. (Fig. 41). The generic placement of this specimen is based on its shell similarity to other specimens of *Basilissopsis*. The genus was originally placed in Trochidae; however, later studies (Quinn, 1991a) transferred it to Seguenziidae. Only one species, *Basilissopsis rhysa* (Dall, 1927), has been reported from the GOM (Rosenberg et al., 2009) but this specimen does not conform to that species. FLORIDA: 27°51.79'N, 84°59.82'W, in 308-323 m.

Seguenzia hapala Woodring, 1928 (Fig. 42). FLORIDA: 27°51.79'N, 84°59.82'W, in 308- 323 m; 24°35.29'N, 83°41.86'W, in 496-509 m; 24°35.29'N, 83°41.86'W, in 496-509 m.

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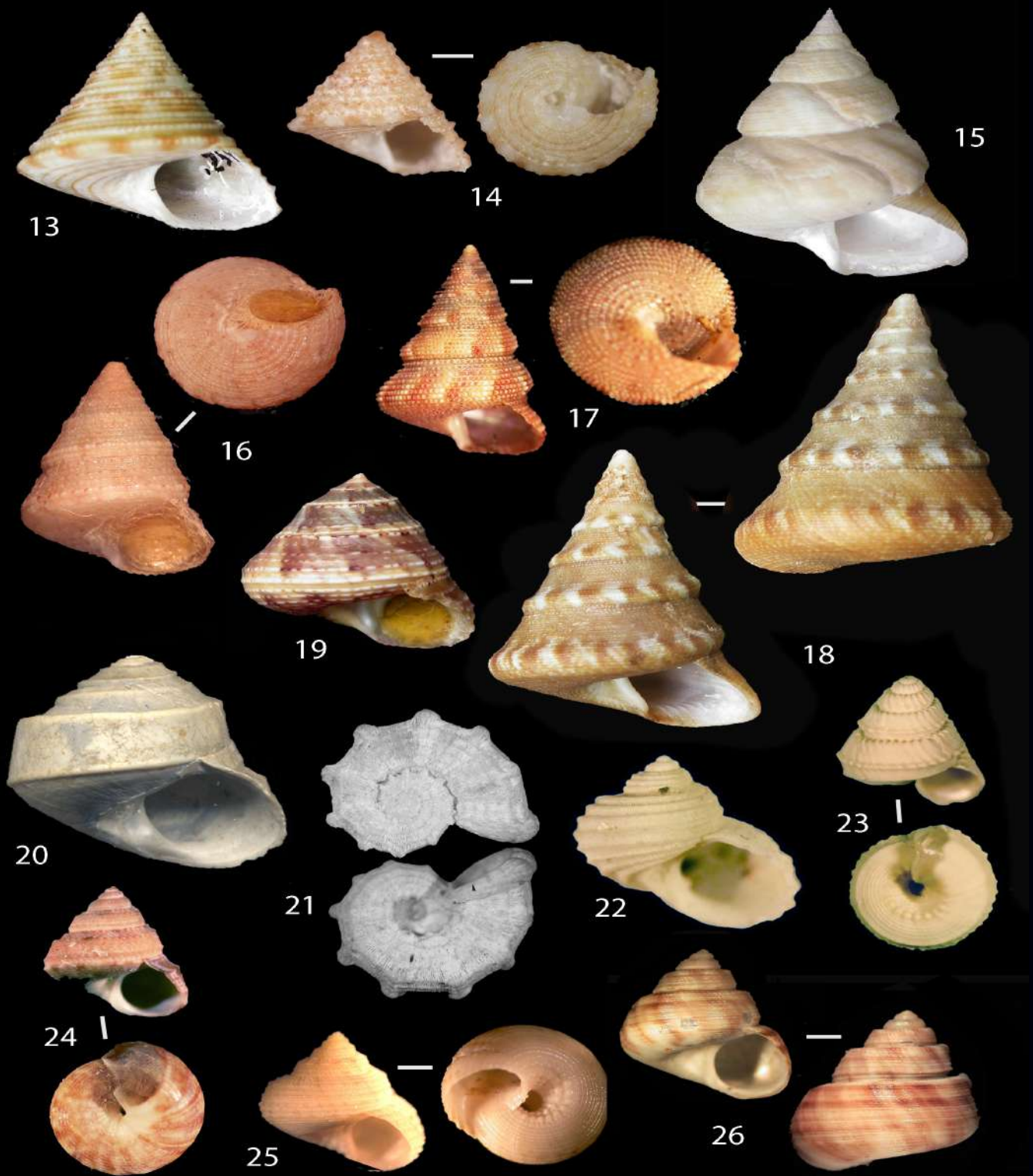
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Plate 1



1. *Arene bairdii* (Dall, 1889) (and operculum), 28°05.009'N, 91°11.365'W, in 110-109 m, 6.6 mm. 2. *Arene tricarinata* (Stearns, 1872), 28°06.78'N, 90°55.58'W, in 101-99 m, 5.5 mm. 3. *Arene variabilis* (Dall, 1889), 28°37.86'N, 89°33.25'W, in 68-104 m, 5 mm. 4. *Arene venustula* Aguayo & Rehder, 1936, 27°49.24'N, 92°53.71'W, in 50-65 m, 7.5 mm, alive. 5a. *Calliostoma barbouri* Clench & Aguayo, 1946, 28°38.16'N, 89°33.19'W, in 60-70 m, 26 mm. 5b. *Calliostoma barbouri* Clench & Aguayo, 1946, 21°48.10'N, 92°06.17'W, in 36-46 m, 7.6 mm. 6. *Calliostoma* cf. *hirtum* Quinn, 1992, 24°35.29'N, 83°41.86'W, in 496-509 m, 4.6 mm. 7. *Calliostoma dentatum* Quinn, 1992, 20°51.49'N, 92°21.44'W, in 63-65 m, in mud, 4.8 mm. 8. *Calliostoma euglyptum* (A. Adams, 1854), 22°10.80'N, 91°09.00'W, in 46-48 m, in rubble, 7 mm. 9. *Calliostoma fascinans* Schwengel, 1942, 29°24.43'N, 87°58.63'W, in 74-72 m, in mud, 6.1 mm. 10. *Calliostoma fascinans* Schwengel, 1942, 20°00.35'N, 92°26.10'W, 73-77m, 9 mm. 11. *Calliostoma jujubinum* (Gmelin, 1791), 22°11.33'N, 91°08.71'W, in 49-53 m, in rubble, 13.2 mm. 12. *Calliostoma marionae* Dall, 1906, 29°15.00'N, 88°20.77'W, in 80 m, in rubble with gorgonia, 24.3 mm.

Plate 2



13. *Calliostoma oregon* Clench & Turner, 1960, 22°36.05'N, 90°27.80'W, in 158-206 m, in sand/ shells, 20.2 mm. 14. *Calliostoma cf. oregon* Clench & Turner, 1960, 22°11.46'N, 91°08.71'W, in 50-55m, in rubble with sediment, 2.9 mm. 15. *Calliostoma orion* Dall, 1889, 22°16.07'N, 90°42.89'W, in 56-57 m, in rubble, 22.2 mm. 16. *Calliostoma pulchrum* (C. B. Adams, 1850), 28°15.754'N, 83°44.996'W, in 33.9-33.9 m, 7.3 mm. 17. *Calliostoma roseolum* Dall, 1880, 25°01.354'N, 83°43.559'W, in 82-85 m, in rubble, 12.3 mm. 18. *Calliostoma scalenum* Quinn, 1992, 28°05.041'N, 91°01.648'W, in 70-75 m, in rubble, 43.4 mm. 19. *Calliostoma yucatecanum* Dall, 1881, 28°44.76'N, 90°14.15'W, in 27-28 m, in rubble/sponges, 17.9 mm, alive. 20. *Cantrainea macleani* Warén & Bouchet, 1993, 27°46.904'N, 91°30.286'W, in 546-555 m; "Bush Hill" site in hydrocarbon cold seeps, 16.3 mm. 21. "*Cirsotrema*" sp., 28°05.897'N, 91°02.169'W, in 56 m, in calcareous rubble, 5.2 mm. 22. *Haplocochlias francesae* (Pilsbry & McGinty, 1945), 26°52.36'N, 83°27.22'W, in 54.1- 54.3 m, 3.1 mm. 23. *Lamellitrochus lamellosus* (Verrill & Smith, 1880), 27°48.62'N, 84°45.80'W, in 205-210.5 m, 3.5 mm. 24. *Solariella cristata* Quinn 1992, 28°38.06'N; 89°33.13'W, in 66 m, 4.2 mm. 25. *Solariella lacunella* (Dall, 1881), 28° 6.21'N, 91° 2.23'W, in 99.3 m, 8 mm. 26. *Solariella multirestis* Quinn, 1979, 27°53.560'N, 91°21.644'W, in 150m, in mud/shell fragments, 10 mm.

Plate 3



27. *Suavotrochus lubricus* (Dall, 1881), 24°35.29'N, 83°41.86'W, in 496-509 m, 3 mm. 28. *Euchelus guttarosea* Dall, 1889, 27°53.65'N, 91°21.48'W, in 219-221 m, in mud, 3 mm. 29. *Gaza superba* (Dall, 1881), 27°35.22'N, 91°52.31'W, in 600-960 m, in hard mud, 37.1 mm. 30. "*Margarites*" sp., 27°51.79'N, 84°59.82'W, in 308-323 m, 2.2 mm. 31. *Lithopoma phoebium* (Röding, 1798), 28°5.7'N, 91°2.15'W, in 60.6 m, 81.7 mm. 32. *Lithopoma tectum* (Lightfoot, 1786), 28°05.548'N, 91°01.040'W, in 58 m, in rubble, 28 mm. 33. *Turbo cailletii* Fischer, 1886, 27°54.895'N, 92°22.594'W, in 60 m, in calcareous rubble: (a) 33.8 mm, (b) 32.3 mm. 34. *Turbo castanea* Gmelin, 1791, 28°05.935'N, 91°02.129'W, in 56.5-56 m, 34.6 mm. 35. *Calliotropis* cf. *globosa* Quinn, 1991, 28°44.43'N, 87°30.53'W, in 1773-1699 m, in mud, 11 mm, alive. 36. *Calliotropis lissocona* (Dall, 1881), 27°51.79'N, 84°59.82'W, in 308-323 m, 2.3 mm. 37. *Calliotropis tiara* (R. B. Watson, 1879), 22°47.17'N, 90°15.97'W, in 350 m, in sediment, 4.5 mm. 38. *Kanola meroglypta* McLean & Quinn, 1987, 28°23.935'N, 89°22.508'W, in 675-765 m, in mud, alive. 39. *Mirachelus clinocnemus* Quinn, 1979, 27°55.14'N, 92°22.71'W, in 168-160 m, 2 mm. 40. *Mirachelus corbis* (Dall, 1889), 28°4.57'N, 90°59.99'W, in 87.9 m, 3.1 mm. 41. *Basilissopsis* sp., 27°51.79'N, 84°59.82'W, in 308-323 m, 1.6 mm. 42. *Seguenzia hapala* Woodring, 1928, 24°35.29'N, 83°41.86'W, in 496-509 m, 5 mm.



Plakobranchidae
Plakobranchus sp.
MNHN La planère revisitée
Koumac 2019
Photographie: KIMBERLEY GARCIA-MENDEZ
Le 24/02/2019

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Keywords

Biodiversity, Greece, Marine Mollusca, Systematic Catalogue

Abstract

This survey provides an updated, systematic catalogue documented with bibliographic and pictorial references on the extant marine Mollusca of the Hellenic Seas by January 2023. It adds 55 extra species for the area and 3 new species for science to the previous taxa register. It incorporates 1479 species belonging to 271 families and referred to in 196 publications. The up-to-date nomenclature is based on the WoRMS Database.

Introduction

The history of the documentation in the form of plain and/or annotated checklists on the Hellenic marine malacofauna has been last presented in detail in a systematic catalogue by Manousis (2021) in *Xenophora Taxonomy*, 34: 26-47. The over 1000 readings of that publication in "ResearchGate" only (accessed, January 2023) constitute an affirmation of its necessity. Since then, some 55 more species new for the area and 3 new for science have been reported and/or image documented in some 43 more publications, while certain taxa names have changed in WoRMS (accessed, January 2023). These later developments are included in the present work with the aim of providing an updated information tool regarding the Hellenic marine malacofauna.

Materials and Methods

All appropriate studies in the last two years depicting specimens collected from the Hellenic Seas were taken into account and added to the previous relevant catalogue (Manousis 2021). As before and for economy of space, the references are numbered in alphabetical and chronological order. The nomenclature followed is that of the WoRMS Database (accessed, January 2023).

Results and discussion

Symbols

! - As drawing
†† - As sp.
‡ - Dubious identification
† - From synonym
? - Inaccurate identification, S. Gofas (WoRMS)
j - Misidentification
* - No image documentation
‡ - Questionable establishment
‡ - Questionable image documentation
‡ - Questionable origin
- Synonym of
x - Unaccepted by WoRMS

This work includes 68 families of Bivalvia with 363 species, 2 families of Caudofoveata with 2 species, 23 families of Cephalopoda with 50 species, 164 families of Gastropoda with 1028 species, 7 families of Polyplacophora with 21 species, 6 families of Scaphopoda with 14 species, and 1 family of Solenogastres with 1 species. In total, 271 families and 1479 species are catalogued and supported by 196 references. There had been optimism that we would "be in the position to reupdate this catalogue in due time to catch up with the fast enriching malacofauna of the area due to more incomers and the finding of as yet unknown local residents" (Manousis, 2021). As a vindication of that optimism, the present work comes to add to that previous catalogue: 7 more species of Bivalvia; 7 more families and 46 more species of Gastropoda; 2 more species of Polyplacophora; 7 more families to the whole; 55 more species to the whole; 43 more references to the whole; 3 more new species for science, namely, *Alvania esmes* Manousis, J. D. Oliver & Zaminos, 2023, *Raphitoma georgiae* Kontadakis, 2022, and *Odostomia widmeri* Tisselli & Manousis, 2022. The overall additions correspond to an increase of 4% in species and 27% in references. Each listed species is accompanied by the references in which is depicted. As before (Manousis 2021): Marked with an asterisk (*) are those references that report but do not depict the relevant species; Dubious identifications due to lack of detailed presentation, misidentifications, species of questionable establishment or questionable documentation and origin are appropriately marked; Accidental and casual records are basically excluded, unless there is no other reference on their finding. The identity of 15 species marked with question mark (?) has been characterized as "inaccurate" by S. Gofas (WoRMS). As those species are depicted in the relevant publications, readers are invited to judge for themselves as to their identity. In accordance with changes in the nomenclature of the taxa, we proceeded to the following changes:

1. *Aegeofusinus angeli* (Russo & Angelidis, 2016) to *Pseudofusus angeli* (Russo & Angelidis, 2016)
2. *Aegeofusinus eviae* (Buzzurro & Russo, 2007) to *Pseudofusus eviae* (Buzzurro & Russo, 2007)
3. *Aegeofusinus margaritae* (Buzzurro & Russo, 2007) to *Pseudofusus margaritae* (Buzzurro & Russo, 2007)
4. *Aegeofusinus patriciae* (Russo & Olivieri, 2013) to *Pseudofusus patriciae* Russo & Olivieri, 2013)
5. *Aegeofusinus profetai* (Nofroni, 1982) to *Pseudofusus profetai* (Nofroni, 1982)
6. *Aegeofusinus rolani* (Buzzurro & Ovalis, 2005) to *Pseudofusus rolani* (Buzzurro & Ovalis, 2005)
7. *Auriculinea bidentata* (Montagu, 1808) to *Leucophytia bidentata* (Montagu, 1808)

8. *Clathrella clathrata* (Philippi, 1844) to *Carinorbis clathrata* (Philippi, 1844)
9. *Fusinus alternatus* Buzzurro & Russo, 2007 to *Pseudofusus alternatus* (Buzzurro & Russo, 2007)
10. *Fusinus dimitrii* Buzzurro & Ovalis in Buzzurro & Russo, 2007 to *Pseudofusus dimitrii* (Buzzurro & Ovalis in Buzzurro & Russo, 2007)
11. *Fusinus parvulus* (Monterosato, 1884) to *Pseudofusus parvulus* Monterosato, 1884
12. *Fusinus pulchellus* (Philippi, 1840) to *Pseudofusus pulchellus* (Philippi, 1849)
13. *Fusinus rusticulus* (Monterosato, 1880) to *Pseudofusus rusticulus* (Monterosato, 1880)
14. *Gracilipurpura rostrata* (Olivi, 1792) to *Pseudofusus rostratus* (Olivi, 1792)
15. *Gyroscala lamellosa* (Lamarck, 1822) to *Gyroscala commutata* (Monterosato, 1877)
16. *Haliotis mykonosensis* Owen, Hanavan & Hall, 2001 to *Haliotis tuberculata tuberculata* Linnaeus, 1758
17. *Haminoea cyanomarginata* Heller & T. E. Thompson, 1983 to *Lamprohaminoea ovalis* (Pease, 1868)
18. *Pogonodon pseudocanaricus* (Bouchet, 1985) to *Ionthoglossa pseudocanarica* (Bouchet, 1985)
19. Genus *Chauvetia* to family CHAUVETIIDAE Kantor, Fedosov, Kosyan, Puillandre, Sorokin, Kano, R. Clark & Bouchet, 2021
20. Genus *Euthria* to family TUDICLIDAE Cossmann, 1901

Acknowledgements

The author is grateful to all the co-authors of his publications on this field of research, and to the unknown referees for their useful contribution to improve the publication.

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Catalogue of species

BIVALVIA	
SOLEMYIDAE Gray, 1840	
<i>Solemya togata</i> (Poli, 1791)	31, 97, 110, 111, 167
NUCULIDAE J. E. Gray, 1824	
<i>Austronucula perminima</i> (Monterosato, 1875)	108, 110, 42
<i>Ennucula aegeensis</i> (Forbes, 1844)	99, 110
<i>Ennucula corbuloides</i> (Seguenza, 1877)	110
<i>Ennucula tenuis</i> (Montagu, 1808)	110
<i>Nucula hanleyi</i> Winckworth, 1931	97, 110
<i>Nucula nitidosa</i> Winckworth, 1930	31, 97, 110, 167
<i>Nucula nucleus</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Nucula sulcata</i> Bronn, 1831	97, 110, 167
MALLETIIDAE H. Adams & A. Adams, 1858 (1846)	
<i>Malletia obtusa</i> (G. O. Sars, 1872)	110
NEILONELLIDAE Schileyko, 1989	
<i>Neilonella latior</i> (Jeffreys, 1876)	75*
NUCULANIDAE H. Adams & A. Adams, 1858	
<i>Ledella messanensis</i> (Jeffreys, 1870)	106, 110
<i>Lembulus pella</i> (Linnaeus, 1758)	31†, 97†, 99, 110, 167†
<i>Nuculana minuta</i> (O. F. Müller, 1776)	97†, 110
<i>Saccella commutata</i> (Philippi, 1844)	31†, 97†, 99, 110, 167†
<i>Saccella illirica</i> (Carrozza, 1987)	97†, 96, 110
PRISTIGLOMIDAE H. L. Sanders & Allen, 1973	
<i>Pristigloma minima</i> (Seguenza, 1877)	106, 110
YOLDIIDAE Dall, 1908	
<i>Microgloma pusilla</i> (Jeffreys, 1879)	110
<i>Microgloma tumidula</i> (Monterosato, 1880)	97i, 106, 110
<i>Yoldia micrometrica</i> G. Seguenza, 1877	108, 110
<i>Yoldiella curta</i> Verrill & Bush, 1898	75*
<i>Yoldiella frigida</i> (Torell, 1859)	75*
<i>Yoldiella lucida</i> (Lovén, 1846)	97i, 110
<i>Yoldiella ovulum</i> La Perna, 2004	106, 110
<i>Yoldiella philippiana</i> (Nyst, 1845)	97, 110
<i>Yoldiella propinqua</i> (Leche, 1878)	106, 110
<i>Yoldiella striolata</i> (Brugnone, 1876)	106, 110
<i>Yoldiella wareni</i> La Perna, 2004	106, 110
ARCIDAE Lamarck, 1809	
<i>Acar clathrata</i> (Defrance, 1816)	31†, 97†, 110, 167†
<i>Acar plicata</i> (Dillwyn, 1817) ?	186
<i>Anadara corbuloides</i> (Monterosato, 1881)	75*, 82, 97i
<i>Anadara gibbosa</i> (Reeve, 1844)	31†, 97†, 110, 167†
<i>Anadara inaequalis</i> (Bruguière, 1789)	75*
<i>Anadara transversa</i> (Say, 1822)	97, 110, 180†, 185†
<i>Arca noae</i> Linnaeus, 1758	31, 83, 97, 110, 167
<i>Arca tetragona</i> Poli, 1795	31, 97, 110, 167†
<i>Asperarca nodulosa</i> (O. F. Müller, 1776)	97, 110, 167†
<i>Asperarca secreta</i> La Perna, 1998	97, 99, 110
<i>Barbatia barbata</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Bathyarca frielei</i> (Friele, 1877)	97i, 110
<i>Bathyarca pentunculoides</i> (Scacchi, 1835)	97, 99, 110, 167
<i>Bathyarca philippiana</i> (Nyst, 1848)	99, 110
NOETIIDAE Stewart, 1930	
<i>Striarca lactea</i> (Linnaeus, 1758)	31, 97, 110, 167

T. MANOUSIS THE MARINE MOLLUSCA OF GREECE (BY JANUARY 2023): AN UPDATED, SYSTEMATIC CATALOGUE

GLYCYMERIDIDAE Dall, 1908	
<i>Glycymeris bimaculata</i> (Poli, 1795)	31†, 97, 110, 167
<i>Glycymeris glycymeris</i> (Linnaeus, 1758)	31, 97i, 110, 167
<i>Glycymeris nummaria</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Glycymeris pilosa</i> (Linnaeus, 1767)	31†, 97, 110, 167
DREISSENIDAE Gray, 1840	
<i>Dreissena polymorpha</i> (Pallas, 1771)	110
MYTILIDAE Rafinesque, 1815	
<i>Arcuatula perfragilis</i> (Dunker, 1857)	97†, 110
<i>Brachidontes pharaonis</i> (P. Fischer, 1870)	108, 110, 167i
<i>Crenella arenaria</i> Monterosato, 1875 ex H. Martin, ms.	97, 99, 110
<i>Crenella decussata</i> (Montagu, 1808)	97, 110
<i>Crenella pellucida</i> (Jeffreys, 1859)	42
<i>Dacrydium hyalinum</i> (Monterosato, 1875)	97, 99, 110
<i>Gregariella petagnae</i> (Scacchi, 1832)	31†, 97, 110, 167†
<i>Gregariella semigranata</i> (Reeve, 1858)	106, 110
<i>Idas argenteus</i> Jeffreys, 1876	110
<i>Idas modiolaeformis</i> (Sturany, 1896)	114
<i>Lioberus ligneus</i> (Reeve, 1858)	106, 110
<i>Lithophaga lithophaga</i> (Linnaeus, 1758)	31, 83, 97, 106, 110, 167
<i>Modiolula phaseolina</i> (Philippi, 1844)	97, 110
<i>Modiolus adriaticus</i> Lamarck, 1819	31, 97, 110, 167
<i>Modiolus barbatus</i> (Linnaeus, 1758)	31, 83, 97, 110, 167
<i>Musculus costulatus</i> (Risso, 1826)	31, 97, 110, 167
<i>Musculus discors</i> (Linnaeus, 1767)	97, 110
<i>Musculus subpictus</i> (Cantraine, 1835)	97†, 110, 167
<i>Mytilaster lineatus</i> (Gmelin, 1791)	96, 110, 167
<i>Mytilaster marioni</i> (Locard, 1889)	96, 97, 110
<i>Mytilaster minimus</i> (Poli, 1795)	31, 97, 110, 167
<i>Mytilaster solidus</i> Monterosato, 1883	97, 110, 167
<i>Mytilus edulis</i> Linnaeus, 1758	97, 110
<i>Mytilus galloprovincialis</i> Lamarck, 1819	31, 83, 97, 110, 167
<i>Rhomboidella prideauxi</i> (Leach, 1815)	114
<i>Septifer cumingii</i> Récluz, 1849	39, 70, 99, 110, 193†*
PINNIDAE Leach, 1819	
<i>Atrina fragilis</i> (Pennant, 1777)	31i, 40*, 97†, 110, 167i
<i>Pinna nobilis</i> Linnaeus, 1758	31, 83, 97, 110, 167
<i>Pinna rudis</i> Linnaeus, 1758	42
MARGARITIDAE Blainville, 1824	
<i>Pinctada imbricata fucata</i> (Gould, 1850)	161*
<i>Pinctada imbricata radiata</i> (Leach, 1814)	31†, 97†, 110, 124, 167†, 168, 195
PTERIIDAE Gray, 1847 (1820)	
<i>Pteria hirundo</i> (Linnaeus, 1758)	31, 97, 110, 167
MALLEIDAE Lamarck 1818	
<i>Malleus regula</i> (Forsskål in Niebuhr, 1775)	49i, 94i, 97†, 110
ISOGNOMONIDAE Woodring, 1925	
<i>Isognomon</i> aff. <i>australicus</i> (Reeve, 1858)	6, 10, 108, 110, 117, 119
<i>Isognomon bicolor</i> (C. B. Adams, 1845)	119
<i>Isognomon legumen</i> (Gmelin, 1791)	10, 110, 117, 118
PROPEAMUSSIIDAE Abbott, 1954	
<i>Cyclopecten brundisiensis</i> Smriglio & Mariottini, 1990	97, 99, 110
<i>Cyclopecten hoskynsi</i> (Forbes, 1844)	99, 110
<i>Parvamussium fenestratum</i> (Forbes, 1844)	110, 167†
<i>Similipecten similis</i> (Laskey, 1811)	97, 110, 167†
PECTINIDAE Rafinesque, 1815	
<i>Aequipecten commutatus</i> (Monterosato, 1875)	97, 110
<i>Aequipecten opercularis</i> (Linnaeus, 1758)	31†, 97, 110, 167

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<i>Delectopecten vitreus</i> (Gmelin, 1791)	97, 110
<i>Flexopecten flexuosus</i> (Poli, 1795)	31†, 110, 167
<i>Flexopecten glaber</i> (Linnaeus, 1758)	31†, 83, 97, 110, 167†
<i>Flexopecten hyalinus</i> (Poli, 1795)	31†, 97†, 110, 167†
<i>Karnekipia sulcata</i> (O. F. Müller, 1776)	97†, 110
<i>Manupecten pesfelis</i> (Linnaeus, 1758)	31†, 97, 110, 167†
<i>Mimachlamys meli</i> Cossignani, 2016	22
<i>Mimachlamys varia</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Palliolium incomparabile</i> (Risso, 1826)	31†, 97, 110, 167†
<i>Palliolium striatum</i> (O. F. Müller, 1776)	97, 110
<i>Pecten jacobaeus</i> (Linnaeus, 1758)	31, 83, 97, 110, 167
<i>Pseudamussium clavatum</i> (Poli, 1795)	31†, 97, 110, 167†
<i>Pseudamussium peslutrae</i> (Linnaeus, 1771)	97, 110
<i>Talochlamys multistriata</i> (Poli, 1795)	31†, 97, 110, 167†
SPONDYLIDAE J. E. Gray, 1826	
<i>Spondylus gaederopus</i> Linnaeus, 1758	31, 97, 110, 167
<i>Spondylus gussonii</i> O. G. Costa, 1830	97, 110
<i>Spondylus</i> cfr. <i>spinosus</i> Schreibers, 1793	97, 99, 110
ANOMIIDAE Rafinesque, 1815	
<i>Anomia ephippium</i> Linnaeus, 1758	31, 97, 110, 167
<i>Heteranomia squamula</i> (Linnaeus, 1758)	97, 110
<i>Pododesmus patelliformis</i> (Linnaeus, 1761)	31, 97, 110, 167
<i>Pododesmus squama</i> (Gmelin, 1791)	97, 99, 110
LIMIDAE d'Orbigny, 1846	
<i>Lima lima</i> (Linnaeus, 1758)	31†, 97, 110, 167
<i>Limaria hians</i> (Gmelin, 1791)	31†, 97, 110, 111, 167†
<i>Limaria loscombi</i> (G. B. Sowerby I, 1823)	97†, 110, 167†, 184
<i>Limaria tuberculata</i> (Olivi, 1792)	31†, 97†, 110, 167†
<i>Limatula gwyni</i> (Sykes, 1903)	99, 110
<i>Limatula subauriculata</i> (Montagu, 1808)	99, 97i, 110, 167†
<i>Limatula subovata</i> (Monterosato, 1875)	99, 110
<i>Limea crassa</i> (Forbes, 1844)	97†, 99, 110
OSTREIDAE Rafinesque, 1815	
<i>Dendostrea</i> cfr. <i>folium</i> (Linnaeus, 1758)	108, 110, 117
<i>Dendostrea frons</i> (Linnaeus, 1758)	117, 119, 193*
<i>Magallana gigas</i> (Thunberg, 1793)	97†, 99, 110
<i>Ostrea edulis</i> Linnaeus, 1758	31, 83, 97, 110, 167
<i>Ostrea stentina</i> Payraudeau, 1826	31†, 97†, 110, 167†
GRYPHAEIDAE Vyalov, 1936	
<i>Neopycnodonte cochlear</i> (Poli, 1795)	31, 97, 110, 167
LUCINIDAE Fleming, 1828	
<i>Ctena decussata</i> (O. G. Costa, 1829)	31, 97, 110, 167
<i>Divalinga arabica</i> Dekker & Goud, 1994	110
<i>Loripes orbiculatus</i> Poli, 1791	31†, 97†, 110, 167†
<i>Loripinus fragilis</i> (Philippi, 1836)	31†, 97†, 110, 167
<i>Lucinella divaricata</i> (Linnaeus, 1758)	97, 110, 167†
<i>Lucinoma borealis</i> (Linnaeus, 1767)	31†, 97, 110
<i>Lucinoma kazani</i> Salas & Woodside, 2002	110
<i>Megaxinus unguiculinus</i> Pallary, 1904	97, 110
<i>Myrtea spinifera</i> (Montagu, 1803)	97, 110, 167
THYASIRIDAE Dall, 1900 (1895)	
<i>Adontorhina similis</i> Barry & McCormack, 2007	106, 110
<i>Axinulus alleni</i> (Carrozza, 1981)	99, 110
<i>Axinulus croulinensis</i> (Jeffreys, 1847)	108, 110
<i>Axinulus exintermedius</i> (Gaglioli, 1992)	106, 110
<i>Axinus grandis</i> (Verrill & S. Smith [in Verrill], 1885)	75*
<i>Genaxinus eumyari</i> (M. Sars, 1870)	110

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<i>Leptaxinus incrassatus</i> (Jeffreys, 1876)	110
<i>Mendicula ferruginosa</i> (Forbes, 1844)	108, 110, 117
<i>Mendicula ockelmanni</i> (Keuning & Schander, 2010)	106, 110
<i>Parathyasira granulosa</i> (Monterosato, 1874)	108, 110
<i>Thyasira biplicata</i> (Philippi, 1836)	99, 110
<i>Thyasira flexuosa</i> (Montagu, 1803)	97, 99, 110
<i>Thyasira obsoleta</i> (Verrill & Bush, 1898)	106, 110
<i>Thyasira sarsi</i> (Philippi, 1845)	75*
<i>Thyasira striata</i> (Sturany, 1896)	97, 110
<i>Thyasira subovata</i> (Jeffreys, 1881)	108, 110
<i>Thyasira succisa</i> (Jeffreys, 1876)	106, 110
UNGULINIDAE Gray, 1854	
<i>Diplodonta brocchii</i> (Deshayes, 1850)	97, 110, 167†j
<i>Diplodonta rotundata</i> (Montagu, 1803)	97, 110
<i>Transkeia bogii</i> (van Aartsen, 2004)	96, 110, 193†*
<i>Transkeia globosa</i> (Forsskål in Niebuhr, 1775)	96, 97†, 110
CHAMIDAE Lamarck, 1809	
<i>Chama asperella</i> Lamarck, 1819	96, 97i, 106, 110, 130
<i>Chama aspersa</i> Reeve, 1846	96, 97, 110, 130
<i>Chama circinata</i> Monterosato, 1878	110, 167
<i>Chama gryphoides</i> Linnaeus, 1758	31, 97, 110, 167
<i>Chama pacifica</i> Broderip, 1835	24, 70, 96, 97, 110
<i>Pseudochama corbierei</i> (Jonas, 1846)	96, 97i, 106, 110
<i>Pseudochama gryphina</i> (Lamarck, 1819)	31, 97, 110, 167
BASTEROTIIDAE Cossmann, 1909	
<i>Basterotia eliptica</i> (Récluz, 1850)	106, 110
<i>Saxicavella jeffreysi</i> Winckworth, 1930	108, 110
GALEOMMATIDAE J. E. Gray, 1840	
<i>Galeomma turtoni</i> W. Turton, 1825	97, 110, 111, 167
<i>Lepirodes politus</i> (Deshayes, 1856)	187†*
<i>Vasconiella jeffreysiana</i> (P. Fischer in de Folin & Périer, 1873)	99, 110
LASAEIDAE Gray, 1842	
<i>Bornia aartseni</i> Gofas, 2012	147
<i>Bornia geoffroyi</i> (Payraudeau, 1826)	75*
<i>Bornia sebetia</i> (O. G. Costa, 1830)	97, 110, 167
<i>Coracuta obliquata</i> (Chaster, 1897)	97i, 99, 110
<i>Devonia perrieri</i> (Malard, 1904)	97, 99, 110
<i>Draculamya uraniae</i> Romani, Bartolini, Oliver & Taviani, 2021	149*
<i>Epilepton clarkiae</i> (W. Clark, 1852)	69, 97, 110, 111
<i>Epilepton parrussetense</i> Giribet & Peñas, 1999	97, 110
<i>Epilepton solidulum</i> (Gagliani, 1992)	106, 110
<i>Epilepton subtrigonum</i> (P. Fischer, 1874)	106, 110
<i>Hemilepton nitidum</i> (W. Turton, 1822)	97, 110
<i>Kellia suborbicularis</i> (Montagu, 1803)	97, 110, 167
<i>Kelliopsis jozinae</i> van Aartsen & Carrozza, 1997	97, 99, 110
<i>Kurtiella bidentata</i> (Montagu, 1803)	97, 109, 110
<i>Kurtiella ovata</i> (Jeffreys, 1881)	109, 110
<i>Kurtiella pellucida</i> (Jeffreys, 1881)	106, 110
<i>Kurtiella triangularis</i> (R. B. Watson, 1897)	109, 110, 114
<i>Kurtiella tumidula</i> (Jeffreys, 1866)	97, 99, 110
<i>Lasaea adansoni</i> (Gmelin, 1791)	97†, 110
<i>Lepton squamosum</i> (Montagu, 1803)	99, 110
<i>Litigiella glabra</i> (P. Fischer in de Folin & Périer, 1873)	69, 97, 99, 110
<i>Montacuta goudi</i> van Aartsen, 1997	97, 99, 110
<i>Montacuta substriata</i> (Montagu, 1808)	97, 110
<i>Phascoliophila phascolionis</i> (Dautzenberg & H. Fischer, 1925)	97†, 99, 110, 167j
<i>Scacchia oblonga</i> (Philippi, 1836)	97, 106, 110
<i>Scacchia zorni</i> van Aartsen & Fehr-de Wal, 1985	106, 110

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<i>Tellimya ferruginosa</i> (Montagu, 1808)	97, 110, 111, 167i
<i>Tellimya semirubra</i> (Gaglini, 1992)	106, 110
<i>Tellimya tenella</i> (Lovén, 1846)	69†, 106, 110
NEOLEPTONIDAE Thiele, 1934	
<i>Neolepton sulcatulum</i> (Jeffreys, 1859)	76†, 97, 110
SPORTELLIDAE Dall, 1899	
<i>Sportella recondita</i> (P. Fischer in de Folin, 1872)	97i, 110
TURTONIIDAE Clark, 1855	
<i>Turtonia minuta</i> (Fabricius, 1780)	110
CARDITIDAE Férussac, 1822	
<i>Cardita calyculata</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Cardites antiquatus</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Centrocardita aculeata</i> (Poli, 1795)	97†, 110, 167†
<i>Coripia corbis</i> (Philippi, 1836)	75*
<i>Glans trapezia</i> (Linnaeus, 1767)	31, 97, 110, 167
ASTARTIDAE d'Orbigny, 1844 (1840)	
<i>Astarte fusca</i> (Poli, 1791)	97, 110, 167
<i>Astarte sulcata</i> (da Costa, 1778)	97, 110, 167
<i>Digitaria digitaria</i> (Linnaeus, 1758)	97, 110
<i>Gonilia calliglypta</i> (Dall, 1903)	97, 110, 167
<i>Goodallia gofasi</i> Giribet & Peñas, 1999	106, 110
<i>Goodallia pusilla</i> (Forbes, 1844)	110
<i>Goodallia triangularis</i> (Montagu, 1803)	97, 110
CARDIIDAE Lamarck, 1809	
<i>Acanthocardia aculeata</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Acanthocardia deshayesii</i> (Payraudeau, 1826)	97, 99, 110
<i>Acanthocardia echinata</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Acanthocardia paucicostata</i> (G. B. Sowerby II, 1834)	31, 97, 110, 167
<i>Acanthocardia spinosa</i> (Lightfoot, 1786)	31, 97, 110, 167
<i>Acanthocardia tuberculata</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Cerastoderma edule</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Cerastoderma glaucum</i> (Bruguière, 1789)	31, 83, 97, 110, 167
<i>Fulvia fragilis</i> (Forsskål in Niebuhr, 1775)	9, 31, 94†, 97, 99, 110, 178
<i>Laevicardium crassum</i> (Gmelin, 1791)	31†, 97, 110, 167†
<i>Laevicardium oblongum</i> (Gmelin, 1791)	31, 97, 110, 167
<i>Papillicardium minimum</i> (Philippi, 1836)	97†, 110, 175†
<i>Papillicardium papillosum</i> (Poli, 1791)	31†, 97†, 110, 167†
<i>Parvicardium carrozzai</i> van Aartsen & Goud, 2001	114, 175
<i>Parvicardium exiguum</i> (Gmelin, 1791)	31, 97, 110, 167
<i>Parvicardium hauniense</i> (Petersen & Russell, 1971) ?	99, 110
<i>Parvicardium pinnulatum</i> (Conrad, 1831)	97, 110, 167†
<i>Parvicardium scabrum</i> (Philippi, 1844)	97, 110, 167†
<i>Parvicardium scriptum</i> (Bucquoy, Dautzenberg & Dollfus, 1892)	96, 97, 110
<i>Parvicardium trapezium</i> Cecalupo & Quadri, 1996	96, 97i, 110, 174
<i>Parvicardium vroomi</i> van Aartsen, Menkhorst & Gittenberger, 1984	75*
<i>Procardium indicum</i> Lamarck, 1819	75*
MACTRIDAE Lamarck, 1809	
<i>Lutraria angustior</i> Philippi, 1844	97, 110
<i>Lutraria lutraria</i> (Linnaeus, 1758)	97, 110
<i>Lutraria oblonga</i> (Gmelin, 1791)	97†, 110
<i>Mactra glauca</i> Born, 1778	31, 97, 110, 167
<i>Mactra stultorum</i> (Linnaeus, 1758)	31†, 110, 167†
<i>Spisula subtruncata</i> (da Costa, 1778)	31, 97, 110, 167
MESODESMATIDAE Gray, 1840	
<i>Donacilla cornea</i> (Poli, 1791)	31, 97, 110, 167
SOLENIIDAE Lamarck, 1809	
<i>Solen marginatus</i> Pulteney, 1799	31, 97, 110, 167
PHARIDAE H. Adams & A. Adams, 1856	

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<i>Ensis ensis</i> (Linnaeus, 1758)	31, 97, 110, 167†
<i>Ensis magnus</i> Schumacher, 1817	97, 110
<i>Ensis minor</i> (Chenu, 1843)	31†, 96, 97, 110, 167
<i>Pharus legumen</i> (Linnaeus, 1758)	31†, 97, 110, 167
<i>Phaxas pellucidus</i> (Pennant, 1777)	97, 110, 111
TELLINIDAE Blainville, 1814	
<i>Arcopagia crassa</i> (Pennant, 1777)	97, 110
<i>Arcopella balaustina</i> (Linnaeus, 1758)	31†, 97, 99, 110, 167†
<i>Asbjornsenia pygmaea</i> (Lovén, 1846)	97†, 110
<i>Bosemprella incarnata</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Fabulina fabula</i> (Gmelin, 1791)	97†, 110, 167†
<i>Gastrana fragilis</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Macomangulus tenuis</i> (da Costa, 1778)	31†, 97†, 110, 167†
<i>Macomopsis cumana</i> (O. G. Costa, 1830)	31†, 97†, 110, 167†
<i>Moerella distorta</i> (Poli, 1791)	31†, 97†, 110, 167†
<i>Moerella donacina</i> (Linnaeus, 1758)	31†, 97†, 110, 111, 167†
<i>Moerella pulchella</i> (Lamarck, 1818)	97†, 110, 167†
<i>Oudardia compressa</i> (Brocchi, 1814)	97†, 110
<i>Peronaea planata</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Peronidia albicans</i> (Gmelin, 1791)	31†, 97†, 110, 167†
<i>Serratina serrata</i> (Brocchi, 1814)	97†, 110, 167†
DONACIDAE J. Fleming, 1828	
<i>Donax semistriatus</i> Poli, 1795	31, 97, 110, 167
<i>Donax trunculus</i> Linnaeus, 1758	31, 83, 97, 110, 167
<i>Donax variegatus</i> (Gmelin, 1791)	31, 97†, 110, 167
<i>Donax venustus</i> Poli, 1795	31, 97, 110, 167
<i>Donax vittatus</i> (da Costa, 1778)	97, 110
PSAMMOBIIDAE J. Fleming, 1828	
<i>Gari costulata</i> (W. Turton, 1822)	31†, 97, 110, 167†
<i>Gari depressa</i> (Pennant, 1777)	31†, 97, 110, 167†
<i>Gari fervensis</i> (Gmelin, 1791)	31†, 97, 110, 167†
<i>Gari tellinella</i> (Lamarck, 1818)	97, 110
SEMELIDAE Stoliczka, 1870	
<i>Abra alba</i> (W. Wood, 1802)	97, 110
<i>Abra longicallus</i> (Scacchi, 1835)	97, 110, 167i
<i>Abra nitida</i> (O. F. Müller, 1776)	97, 110, 111
<i>Abra prismatica</i> (Montagu, 1808)	97, 110, 167i
<i>Abra segmentum</i> (Récluz, 1843)	97, 110, 167†
<i>Abra tenuis</i> (Montagu, 1803)	97, 110, 127
<i>Ervilia castanea</i> (Montagu, 1803)	97, 110
<i>Iacra seychellarum</i> (A. Adams, 1856)	6
<i>Scrobicularia cottardi</i> (Payraudeau, 1826)	97, 110, 167
<i>Scrobicularia plana</i> (da Costa, 1778)	97, 110, 167
SOLECURTIDAE d'Orbigny, 1846	
<i>Azorinus chamasolen</i> (da Costa, 1778)	31, 97, 110, 167
<i>Solecurtus candidus</i> (Brocchi, 1814)	97i, 110, 167†
<i>Solecurtus scopula</i> (W. Turton, 1822)	31, 97, 110
<i>Solecurtus strigilatus</i> (Linnaeus, 1758)	31, 97, 110, 167
KELLIELLIDAE P. Fischer, 1887	
<i>Kelliella miliaris</i> (Philippi, 1844)	110
VESICOMYIDAE Dall & Simpson, 1901	
<i>Isorropodon perplexum</i> Sturany, 1896	97, 106
TRAPEZIDAE Lamy, 1920 (1895)	
<i>Coralliophaga lithophagella</i> (Lamarck, 1819)	97, 99, 110, 167
GLOSSIDAE J. E. Gray, 1847	
<i>Glossus humanus</i> (Linnaeus, 1758)	31, 97, 110, 167
VENERIDAE Rafinesque, 1815	
<i>Callista chione</i> (Linnaeus, 1758)	31, 83, 97, 110, 167

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<i>Chamelea gallina</i> (Linnaeus, 1758)	31, 97i, 110, 167
<i>Chamelea striatula</i> (da Costa, 1778)	97i, 110, 167
<i>Circe scripta</i> (Linnaeus, 1758)†	192*, 183*
<i>Clausinella fasciata</i> (da Costa, 1778)	31†, 96, 97, 110, 167
<i>Clementia papyracea</i> (Gmelin, 1791)	12*, 27
<i>Dosinia exoleta</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Dosinia lupinus</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Globivenus effossa</i> (Philippi, 1836)	31, 97, 110, 167
<i>Gouldia minima</i> (Montagu, 1803)	31, 97, 110, 167
<i>Irus irus</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Lajonkairia lajonkairii</i> (Payraudeau, 1826)	31†, 97†, 110, 167†
<i>Lajonkairia substriata</i> (Montagu, 1808)	97, 110, 167†
<i>Mysia undata</i> (Pennant, 1777)	97, 110, 167
<i>Petricola lithophaga</i> (Retzius, 1788)	31, 97, 110, 167
<i>Petricolaria pholadiformis</i> (Lamarck, 1818)	97, 110, 191
<i>Pitar mediterraneus</i> (Aradas & Benoit, 1872)	97, 110, 167†
<i>Pitar rudis</i> (Poli, 1795)	31, 97, 110, 167
<i>Polititapes aureus</i> (Gmelin, 1791)	31†, 97†, 110, 167†
<i>Polititapes rhomboides</i> (Pennant, 1777)	110, 97†, 167†
<i>Ruditapes decussatus</i> (Linnaeus, 1758)	31†, 83, 97, 110, 167†
<i>Ruditapes philippinarum</i> (Adams & Reeve, 1850)	97, 110
<i>Timoclea ovata</i> (Pennant, 1777)	31, 97, 110, 167
<i>Turtonia minuta</i> (Fabricius, 1780)	97, 110
<i>Venerupis geographica</i> (Gmelin, 1791)	31, 97†, 110, 167
<i>Venus casina</i> Linnaeus, 1758	31, 97, 110, 167
<i>Venus verrucosa</i> Linnaeus, 1758	31, 83, 97, 110, 167
MYIDAE Lamarck, 1809	
<i>Mya arenaria</i> Linnaeus, 1758	97, 110, 186
<i>Sphenia binghami</i> W. Turton, 1822	97, 99, 110
CORBULIDAE Lamarck, 1818	
<i>Lentidium mediterraneum</i> (O. G. Costa, 1830)	97, 110, 167
<i>Varicorbula gibba</i> (Olivi, 1792)	31†, 97†, 110, 167
GASTROCHAENIDAE Gray, 1840	
<i>Cucurbitula cymbium</i> (Spengler, 1783)4 †	31†, 97i, 133*, 167††
<i>Rocellaria dubia</i> (Pennant, 1777)	31†, 97†, 110, 167†
HIATELLIDAE J. E. Gray, 1824	
<i>Hiatella arctica</i> (Linnaeus, 1767)	31†, 97, 110, 167
<i>Hiatella rugosa</i> (Linnaeus, 1767)	97, 110
PHOLADIDAE Lamarck, 1809	
<i>Barnea candida</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Pholadidea loscombiana</i> W. Turton, 1819	97, 110
<i>Pholas dactylus</i> Linnaeus, 1758	31, 97, 110, 167
TEREDINIDAE Rafinesque, 1815	
<i>Bankia gouldi</i> (Bartsch, 1908)	108, 110
<i>Bankia carinata</i> (J. E. Gray, 1827)	97, 99, 110
<i>Lyrodus pedicellatus</i> (Quatrefages, 1849)	97, 99, 110
<i>Nototeredo norvagica</i> (Spengler, 1792)	97, 99, 110
<i>Teredo navalis</i> Linnaeus, 1758	31†, 97, 110, 167†
<i>Teredothyra dominicensis</i> (Bartsch, 1921)	162, 184
XYLOPHAGIDAE Purchon, 1941	
<i>Xylophaga dorsalis</i> (W. Turton, 1819)	97, 99, 110, 167†
<i>Xylophaga praestans</i> E. A. Smith, 1903	97, 110
THRACIIDAE Stoliczka, 1870 (1839)	
<i>Thracia convexa</i> (W. Wood, 1815)	97, 110
<i>Thracia corbuloidea</i> Blainville, 1827	31†, 97†, 110, 167†
<i>Thracia distorta</i> (Montagu, 1803)	97, 99, 110, 167
<i>Thracia phaseolina</i> (Lamarck, 1818)	31†, 97†, 110, 111, 167†
<i>Thracia pubescens</i> (Pulteney, 1799)	31†, 97, 110, 167

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<i>Thracia villosiuscula</i> (MacGillivray, 1827)	97, 110
PERIPLOMATIDAE Dall, 1895	
<i>Cochlodesma praetenu</i> (Pulteney, 1799)	97, 110
CLAVAGELLIDAE d'Orbigny, 1843	
<i>Bryopa aperta</i> (G. B. Sowerby I, 1823)	106, 110
<i>Bryopa melitensis</i> (Broderip, 1834)	75*
PANDORIDAE Rafinesque, 1815	
<i>Pandora inaequalis</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Pandora pinna</i> (Montagu, 1803)	97, 110
LYONSIELLIDAE Dall, 1895	
<i>Allogramma formosa</i> (Jeffreys, 1882)	5†, 110
<i>Lyonsiella compressa</i> Allen & R. D. Turner, 1974	110
<i>Policordia gemma</i> (A. E. Verrill, 1880)	106, 110
LYONSIIDAE P. Fischer, 1887	
<i>Lyonsia arenosa</i> (Møller, 1842)	110
<i>Lyonsia norwegica</i> (Gmelin, 1791)	31, 97, 110, 167†
POROMYIDAE Dall, 1886	
<i>Cetomya neaeroides</i> (Seguenza, 1877)	110
<i>Poromya granulata</i> (Nyst & Westendorp, 1839)	97, 110
VERTICORDIIDAE Stoliczka, 1870	
<i>Haliris berenicensis</i> (Sturany, 1896)	187*
<i>Haliris granulata</i> (Seguenza, 1860)	110
<i>Haliris trapezoidea</i> (Seguenza, 1876)	109, 110
CUSPIDARIIDAE Dall, 1886	
<i>Cardiomya costellata</i> (Deshayes, 1835)	97, 99, 110, 167
<i>Cuspidaria cuspidata</i> (Olivi, 1792)	31, 97, 110, 167
<i>Cuspidaria elliptica</i> Di Geronimo, 1974	75*
<i>Cuspidaria lamellosa</i> (G. O. Sars, 1878)	108, 110
<i>Cuspidaria obesa</i> (Lovén, 1846)	110
<i>Cuspidaria rostrata</i> (Spengler, 1793)	31, 97, 110, 167i
<i>Tropidomya abbreviata</i> (Forbes, 1843)	97, 110
PARILIMYIDAE B. Morton, 1981	
<i>Parilimyia loveni</i> (Jeffreys, 1882)	97, 110
CAUDOFOVEATA	
CHAETODERMATIDAE Théel, 1875	
<i>Falcidens gutturosus</i> (Kowalevsky, 1901)	82*
PROCHAETODERMATIDAE Salvini-Plawen, 1972	
<i>Prochaetoderma raduliferum</i> (Kowalevsky, 1901)	82*
CEPHALOPODA	
SPIRULIDAE Rafinesque, 1815	
<i>Spirula spirula</i> (Linnaeus, 1758)	97, 110
SEPIIDAE Leach, 1817	
<i>Sepia elegans</i> Blainville, 1827	97, 110
<i>Sepia officinalis</i> Linnaeus, 1758	31, 97, 110
<i>Sepia orbignyana</i> Férussac [in d'Orbigny], 1826	97, 110
SEPIOLIDAE Leach, 1817	
<i>Adinaefiola ligulata</i> (Naef, 1912)	75*†, 85*†, 87*†
<i>Heteroteuthis dispar</i> (Rüppell, 1844)	85*, 86*, 89*
<i>Neorossia caroli</i> (Joubin, 1902)	85*, 87*
<i>Rondeletiola minor</i> (Naef, 1912)	85*, 87*
<i>Rossia macrosoma</i> (Delle Chiaje, 1830)	75*, 85*, 87*
<i>Sepietta neglecta</i> Naef, 1916	75*, 87*
<i>Sepietta obscura</i> Naef, 1916	75*
<i>Sepietta oweniana</i> (d'Orbigny, 1841)	75*, 85*, 87*

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<i>Sepiola affinis</i> Naef, 1912	75*, 87*
<i>Sepiola intermedia</i> Naef, 1912	75*, 87*
<i>Sepiola robusta</i> Naef, 1912	75*
<i>Sepiola rondeletii</i> Leach, 1817	75*, 87*
LOLIGINIDAE Lesueur, 1821	
<i>Alloteuthis media</i> (Linnaeus, 1758)	75*, 85*, 173
<i>Alloteuthis subulata</i> (Lamarck, 1798)	93*
<i>Loligo forbesii</i> Steenstrup, 1856	97, 110, 173*
<i>Loligo vulgaris</i> Lamarck, 1798	97, 110
<i>Sepioteuthis lessoniana</i> d'Orbigny, 1826	91
CHTENOPTERYGIDAE Grimpe, 1922	
<i>Ctenopteryx sicula</i> (Vérany, 1851)	85+*
ENOPLUTEUTHIDAE Pfeffer, 1900	
<i>Abralia (Asteroteuthis) veranyi</i> Rüppell, 1844	85*, 173*
HISTIOTEUTHIDAE Verrill, 1881	
<i>Histioteuthis bonnellii</i> (Férussac, 1834)	81, 85*, 89*
<i>Histioteuthis reversa</i> (Verrill, 1880)	85*, 89*, 173*
OCTOPOTEUTHIDAE Berry, 1912	
<i>Octopoteuthis sicula</i> Rüppell, 1844	68†
ONYCHOTEUTHIDAE Gray, 1847	
<i>Ancistroteuthis lichtensteinii</i> (Férussac [in Férussac & d'Orbigny], 1835)	75*, 85*
<i>Onychoteuthis banksii</i> (Leach, 1817)	75*, 85*
PYROTEUTHIDAE Pfeffer, 1912	
<i>Pyroteuthis margaritifera</i> (Rüppell, 1844)	86*, 89*
ANCISTROCHEIRIDAE Pfeffer, 1912	
<i>Ancistrocheirus lesueurii</i> (d'Orbigny [in Férussac & d'Orbigny], 1842)	89*
THYSANOTEUTHIDAE Keferstein, 1866	
<i>Thysanoteuthis rhombus</i> Troschel, 1857	176
CHTENOPTERYGIDAE Grimpe, 1922	
<i>Ctenopteryx sicula</i> (Vérany, 1851)	173*
BRACHIOTEUTHIDAE Pfeffer, 1908	
<i>Brachioteuthis riisei</i> (Steenstrup, 1882)	88*
OMMASTREPHIDAE Steenstrup, 1857	
<i>Illex coindetii</i> (Vérany, 1839)	75, 173*
<i>Ommastrephes bartramii</i> (Lesueur, 1821)	71*, 92, 173*
<i>Todarodes sagittatus</i> (Lamarck, 1798)	97, 110, 173*
<i>Todaropsis eblanae</i> (Ball, 1841)	75, 173*
CHIROTEUTHIDAE Gray, 1849	
<i>Chiroteuthis veranii</i> (Férussac, 1834)	89*
CRANCHIIDAE Prosch, 1847	
<i>Galiteuthis armata</i> Joubin, 1898	85*
BATHYPOLYPODIDAE Robson, 1929	
<i>Bathypolypus sponsalis</i> (P. Fischer & H. Fischer, 1892)	36*
ELEDONIDAE Rochebrune, 1884	
<i>Eledone cirrhosa</i> (Lamarck, 1798)	97, 110
<i>Eledone moschata</i> (Lamarck, 1798)	97, 110
OCTOPODIDAE d'Orbigny, 1840	
<i>Octopus salutii</i> Vérany, 1836	85*, 89*
<i>Octopus vulgaris</i> Cuvier, 1797	31, 97, 110
<i>Macrotritopus defilippi</i> (Vérany, 1851)	90*
<i>Pteroctopus tetracirrhus</i> (Delle Chiaje, 1830)	85*, 86†
<i>Scaevurgus unircirrhus</i> (Delle Chiaje [in Férussac & d'Orbigny], 1841)	85*, 86*
OCYTHOIDAE Gray, 1849	
<i>Ocythoe tuberculata</i> Rafinesque, 1814	19, 59, 75*
ARGONAUTIDAE Cantraine, 1841	
<i>Argonauta argo</i> Linnaeus, 1758	20, 31, 97, 110, 167
TREMOCTOPODIDAE Tryon, 1879	
<i>Tremoctopus violaceus</i> delle Chiaje, 1830	75*, 177

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GASTROPODA	
PATELLIDAE Rafinesque, 1815	
<i>Patella caerulea</i> Linnaeus, 1758	31, 97, 110, 167†
<i>Patella ferruginea</i> Gmelin, 1791	97, 110
<i>Patella rustica</i> Linnaeus, 1758	31†, 97, 110, 167
<i>Patella ulyssiponensis</i> Gmelin, 1791	31†, 97, 110, 167†
NACELLIDAE Thiele, 1891	
<i>Cellana radiata</i> (Born, 1778)	110
LOTTIIDAE Gray, 1840	
<i>Lottia pelta</i> (Rathke, 1833)	108, 110
<i>Tectura virginea</i> (O. F. Müller, 1776)	97, 110, 167†
LEPETIDAE Gray, 1850	
<i>Iothia fulva</i> (O. F. Müller, 1776)	102, 110
<i>Propilidium exiguum</i> (W. Thompson, 1844)	102, 110
FISSURELLIDAE Fleming, 1822	
<i>Diodora demartinorum</i> Buzzurro & Russo, 2005	4, 110
<i>Diodora dorsata</i> (Monterosato, 1878)	4, 97i, 110
<i>Diodora funiculata</i> (Reeve, 1850)	102, 110
<i>Diodora giannispadai</i> Aissaoui, Puillandre & Bouchet, 2017	4, 97i, 110
<i>Diodora gibberula</i> (Lamarck, 1822)	4, 31, 97, 110, 167
<i>Diodora graeca</i> (Linnaeus, 1758)	4, 31, 97, 110, 167
<i>Diodora italica</i> (Defrance, 1820)	4, 31, 97, 110, 167
<i>Diodora producta</i> (Monterosato, 1880)	110
<i>Emarginula adriatica</i> O. G. Costa, 1830	97, 110, 167†i
<i>Emarginula bonfittoi</i> Smriglio & Mariottini, 2001	110
<i>Emarginula decorata</i> Deshayes, 1863	100, 110
<i>Emarginula huzardii</i> Payraudeau, 1826	31, 97, 110, 167
<i>Emarginula octaviana</i> Coen, 1939	31†, 97, 110, 167†
<i>Emarginula punctulum</i> Piani, 1980	97i, 110
<i>Emarginula rosea</i> Bell, 1824	31, 97, 110, 167
<i>Emarginula sicala</i> J.E. Gray, 1825	31, 97, 110, 167
<i>Emarginula solidula</i> O. G. Costa, 1829	97, 110
<i>Emarginula tenera</i> Locard, 1891	110, 116
<i>Fissurella nubecula</i> (Linnaeus, 1758)	31, 97, 110, 167†
<i>Fissurisepta granulosa</i> Jeffreys, 1883	102, 110
<i>Puncturella noachina</i> (Linnaeus, 1771) ?	97, 110
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<i>Anatoma aspera</i> (Philippi, 1844)	110, 167†
<i>Anatoma crispata</i> (J. Fleming, 1828)	97, 110
<i>Anatoma micalii</i> Geiger, 2012	100, 110
<i>Anatoma tenuisculpta</i> (Seguenza, 1880)	102, 110
<i>Scissurella costata</i> d'Orbigny, 1824	97, 110, 167
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<i>Sinezona cingulata</i> (O. G. Costa, 1861)	97, 110, 167†
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<i>Sinezona semicostata</i> Burnay & Rolán, 1990	147
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<i>Haliotis mykonosensis</i> Owen, Hanavan & Hall, 2001 ^x = <i>H. tuberculata</i> Linnaeus, 1758	18†, 97†, 110†, 131†
<i>Haliotis tuberculata</i> Linnaeus, 1758	31†, 97, 110, 167†
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<i>Lepetella barrajoni</i> Dantart & Luque, 1994	114
<i>Lepetella espinosae</i> Dantart & Luque, 1994	97†, 110
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<i>Putzeysia wiseri</i> (Calcara, 1842)	110
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<i>Callumbonella suturalis</i> (Philippi, 1836)	152
<i>Clanculus corallinus</i> (Gmelin, 1791)	31, 97, 110, 167
<i>Clanculus cruciatus</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Clanculus jussieui</i> (Payraudeau, 1826)	31, 97, 110, 167
<i>Clelandella miliaris</i> (Brocchi, 1814)	97, 110
<i>Gibbula albida</i> (Gmelin, 1791)	31, 97, 110, 167†
<i>Gibbula ardens</i> (Salis Marschlin, 1793)	31, 97, 110, 167
<i>Gibbula drepanensis</i> (Brugnone, 1873)	97, 110
<i>Gibbula fanulum</i> (Gmelin, 1791)	31, 97, 110, 167
<i>Gibbula guttadauri</i> (Philippi, 1836)	97, 110, 167†
<i>Gibbula leucophaea</i> (Philippi, 1836)	110, 167
<i>Gibbula magus</i> (Linnaeus, 1758)	31, 97, 110, 167†
<i>Gibbula philberti</i> (Récluz, 1843)	97i, 110, 167
<i>Gibbula ricketti</i> (Payraudeau, 1826)	97i, 110
<i>Gibbula turbinoides</i> (Deshayes, 1835)	97, 110, 167
<i>Gibbula vimontiae</i> Monterosato, 1884	102, 110, 117
<i>Jujubinus exasperatus</i> (Pennant, 1777)	31, 97, 110, 167
<i>Jujubinus gravinae</i> (Dautzenberg, 1881)	97, 110
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<i>Jujubinus montagui</i> (Wood, 1828)	97, 110, 167
<i>Jujubinus striatus</i> (Linnaeus, 1758)	31, 97†, 110, 118, 167
<i>Jujubinus unidentatus</i> (Philippi, 1844)	97, 110, 167
<i>Phorcus articulatus</i> (Lamarck, 1822)	31†, 97†, 110, 167†
<i>Phorcus mutabilis</i> (Philippi, 1851)	31†, 97, 110, 167†
<i>Phorcus richardi</i> (Payraudeau, 1826)	31†, 97, 110, 167†
<i>Phorcus turbinatus</i> (Born, 1778)	31†, 97†, 110, 167†
<i>Steromphala adansonii</i> (Payraudeau, 1826)	31†, 97†, 110, 167†
<i>Steromphala adriatica</i> (Philippi, 1844)	11†, 97†, 110
<i>Steromphala cineraria</i> (Linnaeus, 1758)	97†, 110
<i>Steromphala divaricata</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Steromphala nebulosa</i> (Philippi, 1849)	110
<i>Steromphala rarilineata</i> (Michaud, 1829)	31†, 97†, 110, 167†
<i>Steromphala spratti</i> (Forbes, 1844)	31†, 110, 167†
<i>Steromphala umbilicaris</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Steromphala varia</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
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<i>Calliostoma conulus</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Calliostoma funiculatum</i> Ardevini, 2011	110, 116
<i>Calliostoma granulatum</i> (Born, 1778)	31, 97, 110, 167
<i>Calliostoma gualterianum</i> (Philippi, 1848)	31, 110, 167
<i>Calliostoma kochi</i> Pallary, 1902	110
<i>Calliostoma laugierii</i> (Payraudeau, 1826)	31, 97, 110, 167
<i>Calliostoma virescens</i> Coen, 1933	97, 110
<i>Calliostoma zizyphinum</i> (Linnaeus, 1758)	31, 97, 110, 167
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<i>Akritogyra conspicua</i> (Monterosato, 1880)	102, 110
<i>Cirsonella romettensis</i> (Granata-Grillo, 1877)	102, 110
<i>Dasyskenea suavis</i> Fasulo & Cretella, 2003	108, 110
<i>Dikoleps marianae</i> Rubio, Dantart & Luque, 1998	110, 116
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<i>Dikoleps nitens</i> (Philippi, 1844)	75*, 97

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<i>Dikoleps umbilicostriata</i> (Gaglini, 1987)	109, 110
<i>Lissomphalia bithynoides</i> (Monterosato, 1880)	102, 110
<i>Mikro giustii</i> (Bogi & Nofroni, 1989)	108, 110
<i>Parviturbo alboranensis</i> Peñas & Rolán, 2006	75*
<i>Parviturbo fenestratus</i> (Chaster, 1896)	75*
<i>Skenea catenoides</i> (Monterosato, 1877)	110
<i>Skenea serpuloides</i> (Montagu, 1808)	110, 116
<i>Skeneoides exilissima</i> (Philippi, 1844)	97, 100, 110
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<i>Trochaclis versiliensis</i> Warén, Carrozza & Rocchini in Warén, 1992	108, 110
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<i>Danilia costellata</i> (O. G. Costa, 1861)	31i, 102, 110
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<i>Bolma rugosa</i> (Linnaeus, 1767)	31†, 97, 110, 167†
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<i>Tricolia deschampsii</i> Gofas, 1993 ?	100, 110
<i>Tricolia pullus</i> (Linnaeus, 1758)	31†, 97, 110, 167
<i>Tricolia speciosa</i> (Megerle von Mühlfeld, 1824)	31†, 97, 110, 167
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<i>Nerita sanguinolenta</i> Menke, 1829	110
<i>Smaragdia souverbiana</i> (Montrouzier in Souverbie & Montrouzier, 1863)	39, 102, 110, 194
<i>Smaragdia viridis</i> (Linnaeus, 1758)	31, 97, 110, 167
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<i>Bittium lacteum</i> (Philippi, 1836)	97, 110, 167
<i>Bittium latreillii</i> (Payraudeau, 1826)	97, 110, 167
<i>Bittium reticulatum</i> (da Costa, 1778)	31, 97, 110, 167†
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<i>Cerithium alucastrum</i> (Brocchi, 1814)	31†, 97, 110, 167†
<i>Cerithium litteratum</i> (Born, 1778)	45*
<i>Cerithium lividulum</i> Risso, 1826	97, 110
<i>Cerithium protractum</i> Bivona Ant. in Bivona And., 1838	97, 110, 167†
<i>Cerithium renovatum</i> Monterosato, 1884	97, 110, 127, 167†
<i>Cerithium scabridum</i> Philippi, 1848	39, 70, 97, 110, 127, 190
<i>Cerithium vulgatum</i> Bruguière, 1792	31†, 97, 110, 167†
<i>Rhinoclacis kochi</i> (Philippi, 1848)	94, 102, 110
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<i>Finella pupoides</i> A. Adams, 1860	110
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<i>Fossarus ambiguus</i> (Linnaeus, 1758)	97, 110, 167
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<i>Pirenella conica</i> (Blainville, 1829)	31†, 97†, 110, 167
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<i>Mesalia mesal</i> (Deshayes, 1843) ?	97†, 110
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<i>Turritella turbona</i> Monterosato, 1877	31†, 97, 110
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<i>Euthymella colzumensis</i> (Jousseaume, 1898)	10, 37, 108, 110
<i>Ionthoglossa pseudocanarica</i> (Bouchet, 1985)	110†
<i>Marshallora adversa</i> (Montagu, 1803)	97, 110, 167††
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<i>Monophorus erythrosoma</i> (Bouchet & Guillemot, 1978)	97, 110
<i>Monophorus perversus</i> (Linnaeus, 1758)	31†, 97, 110, 167†
<i>Monophorus thiriota</i> Bouchet, 1985	100, 110
<i>Obesula marisnostris</i> Bouchet, 1985	97, 100, 102, 110
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<i>Strobiliger brychia</i> (Bouchet & Guillemot, 1978)	102, 110
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<i>Viriola corrugata</i> (Hinds, 1843)	118
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<i>Cerithiopsis barleei</i> Jeffreys, 1867	97, 110, 115, 167†‡
<i>Cerithiopsis carlottae</i> Cevalupo & Perugia, 2018	115, 184
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<i>Cerithiopsis fayalensis</i> R. B. Watson, 1880	97, 100, 110, 115, 167†‡
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<i>Cerithiopsis jeffreysi</i> R. B. Watson, 1885	102, 110, 115
<i>Cerithiopsis ladae</i> Prkic & Buzzurro, 2007	102, 110, 115
<i>Cerithiopsis micalii</i> (Cevalupo & Villari, 1997)	97, 100, 110, 115
<i>Cerithiopsis minima</i> (Brusina, 1865)	97, 102, 110, 115, 167†
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<i>Cerithiopsis pulchresculpta</i> Cachia, Mifsud & Sammut, 2004	108, 110, 115
<i>Cerithiopsis pulvis</i> (Issel, 1869)	28, 102, 110, 115, 116
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<i>Cerithiopsis tenthrenois</i> (Melvill, 1896)	28, 97i
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<i>Costulopsis denticulata</i> (Cevalupo & Robba, 2010)	102, 110, 115
<i>Costulopsis nana</i> Jeffreys, 1867	97†, 100, 102, 110, 115
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<i>Dizoniopsis coppolae</i> (Aradas, 1870)	97, 110, 115, 167†‡
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<i>Krachia cylindrata</i> (Jeffreys, 1885)	110, 115, 116
<i>Krachia tiara</i> (Monterosato, 1874)	102, 115
<i>Krachiopsis giannuzzii</i> Smriglio & Mariottini, 1999	110, 115, 116
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<i>Cirsotrema pumiceum</i> (Brocchi, 1814)	31†, 97†, 110, 167†
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<i>Epitonium clathrus</i> (Linnaeus, 1758)	31†, 97, 110, 167
<i>Epitonium jolyi</i> (Monterosato, 1878)	97i, 108, 110, 116
<i>Epitonium muricatum</i> (Risso, 1826)	31i, 97†, 110
<i>Epitonium pseudonanum</i> Bouchet & Warén, 1986	97, 110
<i>Epitonium pulchellum</i> (Bivona, 1832)	97, 110, 167†‡
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<i>Epitonium tryoni</i> (de Boury, 1913)	102, 110, 167†‡
<i>Epitonium turtonis</i> (W. Turton, 1819)	31†, 97, 110, 167†
<i>Gyroscala commutata</i> (Monterosato, 1877) ?	31†, 97†, 108†, 110†, 111†, 167†
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<i>Janthina globosa</i> Swainson, 1822	97†, 110, 167†
<i>Janthina janthina</i> (Linnaeus, 1758)	31†, 97, 110, 167†
<i>Janthina pallida</i> W. Thompson, 1840	102, 110
<i>Narrimania concinna</i> (Sykes, 1925)	102, 110
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<i>Opalia coronata</i> (Philippi & Scacchi, 1840)	31†, 97†, 110, 167†‡
<i>Opalia crenata</i> (Linnaeus, 1758)	102, 110
<i>Opaliopsis atlantis</i> (Clench & R. D. Turner, 1952)	102, 110
<i>Punctiscala cerigottana</i> (Sturany, 1896)	5†, 110
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<i>Aclis ascaris</i> (W. Turton, 1819)	97, 110
<i>Aclis attenuans</i> Jeffreys, 1883	97, 110
<i>Aclis gulsonae</i> (W. Clark, 1850)	110
<i>Aclis minor</i> (Brown, 1827)	97, 110, 167†
<i>Acrochalix callosa</i> Bouchet & Warén, 1986 ?	102, 109, 110
<i>Campylorhaphion famelicum</i> (R. B. Watson, 1883)	97, 102, 110, 116
<i>Crinophtheiros comatulicula</i> (Graff, 1875)	110, 167†
<i>Curveulima beneitoi</i> Peñas & Rolán, 2006	109, 110
<i>Curveulima dautzenbergi</i> (Pallary, 1900)	102, 110
<i>Curveulima devians</i> (Monterosato, 1884)	110, 116, 167†
<i>Eulima bilineata</i> Alder, 1848	95i, 97i, 110
<i>Haliella stenostoma</i> (Jeffreys, 1858)	102, 110
<i>Melanella alba</i> (da Costa, 1778)	97, 110, 116
<i>Melanella boscii</i> (Payraudeau, 1826)	97, 110, 111, 116
<i>Melanella compactilis</i> (Locard, 1891)	108, 110, 116
<i>Melanella conoidea</i> (Kurtz & Stimpson, 1851)	108, 110
<i>Melanella frielei</i> (Jordan, 1895)	97i, 110, 116
<i>Melanella levantina</i> (Oliverio, Buzzurro & Villa, 1994)	110, 116
<i>Melanella lubrica</i> (Monterosato, 1890)	97, 110, 116
<i>Melanella monterosatoi</i> (Monterosato, 1890)	97i, 110, 116, 167†i
<i>Melanella petitiiana</i> (Brusina, 1869)	110, 116
<i>Melanella polita</i> (Linnaeus, 1758)	31†, 97, 108, 110, 111, 167†
<i>Melanella pseudoglobbra</i> (Dautzenberg & H. Fischer, 1912) ?	31†, 97†, 110, 167†
<i>Melanella spiridioni</i> (Dautzenberg & H. Fischer, 1896)	97i, 102, 110
<i>Melanella stalioid</i> (Brusina, 1869)	31i, 75*, 167†i
<i>Melanella translucens</i> (Monterosato, 1890)	110, 116
<i>Nanobalcis nana</i> (Monterosato, 1878)	102, 110
<i>Parvioris ibizenca</i> (F. Nordsieck, 1968)	97, 110, 116, 167†
<i>Pelseneeria minor</i> Koehler & Vaney, 1908	97, 110
<i>Pelseneeria styliifera</i> (W. Turton, 1825)	167
<i>Sabinella bonifaciae</i> (F. Nordsieck, 1974)	97†, 110, 167†i
<i>Sticteulima jeffreysiana</i> (Brusina, 1869)	97, 102, 110
<i>Sticteulima lentiginosa</i> (A. Adams, 1861)	47, 108, 110
<i>Vitreolina antiflexa</i> (Monterosato, 1884)	97, 110, 167†‡
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<i>Vitreolina incurva</i> (Bucquoy, Dautzenberg & Dollfus, 1883)	97, 110, 167†i
<i>Vitreolina perminima</i> (Jeffreys, 1883)	110, 116
<i>Vitreolina philippi</i> (de Rayneval & Ponzi, 1854)	6, 97, 110
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<i>Eatonina matildae</i> Rubio & Rodriguez Babio, 1996 ?	109, 110
<i>Eatonina ochroleuca</i> (Brusina, 1869)	97, 110, 167†‡
<i>Eatonina pumila</i> (Monterosato, 1884)	97, 110
<i>Tubbreva micrometrica</i> (Aradas & Benoit, 1876)	109, 110
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<i>Alvania aspera</i> (Philippi, 1844)	97, 110, 167†
<i>Alvania balearica</i> J. D. Oliver & Templado, 2009	109, 110
<i>Alvania beanii</i> (Hanley in Thorpe, 1844)	97, 110, 167†
<i>Alvania bozcaadensis</i> Tisselli & Giunchi, 2013	104, 110
<i>Alvania cancellata</i> (da Costa, 1778)	31†, 97, 110, 167†
<i>Alvania carinata</i> (da Costa, 1778)	97, 110, 167
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<i>Alvania cimicoides</i> (Forbes, 1844)	97, 110, 167†j
<i>Alvania clarae</i> Nofroni & Pizzini, 1991	110
<i>Alvania clathrella</i> L. Seguenza, 1903	109, 110
<i>Alvania colossophilus</i> Oberling, 1970	97, 110, 123, 167†
<i>Alvania dalmatica</i> Buzzurro & Prkić, 2007	109, 110
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<i>Alvania dictyophora</i> (Philippi, 1844)	110
<i>Alvania discors</i> (T. Brown, 1818)	31, 97, 110, 167†
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<i>Alvania dorbignyi</i> (Audouin, 1826)	97, 110, 167
<i>Alvania elisae</i> Margelli, 2001	104, 110
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<i>Alvania geryonia</i> (Nardo, 1847)	97, 110, 167†
<i>Alvania hallgassi</i> Amati & Oliverio, 1985	147
<i>Alvania hirta</i> (Monterosato, 1884)	97i, 109, 110
<i>Alvania hispidula</i> (Monterosato, 1884)	97, 110, 167†
<i>Alvania josefoi</i> Oliver & Templado, 2009	104, 110
<i>Alvania lactea</i> (Michaud, 1830)	31†, 97, 110, 167†
<i>Alvania lanciae</i> (Calcara, 1845)	97, 110
<i>Alvania lineata</i> Risso, 1826	97, 110, 167
<i>Alvania litoralis</i> (Nordsieck, 1972)	110
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<i>Alvania oetyliaca</i> Amati & Chiarelli, 2017	8
<i>Alvania pagodula</i> (Bucquoy, Dautzenberg & Dollfus, 1884)	97, 110, 167†
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<i>Alvania rudis</i> (Philippi, 1844)	97, 110, 167†
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<i>Benthonella tenella</i> (Jeffreys, 1869)	97, 110

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<i>Crisilla chiarellii</i> (Cecalupo & Quadri, 1995)	114, 184
<i>Crisilla maculata</i> (Monterosato, 1869)	97†, 110
<i>Crisilla marioni</i> (Fasulo & Gaglini, 1987)	110
<i>Crisilla ramosorum</i> J. D. Oliver, Templado & Kersting, 2012	42
<i>Crisilla semistriata</i> (Montagu, 1808)	97, 110, 167†
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<i>Obtusella intersecta</i> (S. Wood, 1857)	97, 110, 167††
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<i>Peringiella elegans</i> (Locard, 1891)	110, 167†
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<i>Pusillina lineolata</i> (Michaud, 1830)	97, 110
<i>Pusillina marginata</i> (Michaud, 1830)	97, 110, 167†
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<i>Rissoa frauenfeldiana</i> Brusina, 1866	97, 110
<i>Rissoa gemmula</i> P. Fischer in de Folin, 1869	104, 97, 110, 167†
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<i>Rissoa parva</i> (da Costa, 1778) ?	97, 110
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<i>Rissoa similis</i> Scacchi, 1836	31†, 97, 104, 110, 167†
<i>Rissoa splendida</i> Eichwald, 1830	31†, 97, 104, 110, 167
<i>Rissoa variabilis</i> (Megerle von Mühlfeld, 1824)	31†, 97, 110, 167
<i>Rissoa ventricosa</i> Desmarest, 1814	31, 97, 110, 167
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<i>Nodulus contortus</i> (Jeffreys, 1856)	97, 110

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<i>Paludinella sicana</i> (Brugnone, 1876)	97, 110
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<i>Caecum clarkii</i> Carpenter, 1859	97, 110, 167
<i>Caecum subannulatum</i> de Folin, 1870	97, 110, 167
<i>Caecum trachea</i> (Montagu, 1803)	97, 110, 167
<i>Parastrophia asturiana</i> de Folin, 1870	97, 110, 167†
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<i>Eupaludestrina stagnorum</i> (Gmelin, 1791)	97†, 110
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<i>Ceratia proxima</i> (Forbes & Hanley, 1850)	97, 110
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<i>Potamopyrgus antipodarum</i> (Gray, 1843)	110, 143‡
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<i>Tornus subcarinatus</i> (Montagu, 1803)	97, 110, 167†
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<i>Truncatella subcylindrica</i> (Linnaeus, 1767)	97, 110, 167
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<i>Petaloconchus glomeratus</i> (Linnaeus, 1758)	97, 110, 167†
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<i>Thylacodes inopertus</i> (Leuckart, 1830)	97†, 110
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<i>Eunaticina papilla</i> (Gmelin, 1791)	42, 144
<i>Euspira catena</i> (da Costa, 1778)	97, 110
<i>Euspira fusca</i> (Blainville, 1825)	110
<i>Euspira guilleminii</i> (Payraudeau, 1826)	31†, 97, 110, 156, 167†
<i>Euspira intricata</i> (Donovan, 1804)	31†, 97†, 110, 111, 156, 167†
<i>Euspira macilenta</i> (Philippi, 1844)	97, 110
<i>Euspira nitida</i> (Donovan, 1804)	97†, 110, 111, 167†
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<i>Charonia variegata</i> (Lamarck, 1816)	31†, 97†, 110, 167†
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<i>Semicassis granulata</i> (Born, 1778)	31†‡, 75†
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<i>Atlanta lesueurii</i> J. E. Gray, 1850	110
<i>Atlanta peronii</i> Lesueur, 1817	110, 167†
<i>Atlanta selvagensis</i> de Vera & Seapy, 2006	110, 116
<i>Protatlanta souleyeti</i> (E. A. Smith, 1888)	110, 116, 167†
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<i>Aspella anceps</i> (Lamarck, 1822)	102, 110
<i>Babelomurex benoiti</i> (Tiberi, 1855)	31†, 97†, 110, 167†
<i>Babelomurex cariniferus</i> (Sowerby, 1834)	31†, 44, 97, 110, 167†
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<i>Coralliophila meyendorffii</i> (Calcara, 1845)	31, 97, 110, 167
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<i>Hexaplex trunculus</i> (Linnaeus, 1758)	31†, 83, 97, 110, 167†
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<i>Murexsul aradasii</i> (Monterosato in Poirier, 1883)	97, 110
<i>Murexsul cevikeri</i> (Houart, 2000)	110
<i>Muricopsis cristata</i> (Brocchi, 1814)	31, 97, 110, 111, 167
<i>Nucella lapillus</i> (Linnaeus, 1758) ?	42, 97, 102, 110
<i>Ocenebra edwardsii</i> (Payraudeau, 1826)	31†, 97†, 110, 167†
<i>Ocenebra erinaceus</i> (Linnaeus, 1758)	31, 97†, 110, 167†
<i>Ocenebra helleri</i> (Brusina, 1865)	110, 116
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<i>Granulina marginata</i> (Bivona, 1832)	97, 110
<i>Granulina melitensis</i> Smriglio, Mariottini & Rufini, 1998	110, 116
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<i>Volvarina mitrella</i> (Risso, 1826)	31†, 97, 110, 167†
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<i>Gibberula jansseni</i> van Aartsen, Menkhorst & Gittenberger, 1984	110, 116
<i>Gibberula miliaria</i> (Linnaeus, 1758)	97, 110, 167i
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<i>Gibberula turgidula</i> (Locard & Caziot, 1900)	97, 110
MITRIDAE Swainson, 1829	
<i>Episcomitra cornicula</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
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<i>Isara cornea</i> (Lamarck, 1811)	61
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<i>Pusia ebenus</i> (Lamarck, 1811)	31†, 97†, 110, 167†
<i>Pusia granum</i> (Forbes, 1844)	31†, 97†, 110, 167†
<i>Pusia savignyi</i> (Payraudeau, 1826)	110, 167
<i>Pusia tricolor</i> (Gmelin, 1791)	31†, 97†, 110, 167
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<i>Chauvetia candidissima</i> (Philippi, 1836)	97, 110, 167
<i>Chauvetia lineolata</i> (Tiberi, 1868)	110, 116
<i>Chauvetia procerula</i> (Monterosato, 1889)	97, 110, 116
<i>Chauvetia turritellata</i> (Deshayes, 1835)	110, 116
<i>Chauvetia ventrosa</i> Nordsieck, 1976	110, 116, 167
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<i>Euthria cornea</i> (Linnaeus, 1758)	31†, 97, 110, 167†
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<i>Cumia reticulata</i> (Blainville, 1829)	31†, 97, 110, 167†
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<i>Nassarius stolatus</i> (Gmelin, 1791)	110
<i>Nassarius turulosus</i> (Risso, 1826)	110
<i>Tritia corniculum</i> (Olivi, 1792)	31†, 97†, 109, 110, 167†
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<i>Tritia elongata</i> (Bucquoy, Dautzenberg & Dollfus, 1882)	109, 110
<i>Tritia gibbosula</i> (Linnaeus, 1758)	31†, 97†, 110, 157, 167†
<i>Tritia grana</i> (Lamarck, 1822)	97†, 110
<i>Tritia incrassata</i> (Strøm, 1768)	31†, 97†, 110, 157, 167†
<i>Tritia lima</i> (Dillwyn, 1817)	31†, 110
<i>Tritia mutabilis</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Tritia neritea</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Tritia nitida</i> (Jeffreys, 1867)	97†, 110, 167†
<i>Tritia pellucida</i> (Risso, 1826)	97†, 110, 167†
<i>Tritia reticulata</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Tritia tavernai</i> T. Cossignani, 2018	23
<i>Tritia unifasciata</i> (Kiener, 1834)	110
<i>Tritia varicosa</i> (W. Turton, 1825)	97†, 110†, 167†
PISANIIDAE Gray, 1857	
<i>Aplus dorbignyi</i> (Payraudeau, 1826)	31†, 97†, 110, 167†
<i>Aplus gaillardoti</i> (Puton, 1856)	3, 117
<i>Aplus scaber</i> (Locard, 1891)	97†, 110
<i>Aplus scacchianus</i> (Philippi, 1844)	31†, 97†, 110, 167†
<i>Enginella leucozona</i> (Philippi, 1844)	31†, 97†, 110, 167†
PISANIIDAE Gray, 1857	
<i>Pisania striata</i> (Gmelin, 1791)	31, 97, 110, 167
COLUMBELLIDAE Swainson, 1840	
<i>Columbella rustica</i> (Linnaeus, 1758)	31, 97, 110, 167
<i>Mitrella canariensis</i> (d'Orbigny, 1840)	102, 110
<i>Mitrella coccinea</i> (Philippi, 1836)	97, 110
<i>Mitrella gervillii</i> (Payraudeau, 1826)	31†, 97, 110, 167†
<i>Mitrella minor</i> (Scacchi, 1836)	97, 110, 167†

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<i>Mitrella scripta</i> (Linnaeus, 1758)	31†, 97, 110, 167
<i>Mitrella svelta</i> Kobelt, 1889	97†, 110, 167†
FASCIOLARIIDAE Gray, 1853	
<i>Aptyxis syracusana</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
<i>Pseudofusus angeli</i> (Russo & Angelidis, 2016)	155†
<i>Pseudofusus eviae</i> (Buzzurro & Russo, 2007)	110†
<i>Pseudofusus margaritae</i> (Buzzurro & Russo, 2007)	110†, 116†
<i>Pseudofusus patriciae</i> (Russo & Olivieri, 2013)	110†
<i>Pseudofusus profetai</i> (Nofroni, 1982)	110†, 118†
<i>Pseudofusus rolani</i> (Buzzurro & Ovalis, 2005)	97†, 110†
<i>Pseudofusus alternatus</i> (Buzzurro & Russo, 2007)	97†, 110†, 116†, 167†
<i>Pseudofusus dimitrii</i> (Buzzurro & Ovalis in Buzzurro & Russo, 2007)	110†
<i>Pseudofusus parvulus</i> Monterosato, 1884	110†, 116†
<i>Pseudofusus pulchellus</i> (Philippi, 1849)	97†, 110†, 167†
<i>Pseudofusus rusticulus</i> (Monterosato, 1880)	108†, 110†, 167†
<i>Pseudofusus rostratus</i> (Olivi, 1792)	97†, 110†, 167†
<i>Tarantinaea lignaria</i> (Linnaeus, 1758)	31†, 97†, 110, 167†
CANCELLARIIDAE Forbes & Hanley, 1851	
<i>Bivetiella cancellata</i> (Linnaeus, 1767)	31†, 97, 110
BORSONIIDAE Bellardi, 1875	
<i>Drilliola emendata</i> (Monterosato, 1872)	75*, 97i,
<i>Drilliola loprestiana</i> (Calcara, 1841)	97, 110, 167†
CLATHURELLIDAE H. Adams & A. Adams, 1858	
<i>Comarmondia gracilis</i> (Montagu, 1803)	31, 97, 110, 167
CONIDAE Fleming, 1822	
<i>Conus ventricosus</i> Gmelin, 1791	31†, 97†, 110, 167†
DRILLIIDAE Olsson, 1964	
<i>Crassopleura maravignae</i> (Bivona Ant. in Bivona And., 1838)	31, 97, 110, 167
HORAICLAVIDAE Bouchet, Kantor, Sysoev & Puillandre, 2011	
<i>Haedropleura secalina</i> (Philippi, 1844)	110, 167†
<i>Haedropleura septangularis</i> (Montagu, 1803)	31†, 97, 110, 167†
MANGELIIDAE P. Fischer, 1883	
<i>Bela cycladensis</i> (Reeve, 1845)	97i, 110
<i>Bela decussata</i> (Locard, 1891)	110
<i>Bela formica</i> F. Nordsieck, 1977 uncertain > taxon inquirendum	140
<i>Bela fuscata</i> (Deshayes, 1835)	97i, 110, 167
<i>Bela menkhorsti</i> van Aartsen, 1988	97, 110, 167†
<i>Bela minoica</i> Bogi, Giannuzzi Savelli & Pusateri, 2021	16
<i>Bela nebula</i> (Montagu, 1803)	97i, 110, 167
<i>Bela nebula</i> (Montagu, 1803) (<i>sensu</i> Nordsieck, 1977)	110
<i>Bela nuperima</i> (Tiberi, 1855)	5†, 97†, 110, 167†
<i>Bela taprurenensis</i> (Pallary, 1904)	97, 110
<i>Bela zenetouae</i> (van Aartsen, 1988)	97i, 110
<i>Bela zonata</i> (Locard, 1891)	31†, 97, 110
<i>Benthomangelia macra</i> (R. B. Watson, 1881)	110
<i>Kurtziella serga</i> (Dall, 1881)	110
<i>Mangelia attenuata</i> (Montagu, 1803)	97, 110, 167
<i>Mangelia barashi</i> (van Aartsen & Fehr-de Wal, 1978)	97, 110, 167
<i>Mangelia brusinae</i> van Aartsen & Fehr-de Wal, 1978	110
<i>Mangelia callosa</i> (F. Nordsieck, 1977)	97, 110, 163, 167†
<i>Mangelia costata</i> (Pennant, 1777)	97i, 110, 167†
<i>Mangelia costulata</i> Risso, 1826	31†, 97, 110, 111, 167†
<i>Mangelia difficilis</i> (Locard & Caziot, 1900)	110*, 167†
<i>Mangelia jerbaensis</i> Della Bella & Spada, 1997	163
<i>Mangelia melitensis</i> Cachia & Mifsud, 2008	110
<i>Mangelia multilineolata</i> (Deshayes, 1835)	31†, 97i, 110, 163, 167†
<i>Mangelia paciniana</i> (Calcara, 1839)	31†, 97, 110, 111, 167†
<i>Mangelia payraudeauti</i> (Deshayes, 1835)	110, 167†

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<i>Mangelia pontica</i> Milaschewitsch, 1908	75*
<i>Mangelia scabrida</i> Monterosato, 1890	110, 167†
<i>Mangelia secreta</i> (van Aartsen & Fehr-de Wal, 1978)	110
<i>Mangelia sicula</i> Reeve, 1846	110, 167†
<i>Mangelia striolata</i> Risso, 1826	97†, 110, 163, 167†
<i>Mangelia taeniata</i> (Deshayes, 1835)	97, 110, 167†
<i>Mangelia tenuicostata</i> (Brugnone, 1862)	110
<i>Mangelia thapsiae</i> (Oberling, 1970)	75*†, 97†j
<i>Mangelia unifasciata</i> (Deshayes, 1835)	97, 110, 111, 167†
<i>Mangelia vauquelini</i> (Payraudeau, 1826)	31†, 97, 110, 167†
<i>Pyrgocythara stosiciana</i> (Brusina, 1869)	97†, 110†, 163, 167†
<i>Sorgenfreispira brachystoma</i> (Philippi, 1844)	97†, 110, 167†
MITROMORPHIDAE T. L. Casey, 1904	
<i>Mitromorpha columbellaria</i> (Scacchi, 1836)	97†j, 110
<i>Mitromorpha crenipicta</i> (Dautzenberg, 1889)	97, 110, 167†
<i>Mitromorpha karpauthensis</i> (F. Nordsieck, 1969)	97, 110, 118
<i>Mitromorpha olivoidea</i> (Cantraine, 1835)	31†, 97, 110, 167†
RAPHITOMIDAE Bellardi, 1875	
<i>Clathromangelia granum</i> (Philippi, 1844)	97, 103, 110, 167
<i>Clathromangelia loiselierii</i> Oberling, 1970	97, 103, 110
<i>Clathromangelia strigilata</i> Pallary, 1904	97, 103, 110
<i>Cyrrillia aequalis</i> (Jeffreys, 1867)	97†, 103, 110, 167†
<i>Cyrrillia ephesina</i> Pusateri, Giannuzzi Savelli & Stahlschmidt, 2017	103, 110, 141
<i>Cyrrillia linearis</i> (Montagu, 1803)	31†, 97†, 110, 111, 167†
<i>Gymnobela abyssorum</i> (Locard, 1897)	103, 110
<i>Leufroyia concinna</i> (Scacchi, 1836)	54, 97†, 103, 110, 167††
<i>Leufroyia erronea</i> Monterosato, 1884	54, 103, 110
<i>Leufroyia leufroyi</i> (Michaud, 1828)	31†, 54, 97†, 103, 110, 167†
<i>Leufroyia villaria</i> (Pusateri & Giannuzzi-Savelli, 2008)	54, 97†, 103, 110
<i>Pleurotomella demosia</i> (Dautzenberg & H. Fischer, 1896)	110
<i>Pleurotomella eurybrocha</i> (Dautzenberg & H. Fischer, 1896)	5†*, 110, 167†
<i>Pleurotomella gibbera</i> Bouchet & Warén, 1980	103, 110
<i>Raphitoma alleryana</i> (Sullioti, 1889)	75*
<i>Raphitoma alternans</i> (Monterosato, 1884)	75*, 97j
<i>Raphitoma andrehoarai</i> Pelorce & Horst, 2020	73, 103†
<i>Raphitoma atropurpurea</i> (Locard & Caziot, 1900)	31i, 103, 110, 167†
<i>Raphitoma bicolor</i> (Risso, 1826)	51, 103, 110, 167†
<i>Raphitoma bracteata</i> (Pallary, 1904)	75*
<i>Raphitoma brunneofasciata</i> Pusateri, Giannuzzi-Savelli & Oliverio, 2013	103, 110, 167††
<i>Raphitoma contigua</i> (Monterosato, 1884)	101, 103, 110
<i>Raphitoma densa</i> (Monterosato, 1884)	103, 110, 167
<i>Raphitoma digiulioi</i> Pusateri & Giannuzzi Savelli, 2017	103, 110
<i>Raphitoma echinata</i> (Brocchi, 1814)	31, 53, 97, 103, 110, 139, 167
<i>Raphitoma farolita</i> F. Nordsieck, 1977	110
<i>Raphitoma georgiae</i> Kontadakis, 2022	73
<i>Raphitoma griseomaculata</i> Pusateri & Giannuzzi-Savelli, 2018	110
<i>Raphitoma hispidella</i> Giannuzzi-Savelli & Pusateri, 2019	97†j, 103†, 110†, 114, 167†† as <i>R. cordieri</i>
<i>Raphitoma horrida</i> (Monterosato, 1884)	97, 103, 110, 154
<i>Raphitoma laviae</i> (Philippi, 1844)	103, 110, 167
<i>Raphitoma lineolata</i> (Bucquoy, Dautzenberg & Dollfus, 1883)	51, 52, 97, 101, 103, 110, 167†
<i>Raphitoma locardi</i> Pusateri, Giannuzzi-Savelli & Oliverio, 2013	101, 103, 110
<i>Raphitoma melitis</i> Kontadakis & Mbazios, 2018	103, 110, 142
<i>Raphitoma mirabilis</i> (Pallary, 1904)	75*
<i>Raphitoma papillosa</i> (Pallary, 1904)	97, 103, 110
<i>Raphitoma philberti</i> (Michaud, 1829)	97, 101, 103, 110, 167†
<i>Raphitoma pruinosa</i> (Pallary, 1906)	97, 103, 110
<i>Raphitoma pseudohystrix</i> (Sykes, 1906)	75*

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<i>Raphitoma pupoides</i> (Monterosato, 1884)	75*, 167†
<i>Raphitoma smriglioi</i> Pusateri & Giannuzzi-Savelli, 2013	101, 103, 110
<i>Raphitoma sophiae</i> Kontadakis & Polyzoulis, 2018	103, 110
<i>Raphitoma spadiana</i> Pusateri & Giannuzzi-Savelli, 2012	101, 103, 110
<i>Taranis moerchii</i> (Malm, 1861)	97, 103, 110, 167
<i>Teretia</i> cfr. <i>neocaledonica</i> Morassi & Bonfitto, 2015	103, 110
<i>Teretia teres</i> (Reeve, 1844)	97, 103, 110
ARCHITECTONICIDAE Gray, 1850	
<i>Discotectonica discus</i> (Philippi, 1844)	31†, 110
<i>Heliacus fallaciosus</i> (Tiberi, 1872)	31, 97, 110, 167†
<i>Heliacus jeffreysianus</i> (Tiberi, 1867)	102, 110, 120
<i>Heliacus implexus</i> (Mighels, 1845)	72
<i>Pseudotorinia architae</i> (O. G. Costa, 1841)	31†, 97, 110
<i>Solatisonax alleryi</i> (G. Seguenza, 1876)	97, 110
<i>Spirolaxis clenchi</i> Jaume & Borro, 1946	102, 110
MATHILDIDAE Dall, 1889	
<i>Mathilda barbadensis</i> Dall, 1889	110
<i>Mathilda cochlaeformis</i> Brugnone, 1873	97, 110
<i>Mathilda coronata</i> Monterosato, 1875	102, 110
<i>Mathilda gemmulata</i> Semper, 1865	97, 110
<i>Mathilda retusa</i> Brugnone, 1873	110, 116, 167
RISSELLIDAE Gray, 1850	
<i>Rissoella angeli</i> Manousis, 2021	109, 110, 116††, 179, 184
<i>Rissoella diaphana</i> (Alder, 1848)	110, 116
<i>Rissoella globularis</i> (Forbes & Hanley, 1853) ?	97, 109, 110, 116
<i>Rissoella inflata</i> (Monterosato, 1880)	97, 109, 110
<i>Rissoella opalina</i> (Jeffreys, 1848)	97, 109, 110, 116
OMALOGYRIDAE G. O. Sars, 1878	
<i>Ammonicera fischeriana</i> (Monterosato, 1869)	97, 110, 167†
<i>Ammonicera lignea</i> (Palazzi, 1988)	110, 116
<i>Ammonicera rota</i> (Forbes & Hanley, 1850)	97, 110
<i>Omalogyra atomus</i> (Philippi, 1841)	97, 110, 167†
<i>Omalogyra disculus</i> Palazzi, 1988	97, 110, 116
<i>Retrotortina fuscata</i> Chaster, 1896	97, 110, 116
XYLODISCULIDAE Warén, 1992	
<i>Xylodiscula boucheti</i> Warén, Carrozza & Rocchini in Warén, 1992	108, 110
<i>Xylodiscula lens</i> Warén, 1992	108, 110
CORNIROSTRIDAE Ponder, 1980	
<i>Tomura depressa</i> (Granata-Grillo, 1877)	97, 98, 110, 111, 116
HYALOGYRINIDAE Warén & Bouchet, 1993	
<i>Hyalogyra zibrowii</i> Warén in Warén, Carrozza & Rocchini, 1997	109, 110, 151
<i>Hyalogyrina amphorae</i> Warén, Carrozza & Rocchini, 1997	110
<i>Xenoskenea pellucida</i> (Monterosato, 1874)	97, 110, 167††
PYRAMIDELLIDAE Gray, 1840	
<i>Auristomia barashi</i> (Bogi & Galil, 2000)	110
<i>Auristomia erjaveciana</i> (Brusina, 1869)	105, 110
<i>Auristomia fusulus</i> (Monterosato, 1878)	108, 110, 118
<i>Auristomia ignorata</i> Monterosato, 1917	105, 110
<i>Brachystomia angusta</i> (Jeffreys, 1867)	97†i, 105, 110
<i>Brachystomia carrozzai</i> (van Aartsen, 1987)	105, 110
<i>Brachystomia eulimoides</i> (Hanley, 1844)	97†, 105, 110
<i>Brachystomia scalaris</i> (MacGillivray, 1843)	105, 110, 167††
<i>Cingulina isseli</i> (Tryon, 1886)	29, 117
<i>Doliella nitens</i> (Jeffreys, 1870)	110, 167††
<i>Eulimella acicula</i> (Philippi, 1836)	97, 110, 167†
<i>Eulimella bogii</i> van Aartsen, 1994	105, 110
<i>Eulimella cerullii</i> (Cossmann, 1916)	110, 167†
<i>Eulimella cossignaniorum</i> van Aartsen, 1994	105, 110

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<i>Eulimella neoattenuata</i> Gaglini, 1992	105, 110
<i>Eulimella oliveri</i> Peñas & Rolán, 2006	105, 110
<i>Eulimella scillae</i> (Scacchi, 1835)	110
<i>Eulimella ventricosa</i> (Forbes, 1844)	97, 110, 167†
<i>Euparthenia bulinea</i> (R. T. Lowe, 1841)	97, 110, 167††
<i>Euparthenia humboldti</i> (Risso, 1826)	97, 110,
<i>Folinella excavata</i> (Phillippi, 1836)	97†, 110, 167†
<i>Folinella ghisottii</i> van Aartsen, 1984	110
<i>Kejdonia cachiai</i> (Mifsud, 1998)	105, 110
<i>Liostomia afzelii</i> Warén, 1991	105, 110
<i>Liostomia clavula</i> (Lovén, 1846)	110
<i>Liostomia mamoi</i> Mifsud, 1993	105, 110
<i>Megastomia conoidea</i> (Brocchi, 1814)	97†, 110, 167††, 170!†
<i>Megastomia conspicua</i> (Alder, 1850)	97†j, 110, 167††
<i>Megastomia crovatoi</i> Nofroni, Renda, Vannozzi, 2022	110j, 129
<i>Noemiamea dolioliformis</i> (Jeffreys, 1848)	105, 110
<i>Notodiaphana atlantica</i> Ortea, Moro & Espinosa, 2013	150*†
<i>Odostomella bicincta</i> (Tiberi, 1868)	105, 110
<i>Odostomella doliolum</i> (Philippi, 1844)	97, 110, 167††
<i>Odostomia aartseni</i> Nofroni, 1988	105, 110
<i>Odostomia acuta</i> Jeffreys, 1848	97, 110, 167†
<i>Odostomia dijkhuizeni</i> van Aartsen, Gittenberger & Goud, 1998	114
<i>Odostomia eremita</i> Peñas & Rolán, 1999	105, 110
<i>Odostomia</i> aff. <i>extenuata</i> Peñas & Rolán, 1999	42
<i>Odostomia improbabilis</i> Oberling, 1970	105, 110
<i>Odostomia kromi</i> van Aartsen, Menkhorst & Gittenberger, 1984	105, 110
<i>Odostomia lorellae</i> Micali, 1987	105, 110
<i>Odostomia lorioli</i> (Hornung & Mermoud, 1924)	110j = <i>Megastomia crovatoi</i>
<i>Odostomia lukisi</i> Jeffreys, 1859	97, 105, 110, 167†
<i>Odostomia megerlei</i> (Locard, 1886)	97, 105, 110, 118
<i>Odostomia nardoi</i> Brusina, 1869	110
<i>Odostomia plicata</i> (Montagu, 1803)	97, 110, 167†
<i>Odostomia rissoiformis</i> Milaschewitsch, 1909	108, 110
<i>Odostomia schrami</i> van Aartsen, E. Gittenberger & Goud, 1998	105, 110
<i>Odostomia sorianoii</i> Peñas & Rolán, 2006	105, 110
<i>Odostomia striolata</i> Forbes & Hanley, 1850	97, 110
<i>Odostomia suboblonga</i> Jeffreys, 1884	105
<i>Odostomia turriculata</i> Monterosato, 1869	97, 110, 167†
<i>Odostomia turrita</i> Hanley, 1844	97, 110, 167†
<i>Odostomia unidentata</i> (Montagu, 1803)	97, 110, 167††
<i>Odostomia widmeri</i> Tisselli & Manousis, 2022	50, 55, 112, 114, 146!, 184
<i>Ondina anceps</i> Gaglini, 1992	105, 110
<i>Ondina crystallina</i> Locard, 1891	105, 110
<i>Ondina diaphana</i> (Jeffreys, 1848)	105, 110
<i>Ondina dilucida</i> (Monterosato, 1884)	97, 105, 110††
<i>Ondina divisa</i> (J. Adams, 1797)	97i, 105, 108, 110
<i>Ondina modiola</i> (Monterosato, 1884)	105, 110
<i>Ondina neocrystallina</i> Gaglini, 1992	105, 110
<i>Ondina vitrea</i> (Brusina, 1866)	97, 110, 167††
<i>Ondina warreni</i> (Thompson, 1845)	97†, 110, 111, 167††
<i>Oscilla galilae</i> Bogi, Karhan & Yokes, 2012	118
<i>Parthenina alesii</i> Micali, Nofroni & Perna, 2012	105, 110
<i>Parthenina clathrata</i> (Jeffreys, 1848)	97†, 105, 110, 118, 167††
<i>Parthenina dantarti</i> (Peñas & Rolán, 2008)	105, 110
<i>Parthenina decussata</i> (Montagu, 1803)	110, 167††
<i>Parthenina dollfusi</i> (Kobelt, 1903)	110
<i>Parthenina emaciata</i> (Brusina, 1866)	97†, 110, 167††
<i>Parthenina eximia</i> (Jeffreys, 1849)	105, 110

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<i>Parthenina indistincta</i> (Montagu, 1808)	97†, 110, 167†‡
<i>Parthenina interstincta</i> (J. Adams, 1797)	97†, 110, 167†‡
<i>Parthenina juliae</i> (de Folin, 1872)	97†, 110, 167†‡
<i>Parthenina monozona</i> (Brusina, 1869)	110, 167†‡
<i>Parthenina monterosatii</i> (Clessin, 1900)	110, 167†‡
<i>Parthenina multicostata</i> (Jeffreys, 1884)	105, 110
<i>Parthenina palazzii</i> (Micali, 1984)	105, 110
<i>Parthenina penchynati</i> (Bucquoy, Dautzenberg & Dollfus, 1883)	105, 110
<i>Parthenina suturalis</i> (Philippi, 1844)	97†, 110, 167†‡
<i>Parthenina terebellum</i> (Philippi, 1844)	97†, 110, 167†‡
<i>Pyrgiscus crenatus</i> (Brown, 1827)	97†, 105, 110
<i>Pyrgiscus jeffreysii</i> (Jeffreys, 1848)	97†, 110, 167†
<i>Pyrgiscus rufus</i> (Philippi, 1836)	97, 110, 167†, 170!†
<i>Pyrgostylus striatulus</i> (Linnaeus, 1758)	97†, 110, 167†‡
<i>Pyrgulina pupaeformis</i> (Souverbie, 1865)	105, 110
<i>Pyrgulina stefanisi</i> (Jeffreys, 1869)	105, 110
<i>Spiralinella incerta</i> (Milaschewich, 1916)	97†, 110, 167†‡
<i>Spiralinella spiralis</i> (Montagu, 1803)	105, 110
<i>Syrnola fasciata</i> Jickeli, 1882	39, 42, 70, 110
<i>Syrnola minuta</i> H. Adams, 1869	105, 110
<i>Tiberia minuscula</i> (Monterosato, 1880)	110
<i>Tibersyrnola unifasciata</i> (Forbes, 1844)	97†, 105, 110, 117, 167†‡
<i>Trabecula jeffreysiana</i> Monterosato, 1884	97†, 105, 110
<i>Tragula fenestrata</i> (Jeffreys, 1848)	97†, 110, 167†‡
<i>Turbonilla acuta</i> (Donovan, 1804)	97, 110, 167†
<i>Turbonilla acutissima</i> Monterosato, 1884	97, 105, 110
<i>Turbonilla cangeyrani</i> Ovalis & Mifsud, 2017	118
<i>Turbonilla edgarii</i> (Melvill, 1896)	108, 110
<i>Turbonilla gradata</i> Bucquoy, Dautzenberg & Dollfus, 1883	97i, 110, 167‡, 170!
<i>Turbonilla lactea</i> (Linnaeus, 1758)	75*, 97i, 167‡
<i>Turbonilla micans</i> (Monterosato, 1875)	105, 110
<i>Turbonilla obliquata</i> (Philippi, 1844)	97, 110, 111, 167‡
<i>Turbonilla pumila</i> G. Seguenza, 1876	97i, 110, 167†‡
<i>Turbonilla pusilla</i> (Philippi, 1844)	97i, 110, 167‡
<i>Turbonilla sinuosa</i> (Jeffreys, 1884)	97, 110, 167‡
<i>Turbonilla subulina</i> Monterosato, 1889	97, 105, 110
AMATHINIDAE Ponder, 1987	
<i>Carinorbis clathrata</i> (Philippi, 1844)	110†
CIMIDAE Warén, 1993	
<i>Cima apicisbelli</i> Rolán, 2003	105, 110
<i>Cima cylindrica</i> (Jeffreys, 1856)	97, 105, 110, 111
<i>Cima minima</i> (Jeffreys, 1858)	97, 110
<i>Graphis albida</i> (Kanmacher, 1798)	97, 105, 110, 111
<i>Graphis barashi</i> van Aartsen, 2002	97, 98, 110, 111
<i>Graphis gracilis</i> (Monterosato, 1874)	146!
MURCHISONELLIDAE Casey, 1904	
<i>Ebala</i> aff. <i>communis</i> Bandel, 2005	42
<i>Ebala gradata</i> (Monterosato, 1878)	97, 98, 110
<i>Ebala nitidissima</i> (Montagu, 1803)	97, 110, 167‡
<i>Ebala pointeli</i> (de Folin, 1868)	97, 110, 111, 167‡
<i>Ebala striatula</i> (Jeffreys, 1856)	97, 98, 110
<i>Ebala trigonostoma</i> (de Folin, 1872)	105, 110, 118
<i>Murchisonella anabathron</i> (Hedley, 1906)	105, 110
<i>Murchisonella mediterranea</i> Peñas & Rolán, 2013	97†, 98, 105, 110, 111
TJAERNOEIIDAE Warén, 1991	
<i>Tjaernoelia exquisita</i> (Jeffreys, 1883)	75*
ACTEONIDAE d'Orbigny, 1843	

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<i>Acteon monterosatoi</i> Dautzenberg, 1889	110
<i>Acteon tornatilis</i> (Linnaeus, 1758)	97, 110, 167, 170!
<i>Callostracon tyrrhenicum</i> (Smriglio & Mariottini, 1996)	108, 110
<i>Crenilabium exile</i> (Jeffreys, 1870)	110
<i>Rapturella globulina</i> (Forbes, 1844)	110
PLAKOBRANCHIDAE Gray, 1840	
<i>Elysia flava</i> Verrill, 1901	12*
<i>Elysia hetta</i> Perrone, 1990	66
<i>Elysia margaritae</i> Fez, 1962	66, 137†‡
<i>Elysia nealae</i> Ostergaard, 1955	107, 110
<i>Elysia timida</i> (Risso, 1818)	20, 46, 61, 66
<i>Elysia viridis</i> (Montagu, 1804)	97, 110, 111, 169†
<i>Thuridilla hopei</i> (Vérany, 1853)	66, 110, 111
DIAPHANIDAE Odhner, 1914 (1857)	
<i>Diaphana cretica</i> (Forbes, 1844)	97i, 110
<i>Diaphana minuta</i> T. Brown, 1827	97, 110
MNESTIIDAE Oskars, Bouchet & Malaquias, 2015	
<i>Mnestia girardi</i> (Audouin, 1826)	110, 194†
NOTODIAPHANIDAE Thiele, 1931, taxon inquirendum	
<i>Notodiaphana atlantica</i> Ortea, Moro & Espinosa, 2013	150
RETUSIDAE Thiele, 1925	
<i>Pyrunculus fourierii</i> (Audouin, 1826)	12, 97i, 110; 150
<i>Pyrunculus hoernesii</i> (Weinkauff, 1866)	97, 110
<i>Pyrunculus ovatus</i> (Jeffreys, 1871)	107, 110
<i>Retusa candidula</i> (Locard, 1891)	97†, 110
<i>Retusa crebrisculpta</i> (Monterosato, 1884)	97†, 107, 110, 111
<i>Retusa crosseii</i> (Bucquoy, Dautzenberg & Dollfus, 1886)	97†i, 107, 110, 167†‡
<i>Retusa desgenettii</i> (Audouin, 1826)	107, 110
<i>Retusa laevisculpta</i> (Granata-Grillo, 1877)	25, 97†, 107, 110
<i>Retusa leptoneilema</i> (Brusina, 1866)	2, 97, 110, 111
<i>Retusa mammillata</i> (Philippi, 1836)	97, 110, 167†
<i>Retusa minutissima</i> (Monterosato, 1878)	2, 26, 97, 107, 110
<i>Retusa nitidula</i> (Lovén, 1846)	25, 107, 110
<i>Retusa obtusa</i> (Montagu, 1803)	97i, 110, 167‡
<i>Retusa parvula</i> (Jeffreys, 1883)	25, 97†, 107, 110
<i>Retusa tornata</i> (R. B. Watson, 1886)	97, 107, 110
<i>Retusa truncatula</i> (Bruguière, 1792)	97, 107, 110, 167†, 170!
<i>Retusa umbilicata</i> (Montagu, 1803)	97†, 110
<i>Retusa variabilis</i> (Milaschewitsch, 1912)	107, 110
RHIZORIDAE Dell, 1952	
<i>Volvulella acuminata</i> (Bruguière, 1792)	97, 110, 167†‡, 170!†
TORNATINIDAE P. Fischer, 1883	
<i>Acteocina mucronata</i> (Philippi, 1849)	110
RINGICULIDAE Philippi, 1853	
<i>Ringicula auriculata</i> (Ménard de la Groye, 1811)	97, 110, 167
<i>Ringicula conformis</i> Monterosato, 1877	97, 110, 167, 170!
<i>Ringicula gianninii</i> F. Nordsieck, 1974	97, 110
<i>Ringicula minuta</i> H. Adams, 1872	121, 196††
RUNCINIDAE H. Adams & A. Adams, 1854	
<i>Runcina adriatica</i> T. E. Thompson, 1980	107, 109, 110
BULLIDAE Gray, 1827	
<i>Bulla ampulla</i> Linnaeus, 1758	97‡
<i>Bulla arabica</i> Malaquias & D. Reid, 2008	110, 117, 178i as <i>B. ampulla</i>
<i>Bulla striata</i> Bruguière, 1792	31, 97, 110, 167, 170!
HAMINOEIDAE Pilsbry, 1895	
<i>Aliculastrum cylindricum</i> (Helbling, 1779)	107, 110
<i>Atys angustatus</i> E. A. Smith, 1872	2, 03
<i>Atys ehrenbergi</i> (Issel, 1869)	2

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<i>Haloa japonica</i> (Pilsbry, 1895)	107, 109, 110
<i>Haminoea hydatis</i> (Linnaeus, 1758)	97, 110, 111, 167
<i>Haminoea navicula</i> (da Costa, 1778)	31, 97, 110, 167
<i>Haminoea orbignyana</i> (Férussac, 1822)	97, 107, 110
<i>Haminoea ortei</i> Talavera, Murillo & Templado, 1987	43, 107, 110
<i>Lamprohaminoea ovalis</i> (Pease, 1868)	66†, 97†, 110†, 122†, 153†, 186†
<i>Liloea mongii</i> (Audouin, 1826)	134†, 150
<i>Roxaniella jeffreysi</i> (Weinkauff, 1866)	97†, 110, 111, 167i, 170!†
<i>Weinkauffia macandrewi</i> (E. A. Smith, 1872)	2, 70†, 97†i, 107, 110
<i>Weinkauffia turgidula</i> (Forbes, 1844)	2, 97, 110, 111, 167†, 170!†
LAONIDAE Pruvot-Fol, 1954	
<i>Laona pruinosa</i> (W. Clark, 1827)	107, 110
<i>Laona quadrata</i> (S. Wood, 1839)	97†i, 107, 110
PHILINIDAE Gray, 1850	
<i>Hermania scabra</i> (O. F. Müller, 1784)	110, 170!†
<i>Philine angasi</i> (Crosse, 1865)	107, 110
<i>Philine angulata</i> Jeffreys, 1867	107, 110
<i>Philine catena</i> (Montagu, 1803)	97, 98, 110, 111, 167‡
<i>Philine denticulata</i> (J. Adams, 1800)	97, 98, 107, 110, 111
<i>Philine intricata</i> Monterosato, 1884	97, 98, 110
<i>Philine iris</i> Tringali, 2001	97, 98, 110
<i>Philine monterosati</i> Monterosato, 1874	107, 110
<i>Philine powelli</i> Rudman, 1970	107, 110
<i>Philine punctata</i> (J. Adams, 1800)	97, 98, 110
<i>Philine quadripartita</i> Ascanius, 1772	31†, 97†, 110, 111, 167, 170!†
<i>Philine vestita</i> (Philippi, 1840)	110
SCAPHANDRIDAE G. O. Sars, 1878	
<i>Scaphander lignarius</i> (Linnaeus, 1758)	31, 97, 107, 110, 167
<i>Scaphander punctostriatus</i> (Mighels & C. B. Adams, 1842)	107, 110
AGLAJIDAE Pilsbry, 1895	
<i>Aglaja tricolorata</i> Renier, 1807	97i, 170!
<i>Biuve fulvipunctata</i> (Baba, 1938)	107, 110
<i>Camachoaglaja africana</i> (Pruvot-Fol, 1953)	107, 110, 111
<i>Melanochlamys algirae</i> (A. Adams, 1850)	109, 110
<i>Melanochlamys wildpretii</i> Ortea, Bacallado & Moro, 2003	97, 98, 107, 110
<i>Philinopsis depicta</i> (Renier, 1807)	66
<i>Tubulophilinopsis</i> sp.	42
ALACUPPIDAE Oskars, Bouchet & Malaquias, 2015	
<i>Roxania monterosatoi</i> Dautzenberg & H. Fischer, 1896	110
<i>Roxania utriculus</i> (Brocchi, 1814)	97, 110, 167
COLPODASPIDIDAE Oskars, Bouchet & Malaquias, 2015	
<i>Colpodaspis pusilla</i> M. Sars, 1870	107, 110
CYLICHNIDAE H. Adams & A. Adams, 1854	
<i>Cylichna alba</i> (T. Brown, 1827)	97, 110, 167 ‡
<i>Cylichna cylindracea</i> (Pennant, 1777)	97, 110, 167, 170!
GASTROPTERIDAE Swainson, 1840	
<i>Gastropteron rubrum</i> (Rafinesque, 1814)	75*
CAVOLINIIDAE Gray, 1850 (1815)	
<i>Cavolinia gibbosa</i> (d'Orbigny, 1835)	97, 110
<i>Cavolinia inflexa</i> (Lesueur, 1813)	97, 110, 167
<i>Cavolinia tridentata</i> (Forsskal [in Niebuhr], 1775)	31, 97, 110, 167
<i>Diacria trispinosa</i> (Lesueur, 1821)	97, 110
CLIIDAE Jeffreys, 1869	
<i>Clio cuspidata</i> (Bosc, 1801)	97, 110
<i>Clio pyramidata</i> Linnaeus, 1767	97, 110, 167
CRESEIDAE Rampal, 1973	
<i>Creseis clava</i> (Rang, 1828)	97†, 110, 167‡‡

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<i>Creseis virgula</i> (Rang, 1828)	97, 110
<i>Styliola subula</i> (Quoy & Gaimard, 1827)	97, 110, 167†
HYALOCYLIDAE A. W. Janssen, 2020	
<i>Hyalocylis striata</i> (Rang, 1828)	110
LIMACINIDAE Gray, 1840	
<i>Heliconoides inflatus</i> (d'Orbigny, 1835)	97†, 110, 167††
<i>Limacina bulimoides</i> (d'Orbigny, 1835)	97, 110
<i>Limacina retroversa</i> (J. Fleming, 1823)	110
<i>Limacina trochiformis</i> (d'Orbigny, 1835)	97, 110
<i>Thielea helicoides</i> (Jeffreys, 1877)	110
CYMBULIIDAE Gray, 1840	
<i>Cymbulia peronii</i> Blainville, 1818	107, 110
<i>Gleba cordata</i> Forsskål in Niebuhr, 1776	110
PERACLIDAE Tesch, 1913	
<i>Peracle reticulata</i> (d'Orbigny, 1835)	110
<i>Peracle elata</i> (Seguenza, 1875)	75*
OXYNOIDAE Stoliczka, 1885	
<i>Lobiger serradifalci</i> (Calcara, 1840)	97, 110, 111, 167
<i>Oxynoe olivacea</i> Rafinesque, 1814	97, 110, 167
VOLVATELLIDAE Pilsbry, 1895	
<i>Ascobulla fragilis</i> (Jeffreys, 1856)	97, 110, 111, 167, 170!†
HERMAEIDAE H. Adams & A. Adams, 1854	
<i>Aplysiopsis elegans</i> Deshayes, 1853	171*
<i>Cyerce graeca</i> T. E. Thompson, 1988	171*
LIMAPONTIIDAE Gray, 1847	
<i>Calliopaea bellula</i> d'Orbigny, 1837	97, 98, 110, 111
<i>Limapontia capitata</i> (O. F. Müller, 1774)	107, 110
<i>Placida viridis</i> (Trinchese, 1874)	12*
UMBRACULIDAE Dall, 1889	
<i>Umbraculum umbraculum</i> (Lightfoot, 1786)	31†, 66, 97, 110, 167†
TYLODINIDAE Gray, 1847	
<i>Tylodina perversa</i> (Gmelin, 1791)	25, 66, 97, 110, 167
PLEUROBRANCHIDAE Gray, 1827	
<i>Berthella aurantiaca</i> (Risso, 1818)	66, 97, 110
<i>Berthella ocellata</i> (Delle Chiaje, 1830)	25, 66
<i>Berthella plumula</i> (Montagu, 1803)	97, 98, 110, 167
<i>Berthella stellata</i> (Risso, 1826)	110, 111
<i>Berthellina edwardsii</i> (Vayssi�re, 1897)	66, 107, 110
<i>Pleurobranchus forskalii</i> R�ppell & Leuckart, 1828	110
<i>Pleurobranchus membranaceus</i> (Montagu, 1816)	110, 111
<i>Pleurobranchus testudinarius</i> Cantraine, 1835	66, 135
HEDYLOPSIDAE Odhner, 1952	
<i>Hedylopsis spiculifera</i> (Kowalevsky, 1901)	107, 110
AKERIDAE Mazzarelli, 1891	
<i>Akera bullata</i> O. F. M�ller, 1776	97, 110
APLYSIIDAE Lamarck, 1809	
<i>Aplysia dactylomela</i> Rang, 1828	62, 66, 110, 127
<i>Aplysia depilans</i> Gmelin, 1791	97, 110
<i>Aplysia fasciata</i> Poiret, 1789	25, 31, 66, 97, 110, 167
<i>Aplysia parvula</i> M�rch, 1863	21, 25, 97, 110
<i>Aplysia punctata</i> (Cuvier, 1803)	66, 97, 110
<i>Bursatella leachii</i> Blainville, 1817	30, 66, 97, 98, 110
<i>Notarchus punctatus</i> Philippi, 1836	66, 110, 135
<i>Petalifera gravieri</i> (Vayssi�re, 1906)	97, 98, 110
<i>Petalifera petalifera</i> (Rang, 1828)	25, 66, 97, 110, 111, 170!
<i>Phyllaplysia lafonti</i> (P. Fischer, 1870)	110
<i>Syphonota geographica</i> (A. Adams & Reeve, 1850)	66, 70, 122*
ARMINIDAE Iredale & O'Donoghue, 1923 (1841)	

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<i>Armina neapolitana</i> (Delle Chiaje, 1824)	79*
CORYPHELLIDAE Bergh, 1889	
<i>Fjordia lineata</i> (Lovén, 1846)	66
EUBRANCHIDAE Odhner, 1934	
<i>Amphorina farrani</i> (Alder & Hancock, 1844)	125†
<i>Amphorina linensis</i> (Garcia-Gomez, Cervera & Garcia, 1990)	117
FIONIDAE Gray, 1857	
<i>Fiona pinnata</i> (Eschscholtz, 1831)	25
DORIDIDAE Rafinesque, 1815	
<i>Doris bertheloti</i> (d'Orbigny, 1839)	66
<i>Doris marmorata</i> Risso, 1818	79*
<i>Doris ocelligera</i> (Bergh, 1881)	110
<i>Doris pseudoargus</i> Rapp, 1827	79*†, 97, 110, 111
CHROMODORIDIDAE Bergh, 1892	
<i>Felimida binza</i> (Ev. Marcus & Er. Marcus, 1963)	15†, 66, 110†
<i>Felimare orsinii</i> (Vérany, 1846)	66
<i>Felimare picta</i> (Schultz in Philippi, 1836)	66, 97†, 110
<i>Felimare villafranca</i> (Risso, 1818)	66, 97†, 98, 110, 111
<i>Felimida krohni</i> (Vérany, 1846)	110
<i>Felimida luteorosea</i> (Rapp, 1827)	25, 66, 79*†, 97†, 110
<i>Felimida purpurea</i> (Risso, 1831)	66
<i>Goniobranchus annulatus</i> (Eliot, 1904)	30†, 125, 172, 188†*
<i>Hypselodoris infucata</i> (Rüppell & Leuckart, 1830)	63, 193*
DISCODORIDIDAE Bergh, 1891	
<i>Discodoris rosi</i> Ortea, 1979	138
<i>Jorunna onubensis</i> Cervera, Garcia-Gomez & Garcia, 1986	107, 110
<i>Jorunna tomentosa</i> (Cuvier, 1804)	79†*
<i>Paradoris indecora</i> (Bergh, 1881)	66
<i>Peltodoris atromaculata</i> Bergh, 1880	66, 110
<i>Platydoris argo</i> (Linnaeus, 1767)	66, 67, 77
<i>Tayuva lilacina</i> (Gould, 1852)	25, 66
PHYLLIDIIDAE Rafinesque, 1814	
<i>Phyllidia flava</i> Aradas, 1847	66
CALYCIDORIDIDAE Roginskaya, 1972	
<i>Diaphorodoris papillata</i> Portmann & Sandmeier, 1960	66
DENDRODORIDIDAE O'Donoghue, 1924	
<i>Dendrodoris grandiflora</i> (Rapp, 1827)	66, 97†, 110, 111
<i>Dendrodoris herytra</i> Valdés & Ortea in Valdés, Ortea, Avila & Ballesteros, 1996	107, 110, 111
<i>Dendrodoris limbata</i> (Cuvier, 1804)	66, 110, 111
GONIODORIDIDAE H. Adams & A. Adams, 1854	
<i>Okenia elegans</i> (Leuckart, 1828)	79*
<i>Okenia picoensis</i> Paz-Sedano, Ortigosa & Pola, 2017	60
<i>Trapania lineata</i> Haefelfinger, 1960	66
<i>Trapania maculata</i> Haefelfinger, 1960	66
POLYCERIDAE Alder & Hancock, 1845	
<i>Plocamopherus ocellatus</i> Rüppell & Leuckart, 1828	66, 144
<i>Polycera quadrilineata</i> (O. F. Müller, 1776)	66
<i>Polycerella emertoni</i> A. E. Verrill, 1880†	64†, 82*, 133*
TRITONIIDAE Lamarck, 1809	
<i>Duvaucelia manicata</i> (Deshayes, 1853)	66, 110†, 111
HANCOCKIIDAE MacFarland, 1923	
<i>Hancockia uncinata</i> (Hesse, 1872)	25, 110
SCYLLAEIDAE Alder & Hancock, 1855	
<i>Notobryon bijecurum</i> Baba, 1937	107, 110, 111
<i>Scyllaea pelagica</i> Linnaeus, 1758	25, 66, 135
TETHYDIDAE Rafinesque, 1815	
<i>Melibe viridis</i> (Kelaart, 1858)	66, 80†, 97, 110, 181
JANOLIDAE Pruvot-Fol, 1933	

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<i>Antiopella cristata</i> (Delle Chiaje, 1841)	66, 110, 135+
AEOLIDIIDAE Gray, 1827	
<i>Anteaeolidiella lurana</i> (Ev. Marcus & Er. Marcus, 1967)	14
<i>Baeolidia moebii</i> Bergh, 1888	84
<i>Berghia coerulescens</i> (Laurillard, 1832)	15, 66
<i>Berghia verrucicornis</i> (A. Costa, 1867)	66, 97, 98, 110, 111
<i>Spurilla neapolitana</i> (Delle Chiaje, 1841)	25, 66, 107, 110
FACELINIDAE Bergh, 1889	
<i>Caloria elegans</i> (Alder & Hancock, 1845)	66, 135
<i>Caloria quatrefagesi</i> (Vayssière, 1888)	66
<i>Cratena peregrina</i> (Gmelin, 1791)	66, 79*, 97, 98, 110, 111
<i>Dicata odhneri</i> Schmekel, 1967	58
<i>Dondice banyulensis</i> Portmann & Sandmeier, 1960	66, 135
<i>Dondice trainitoi</i> Furfaro & Mariottini, 2020	48
<i>Facelina annulicornis</i> (Chamisso & Eysenhardt, 1821)	97, 110, 111
<i>Facelina rubrovittata</i> (Costa A., 1866)	66
PISEINOTECIDAE Edmunds, 1970	
<i>Paraflabellina gabinieri</i> (Vicente, 1975)	136+
FLABELLINIDAE Bergh, 1889	
<i>Coryphellina rubrolineata</i> O'Donoghue, 1929	38+, 66, 71+, 110
<i>Edmundsella pedata</i> (Montagu, 1816)	66, 97+, 110, 111
<i>Flabellina affinis</i> (Gmelin, 1791)	41*, 66, 110, 160*
<i>Flabellina cavolini</i> (Vérany, 1846)	66
<i>Luisella babai</i> (Schmekel, 1972)	66, 135+
<i>Paraflabellina ischitana</i> (Hirano & T. E. Thompson, 1990)	41*, 66
CUMANOTIDAE Odhner, 1907	
<i>Cumanotus beaumonti</i> (Eliot, 1906)	66
SIPHONARIIDAE Gray, 1827	
<i>Siphonaria pectinata</i> (Linnaeus, 1758)	31, 97, 110, 128, 167
<i>Williamia gussoni</i> (O. G. Costa, 1829)	97, 110, 167
TRIMUSCULIDAE J. Q. Burch, 1945 (1840)	
<i>Trimusculus mammillaris</i> (Linnaeus, 1758)	110, 167
TRINCHESIIDAE F. Nordsieck, 1972	
<i>Trinchesia caerulea</i> (Montagu, 1804)	109, 110
ELLOBIIDAE L. Pfeiffer, 1854	
<i>Leucophytia bidentata</i> (Montagu, 1808)	97+, 110+
<i>Myosotella myosotis</i> (Draparnaud, 1801)	97, 110, 167+
<i>Ovatella aequalis</i> (R. T. Lowe, 1832)	108, 110
<i>Ovatella firminii</i> (Payraudeau, 1827)	97, 110, 132!, 167
POLYPLACOPHORA	
ACANTHOCHITONIDAE Pilsbry, 1893	
<i>Acanthochitona crinita</i> (Pennant, 1777)	97, 110
<i>Acanthochitona fascicularis</i> (Linnaeus, 1767)	31, 97, 110
CALLOCHITONIDAE Plate, 1901	
<i>Callochiton septemvalvis</i> (Montagu, 1803)	97, 110
CHITONIDAE Rafinesque, 1815	
<i>Rhyssoplax corallinus</i> (Risso, 1826)	31+, 110
<i>Rhyssoplax olivacea</i> Spengler, 1797	31+, 97+, 110
HANLEYIDAE Bergenhayn, 1955	
<i>Hanleya hanleyi</i> (Bean, 1844)	75*, 165
<i>Hanleya mediterranea</i> Sirenko, 2014	34*
ISCHNOCHITONIDAE Dall, 1889	
<i>Callistochiton pachylasmae</i> (Monterosato, 1879)	33, 97, 110
<i>Ischnochiton rissoi</i> (Payraudeau, 1826)	110
<i>Ischnochiton tsekosi</i> Koukouras & Karachle, 2005	74!, 75*
<i>Ischnochiton usticensis</i> Dell'Angelo & Castriota, 1999	118

LEPTOCHITONIDAE Dall, 1889	
<i>Lepidopleurus cajetanus</i> (Poli, 1791)	110
<i>Leptochiton algesirensis</i> (Capellini, 1859)	110
<i>Leptochiton bedullii</i> Dell'Angelo & Palazzi, 1986	56*
<i>Leptochiton boettgeri</i> Sulc, 1934	75*
<i>Leptochiton cancellatus</i> (G. B. Sowerby II, 1840)	110
<i>Leptochiton scabridus</i> (Jeffreys, 1880)	110
<i>Parachiton africanus</i> (Nierstrasz, 1906)	110
TONICELLIDAE Simroth, 1894	
<i>Lepidochitona caprearum</i> (Scacchi, 1836)	110
<i>Lepidochitona cinerea</i> (Linnaeus, 1767)	110
<i>Lepidochitona monterosatoi</i> Kaas & Van Belle, 1981	75*
SCAPHOPODA	
DENTALIIDAE Gray, 1847	
<i>Antalis agilis</i> (M. Sars in G. O. Sars, 1872)	97, 110
<i>Antalis dentalis</i> (Linnaeus, 1758)	97, 110
<i>Antalis entalis</i> (Linnaeus, 1758) ?	97, 110
<i>Antalis inaequicostata</i> (Dautzenberg, 1891)	31†, 97, 110, 167†
<i>Antalis panorma</i> (Chenu, 1843)	110
<i>Antalis rossati</i> (Caprotti, 1966)	97, 110
<i>Antalis vulgaris</i> (da Costa, 1778)	31†, 97, 110, 167†
FUSTIARIIDAE Steiner, 1991	
<i>Fustiaria rubescens</i> (Deshayes, 1825)	31, 97, 110, 167
GADILINIDAE Chistikov, 1975	
<i>Episiphon filum</i> (G. B. Sowerby II, 1860)	97, 110
ENTALINIDAE Chistikov, 1979	
<i>Entalina tetragona</i> (Brocchi, 1814)	97, 110
PULSELLIDAE Boss, 1982	
<i>Pulsellum lofotense</i> (M. Sars, 1865)	97, 110
GADILIDAE Stoliczka, 1868	
<i>Cadulus jeffreysi</i> (Monterosato, 1875)	97, 110
<i>Cadulus subfusiformis</i> (M. Sars, 1865)	108, 110
<i>Dischides politus</i> (S. Wood, 1842)	97, 110, 167†
SOLENOGASTRES	
RHOPALOMENIIDAE Salvini-Plawen, 1978	
<i>Rhopalomenia aglaopheniae</i> (Kowalevsky & Marion, 1887)	32*

TWO NEW *EMARGINULA* LAMARCK, 1801 SPECIES FOR THE HELLENIC SEAS

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Keywords

Emarginula, Greece, Mediterranean Sea, Mollusca, New Records

Abstract

Additional findings from the ongoing study on the marine malacofauna of the Hellenic Seas by June 2023. It incorporates 3 species out of which 2 are new records for the Hellenic malacofauna and 1 is a second image documented record for the Hellenic territory. Taxonomic characteristics and ecological data are presented and discussed.

Introduction

The recent faunistic works on the Hellenic marine malacofauna has been presented in Galinou-Mitsoudi *et al.* (2023) and the relevant biodiversity was last presented in an annotated catalogue by Manousis (2023). Attentive search based on material management and multifaceted research activity in the field continues to produce and present new records for the area.

Methods

Sampling, handling and species recognition were conducted according to Manousis & Galinou-Mitsoudi (2014) and Mbazios *et al.* (2020). The specimens are deposited in the premises of K. Papavasileiou. Scientists are welcome to have access to the biological material for inspection by appointment.

Abbreviations

CS - Coralligenous substrate
G1 - First record for Greece
G2 - Second record for Greece

Results and Discussion

The records

From the deep waters of North Aegean Sea two *Emarginula* species were identified as first records for the Hellenic seas namely *E. christiaensi* and *E. fissura*:

Emarginula christiaensi Piani, 1985 (Fig. 2b). 1 live individual associated with the coral *Desmophyllum pertusum* (Linnaeus, 1758) (Fig. 2c), CS, 350 m, Psathoura Isl., 39°35'N - 24°10'E, June 2023, G1 (description in: Gofas *et al.*, 2011).

Emarginula fissura (Linnaeus, 1758) (Fig. 1a, b). 2 shells associated also with *Desmophyllum pertusum* (Linnaeus, 1758) (Fig. 2c), 350 m, Psathoura Isl., 39°35'N - 24°10'E, June 2023, G1 (description in: Gofas *et al.*, 2011). Up to this work, 10 *Emarginula* species have been referred for the Hellenic seas (Manousis, 2021) and now two more species are added while the genus is represented by 17 species in the Mediterranean Sea (Piani, 1985; Manousis & Galinou-Mitsoudi, 2014). The majority of *Emarginula* species are referred as rare or uncommon, except for the common species *E. adriatica*, *E. huzardii*, *E. octaviana* and *E. tenera* (Repetto *et al.*, 2005). Besides the two *Emarginula* species, the bivalve *Parathyasira granulosa* (Monterosato, 1874) (Fig. 2a) was also recorded as 2 shells, from the same location of 350 m, Psathoura Isl., 39°35'N - 24°10'E, June 2023, G2 (G1 in: Manousis *et al.*, 2021, Fig. 2c.) (description in: Oliver & Killeen, 2002; Gofas *et al.*, 2011). The species is referred as rare (Repetto *et al.*, 2005) from deep waters of 100-1800 m (Oliver & Killeen, 2002).

Acknowledgements

The authors are grateful to the anonymous referees for their useful contribution to improve the publication.

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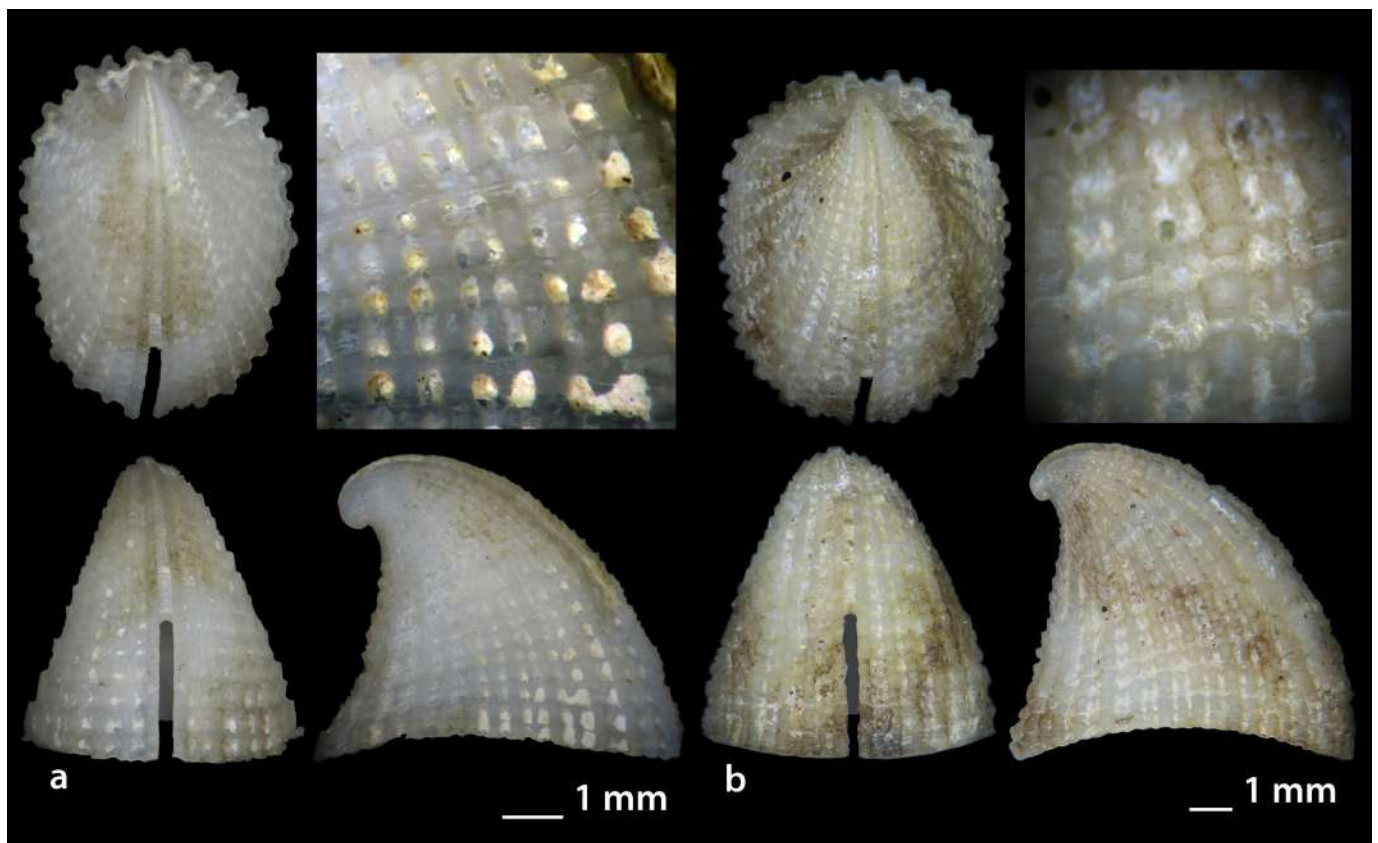
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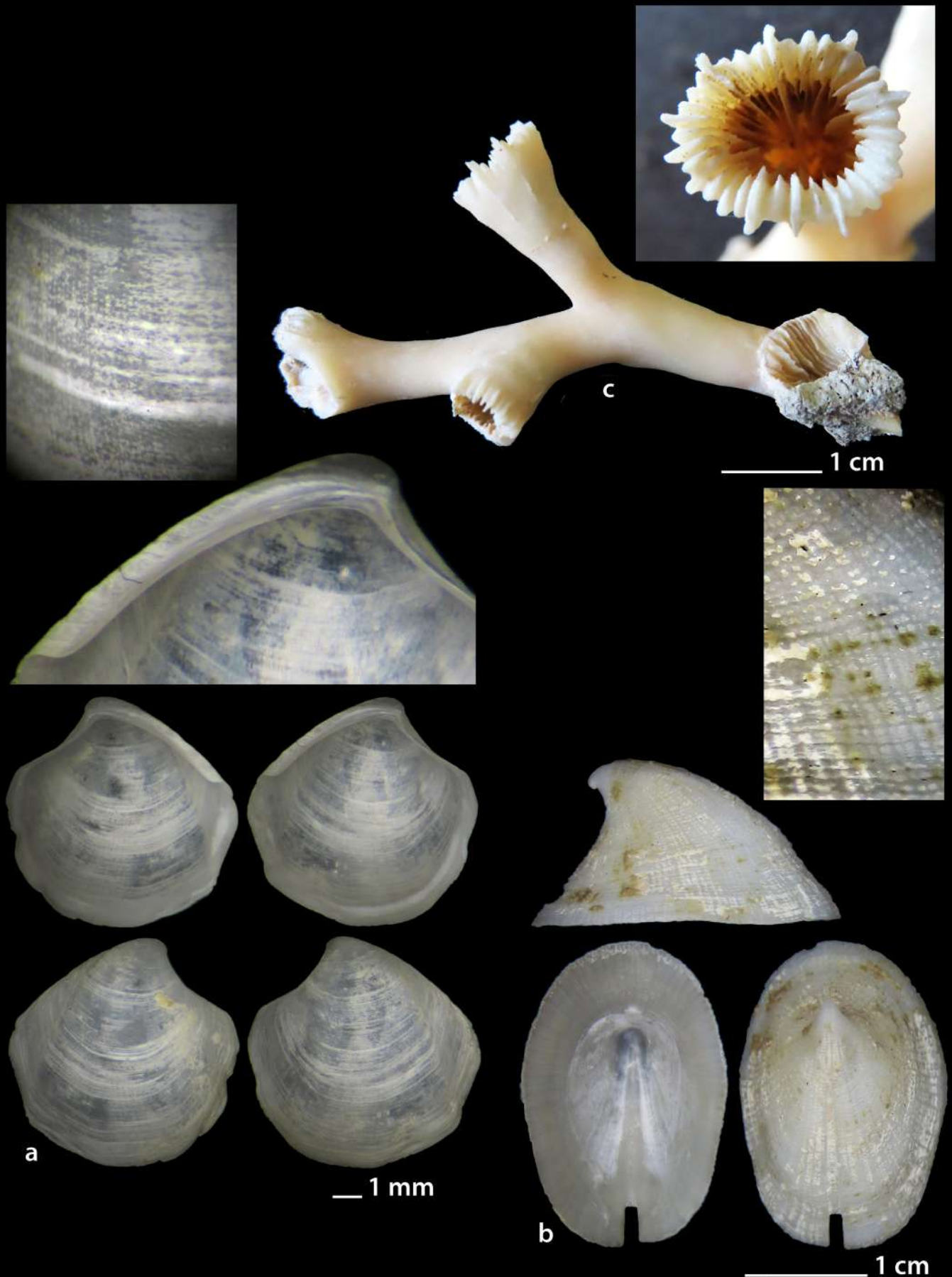
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Figure 1



a and b. *Emarginula fissura* (Linnaeus, 1758).

Figure 2



a. *Parathyasira granulosa* (Monterosato, 1874); b. *Emarginula christiaensi* Piani, 1985; c. *Desmophyllum pertusum* (Linnaeus, 1758) host.



Nassarulidae
Nassarulus vitiensis
MNHN La planète revisitée
Koumac 2018
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