

XENOPHORA

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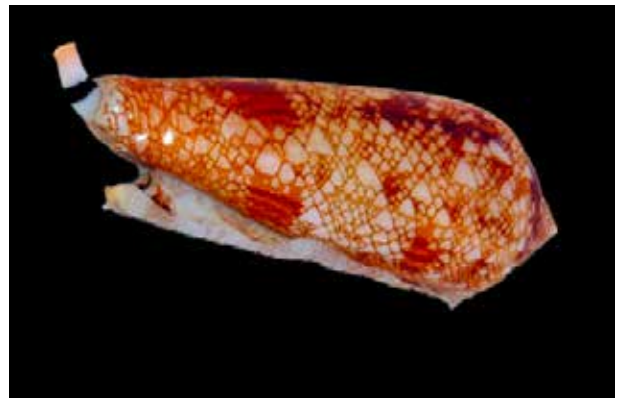
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Couverture:

Cylinder patmartii nov. sp.,
paratype 6
photographié vivant après sa collecte, en bac,
à bord.
Photographie : Patrick Marti.



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DESCRIPTION DE *CYLINDER PATMARTII* (GASTROPODA: CONIDAE), DE L'ARCHIPEL DES ÎLES MARQUISES (POLYNÉSIE FRANÇAISE)

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MOTS-CLÉS

Conidae, *Cylinder patmartii*, Marquises, Polynésie française, diagnose, endémisme.

KEYWORDS

Conidae, *Cylinder patmartii*, Marquesas Islands, French Polynesia, Diagnosis, Endemism.

RÉSUMÉ

Cet article décrit *Cylinder patmartii*, une nouvelle espèce de cône découverte dans l'archipel des îles Marquises en Polynésie française. Les auteurs examinent ses caractéristiques morphologiques distinctives et la comparent à trois autres espèces de cônes présentes en Polynésie française : *C. aureus* (Hwass, 1792), *C. canonicus* (Hwass, 1792), et *C. legatus* (Lamarck, 1810).

ABSTRACT

This article describes *Cylinder patmartii*, a new cone species discovered in the Marquesas Islands, French Polynesia. The authors examine its distinctive morphological features and compare it to three other cone species found in French Polynesia: *C. aureus* (Hwass, 1792), *C. canonicus* (Hwass, 1792), and *C. legatus* (Lamarck, 1810).

INTRODUCTION

Les îles Marquises, situées au cœur du Pacifique central, comptent parmi les archipels les plus isolés au monde. Cet éloignement a favorisé un endémisme remarquable, en particulier au sein des Conidae, une famille de gastéropodes marins. L'isolement géographique des Marquises a ainsi conduit à l'émergence d'espèces uniques, comme le démontrent plusieurs études (Petuch & Berschauer, 2020 ; Touitou & Balleton, 2022). Plus récemment, Lum (2023) a décrit *Textilia minamiae*, une autre espèce endémique de cet archipel, illustrant à nouveau la singularité de la biodiversité marine des Marquises.

Plus largement, la Polynésie française a fait l'objet de nombreuses recherches scientifiques, récentes et approfondies, attestant de la richesse de sa faune marine. L'ouvrage *Mollusques marins de Polynésie française* de Boutet, Gourguet

et Letourneux (2021) offre une vue d'ensemble des espèces présentes dans cette région. De plus, *Conidae de Polynésie française* de Balleton et Marti (2017), ainsi que *Panorama sur la diversité des Conidae en Polynésie française* par Richard et Rabiller (2022), fournissent des synthèses précieuses sur cette famille, répertoriant et illustrant de nombreuses espèces. Les travaux de Tenorio et Tucker (2013) ainsi que de Monnier *et al.* (2018) viennent également compléter cette documentation en illustrant la majorité des espèces contemporaines de Conidae. Cependant, il est surprenant de constater que l'espèce décrite dans cet article avait jusqu'ici été très peu documentée. Cette découverte souligne l'importance de poursuivre les recherches taxonomiques dans ces régions isolées, où de nombreuses espèces restent probablement à découvrir.

La description de *Cylinder patmartii* enrichit ainsi notre compréhension de la biodiversité marine de l'archipel des îles Marquises, prouvant que des espèces inédites continuent d'émerger de ces écosystèmes uniques de Polynésie française.

MATÉRIELS ET MÉTHODES

Les spécimens de *Cylinder patmartii* ont été collectés au cours de plongées sous-marines, réalisées entre 2014 et 2024, de jour comme de nuit, autour des îles de l'archipel des Marquises par Patrick Marti, Patrick Vappereau, et Michel Luciani, trois malacologues amateurs expérimentés résidant en Polynésie française. Malgré environ 300 plongées, seuls sept spécimens ont été trouvés à des profondeurs variant entre 32 et 45 mètres, exclusivement autour des îles de Fatu Hiva et Tahuata, ce qui souligne la rareté et la difficulté d'accès de cette espèce. Des plongées menées ailleurs en Polynésie française n'ont pas permis d'identifier d'autres individus, renforçant ainsi son caractère strictement localisé.

Les spécimens ont été découverts sur des substrats variés, tels que du sable, des roches, ou en étant manuellement délogés jusqu'à 20 cm de profondeur dans un substrat sablo-graveleux. Seuls des individus adultes, ayant atteint leur développement complet, ont été collectés. Une partie des spécimens a été séchée pour l'analyse morphologique, tandis que les autres ont été conservés dans de l'alcool neutre à 70 % en vue d'analyses moléculaires futures.

Les sept spécimens ont ensuite été comparés à un large échantillon d'espèces similaires provenant de diverses localités, avec des comparaisons taxonomiques principalement fondées sur l'holotype.

Les photographies des spécimens ont été prises à l'aide d'un appareil NIKON équipé d'un objectif macro de 105 mm f/2.8, pour minimiser les distorsions optiques. Un flash annulaire a permis d'assurer une illumination homogène, et les mesures ont été effectuées avec un pied à coulisse électronique, garantissant une précision optimale.

Abréviations et acronymes

- spm : spécimen.
- CFV : Collecte et collection Fabrice Volle.
- CML : Collecte et collection Michel Luciani.
- CPM : Collecte et collection Patrick Marti.
- CPV : Collecte et collection Patrick Vappereau.
- CVW : Collecte et collection Vincent Wagnier.
- Fig(s) : Figure(s).
- LC : longueur de la coquille.
- MNHN : Muséum national d'Histoire naturelle, Paris.
- MHNLR : Muséum d'Histoire Naturelle, La Rochelle.

Systématique

Phylum	Mollusca
Classe	Gastropoda
Sous-classe	Caenogastropoda
Ordre	Neogastropoda
Superfamille	Conoidea J. Fleming, 1822
Famille	Conidae J. Fleming, 1822
Genre	<i>Cylinder</i> Montfort, 1810

Cylinder patmartii sp. nov.

(Figures 1 à 8)

Matériel type.

- Holotype MNHN-IM-2019-1971, LC 44.6 mm, île de Fatu Hiva (Fig. 1).
- Trois paratypes CML, LC 45.5 & 58.4 mm, île de Fatu Hiva et 52.2 mm, île de Tahuata (Figs. 2-4).
- Deux paratypes CPV, LC 54.1 & 68.3 mm, île de Fatu Hiva (Figs. 5-6).
- Un paratype CPM, LC 52.3 mm, île de Fatu Hiva (Figs. 7-8).

Localité type. Archipel des îles Marquises, île de Fatu Hiva, à 45 mètres de profondeur, à la base d'un tombant, à l'interface entre zones sableuses et rocheuses.

Description. La coquille de *Cylinder patmartii* varie en taille entre 44,6 mm et 68,3 mm selon les spécimens collectés. La spire comporte de 6 à 9 tours, avec un dernier tour légèrement concave au niveau de l'épaule. La spire est rectiligne, et la surface du test est décorée de deux bandes transversales brunes, plus ou moins marquées, agrémentées de taches triangulaires blanches. L'ouverture, blanche et étroite, s'élargit progressivement vers la base. La lèvre externe, recourbée, laisse apparaître les motifs du test en bordure. La coquille, lisse et dépourvue de stries spirales apparentes, se distingue nettement de certaines espèces proches, comme *C. aureus*.

Caractéristiques de l'animal. L'animal se distingue par un pied

crème parsemé de taches brun-rouge diffuses, également visibles sur la face ventrale. Le siphon est d'un rouge orangé vif, avec un anneau noir bien défini, particulièrement remarquable. Les tentacules oculaires présentent une teinte blanche sur leur seconde moitié, du côté de l'œil (Fig. 8).

Comparaisons et critères de diagnose. *Cylinder canonicus* se distingue nettement de *C. patmartii* par son fond jaune, parcouru longitudinalement de motifs bruns ondulés et de lignes circulaires brunes. Son ouverture est généralement rose, et des taches écailleuses blanches, entourées de traits bruns, caractéristiques de *C. canonicus*, sont absentes chez *C. patmartii* (Fig. 11).

Cylinder legatus, quant à lui, est plus petit et plus élancé que *C. patmartii*. Sa coquille, aux teintes rosées ou orangées, présente de larges traits bruns ainsi que de petites taches blanches disposées en forme de cordes, avec une ouverture souvent rose. Ces caractéristiques permettent de le distinguer clairement de *C. patmartii*. De plus, à ce jour, *C. legatus* n'a pas été observé dans l'archipel des îles Marquises (Fig. 11).

Bien que *C. aureus* partage une taille similaire avec *C. patmartii*, il s'en différencie par ses taches légèrement plus larges, disposées verticalement sur un test traversé de fines stries spirales, surtout visibles sur le dernier tour de la coquille. De plus, *C. aureus* présente une coquille plus allongée et cylindrique, avec une spire plus haute et convexe. Le periostracum de *C. aureus* est fin et lisse, contrairement à celui de *C. patmartii*, qui est plus rugueux (Fig. 11).

Remarque. Bien que *C. patmartii* présente une ressemblance notable avec *C. aureus*, les différences morphologiques entre ces deux espèces sont constantes et significatives, suggérant une spéciation liée à l'isolement géographique de *C. patmartii* dans les îles Marquises. Cette nouvelle espèce semble être le fruit d'adaptations écologiques spécifiques à son environnement insulaire.

Étymologie. Cette espèce est dédiée à Patrick Marti, collectionneur passionné et naturaliste reconnu pour ses contributions significatives à l'étude des Conidae. Son dévouement à la malacologie et son appui constant aux recherches sur la biodiversité marine en font un allié précieux pour la communauté scientifique.

DISCUSSION

Parmi les espèces proches, *Cylinder aureus* est celle avec laquelle *C. patmartii* présente le plus de similitudes, pouvant entraîner des confusions taxonomiques. Dans leurs travaux, Boutet, Gourguet et Letourneux (2021) répertorient *C. aureus* en Polynésie française, et Richard et Rabiller (2022) indiquent qu'il se trouve «*sous les blocs, dans tout le complexe récifal, jusqu'à 30 m de profondeur*», en illustrant un spécimen provenant de Rangiroa, dans l'archipel des Tuamotu (taille : 47,7 mm, Collection MHNLR).

Bien que les auteurs n'aient pas eux-mêmes collecté cette espèce, ils ont observé deux spécimens pêchés vivants : l'un de 63,2 mm collecté par Vincent Wagnier sur l'île de Tahiti, archipel de la Société (Fig. 9), et un autre de 61,1 mm collecté par Fabrice Volle sur l'île de Hao, archipel des Tuamotu (Fig. 10), également illustré par Boutet, Gourguet et Letourneux (2021). Il est probable que les observations de *C. aureus* dans les

Marquises rapportées par Boutet, Gourguet et Letourneux (2021) ainsi que par Richard et Rabiller (2022) résultent d'une confusion avec *C. patmartii*, en raison du manque de distinctions taxonomiques précises à l'époque.

À notre connaissance, seuls Touitou et Balleton (2022) ont proposé une distinction, en examinant plusieurs spécimens de cônes récoltés lors de l'expédition MUSORSTOM 9 en 1997, menée par le MNHN dans les îles Marquises. Ces spécimens ont été considérés comme appartenant à une nouvelle espèce, qu'ils ont provisoirement rattachée à *Cylinder* aff. *aureus* (Hwass in Bruguière, 1792).

La reconnaissance de *C. patmartii* comme une espèce distincte repose sur plusieurs critères :

- Des critères morphologiques constants et uniques, qui ne varient pas entre les spécimens.
- L'absence d'espèce connue avec une similitude morphologique réellement évidente.
- Un fort taux d'endémisme dans l'archipel des îles Marquises, illustré par des espèces comme *Lambis pilsbryi* Abbott, 1961, différencié de *L. crocata* (Link, 1807), *Conus conco* Puillandre, Stöcklin, Favreau, Bianchi, Perret, Rivasseau, Limpalaër, Monnier & Bouchet, 2015, différencié de *C. lividus* Hwass, 1792, *Harpa kolaceki* Cossignani, 2011, différenciée de *H. amouretta* Röding, 1798, et, plus récemment, *Textilia minamiae* (Lum, 2023), différencié de *T. bullatus* (Linnaeus, 1758).

La découverte de *C. patmartii* suscite des questions intéressantes sur la dynamique évolutive des Conidae dans les îles Marquises, une région réputée pour son endémisme. Les ressemblances avec *C. aureus* pourraient indiquer une relation évolutive, où *C. patmartii* aurait divergé de cet ancêtre commun sous l'effet de l'isolement géographique extrême et des pressions écologiques particulières des Marquises.

Les différences morphologiques, bien que subtiles, sont constantes et significatives, suggérant une adaptation à des niches écologiques spécifiques aux îles Marquises. Le phénomène de micro-insularité, où des populations distinctes évoluent de manière isolée sur des îles ou dans des zones géographiques restreintes, pourrait expliquer cette possible endémicité stricte de *C. patmartii*.

La faible fréquence de collecte de *C. patmartii* pourrait témoigner soit d'une distribution saisonnière, soit d'une rareté réelle. Dans les deux cas, cela met en lumière l'importance cruciale de protéger les habitats marins de l'archipel des îles Marquises, où cette biodiversité unique et fragile est encore en grande partie méconnue.

Bien que l'analyse moléculaire reste nécessaire pour confirmer la position taxonomique de *C. patmartii*, les preuves morphologiques actuelles justifient son statut d'espèce nouvelle.

La reconnaissance de *C. patmartii* comme nouvelle espèce non seulement enrichit notre compréhension de la faune malacologique de l'archipel des îles Marquises, mais souligne également la nécessité pressante de poursuivre les recherches taxonomiques dans ces régions isolées. La protection de cet environnement unique, récemment inscrit au patrimoine mondial par l'UNESCO, est essentielle pour préserver la richesse et la diversité de la biodiversité mondiale.

CONCLUSION

L'isolement géographique et les conditions environnementales uniques des îles Marquises ont favorisé l'émergence d'espèces endémiques, telles que *Cylinder patmartii*. Cette découverte souligne non seulement l'importance de l'archipel dans l'étude de la spéciation et de l'endémisme au sein des Conidae, mais également la richesse biologique de cette région peu explorée. Le faible nombre de spécimens collectés indique une nécessité pressante de conservation des habitats marins de la région. La préservation de ces environnements est cruciale pour la survie de cette espèce et d'autres organismes marins qui y évoluent. De futures études moléculaires seront essentielles pour approfondir notre compréhension des relations phylogénétiques entre *C. patmartii* et ses espèces apparentées. Ces recherches permettront également d'évaluer les menaces potentielles pesant sur cette espèce nouvellement décrite, renforçant ainsi l'importance d'une approche proactive pour la protection de la biodiversité des îles Marquises.

La valorisation de cette biodiversité unique doit aller de pair avec des efforts de conservation adaptés, en vue de préserver le patrimoine naturel de cette région fragile et irremplaçable.

REMERCIEMENTS

Les auteurs expriment leur profonde gratitude au Dr. Philippe Bouchet, professeur émérite au Muséum national d'Histoire naturelle de Paris (MNHN) et au Dr. Éric Monnier pour leur relecture attentive du manuscrit et leurs contributions significatives à son amélioration ; ils remercient également chaleureusement les plongeurs locaux pour leur aide précieuse et Patrick Marti pour son soutien continu à la malacologie.

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Planche 1



Figure 1ab : *Cylinder patmartii* nov. sp., holotype, MNHN-IM-2019-1971, LC 44.6 mm, île de Fatu Hiva, - 45 m, collecté en février 2022. Photographies : Patrick Marti.

Planche 2



Figure 2ab : *Cylinder patmartii* nov. sp., paratype n° 1, CML, LC 45.5 mm, île de Fatu Hiva, - 35 m, collecté en février 2023.

Figure 3ab : *Cylinder patmartii* nov. sp., paratype n° 2, CML, LC 58.4 mm, île de Fatu Hiva, - 40 m, collecté en février 2023.

Figure 4ab : *Cylinder patmartii* nov. sp., paratype n° 3, CML, LC 52.2 mm, île de Tahuata, - 41 m, collecté en février 2023.

Photographies : Michel Luciani.

Planche 3



Figure 5ab : *Cylinder patmartii* nov. sp., paratype n° 4, CPV, LC 54.1 mm, île de Fatu Hiva (avec periostracum), - 37 m, collecté en mai 2021.

Figure 6ab : *Cylinder patmartii* nov. sp., paratype n° 5, CPV, LC 68.3 mm, île de Fatu Hiva (avec periostracum), - 35 m, collecté en mai 2021.

Photographies : Patrick Vappereau.

Planche 4

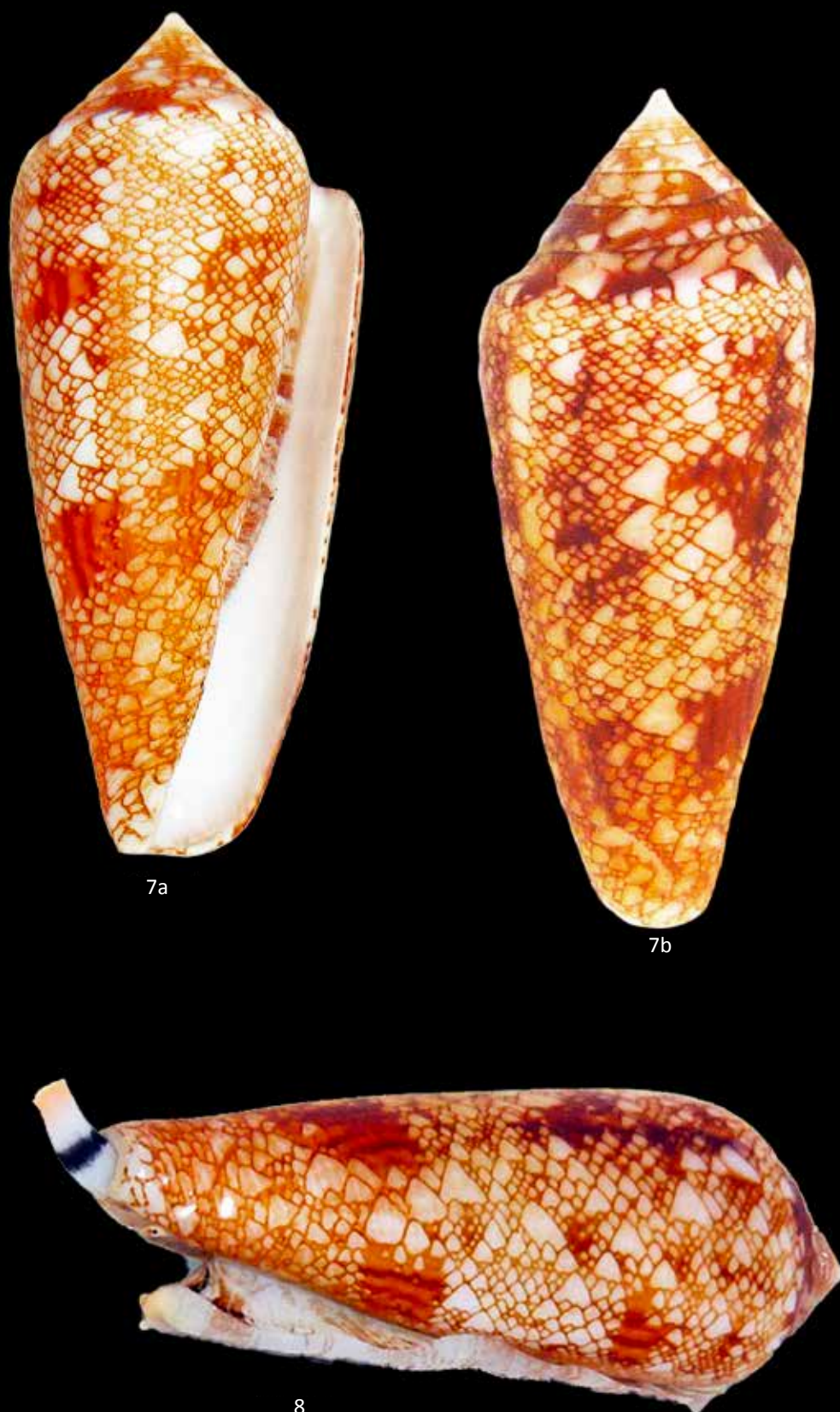


Figure 7ab : *Cylinder patmartii* nov. sp., paratype n° 6, CPM, LC 52.3 mm, île de Fatu Hiva, - 32 m, collecté en novembre 2019.

Figure 8 : *Cylinder patmartii* nov. sp., paratype 6 photographié vivant après sa collecte, en bac, à bord. Photographies : Patrick Marti.

Planche 5



Figure 9ab : *Cylinder aureus* (Hwass, 1792), CVW, LC 63.2 mm, île de Tahiti, archipel de la Société, collecté en 2012. Photographies : Vincent Wagnier.

Figure 10ab : *Cylinder aureus* (Hwass, 1792), CFV, LC 61.1 mm, île de Hao, archipel des Tuamotu, collecté en 2019. Photographies : Fabrice Volle.

Planche 6

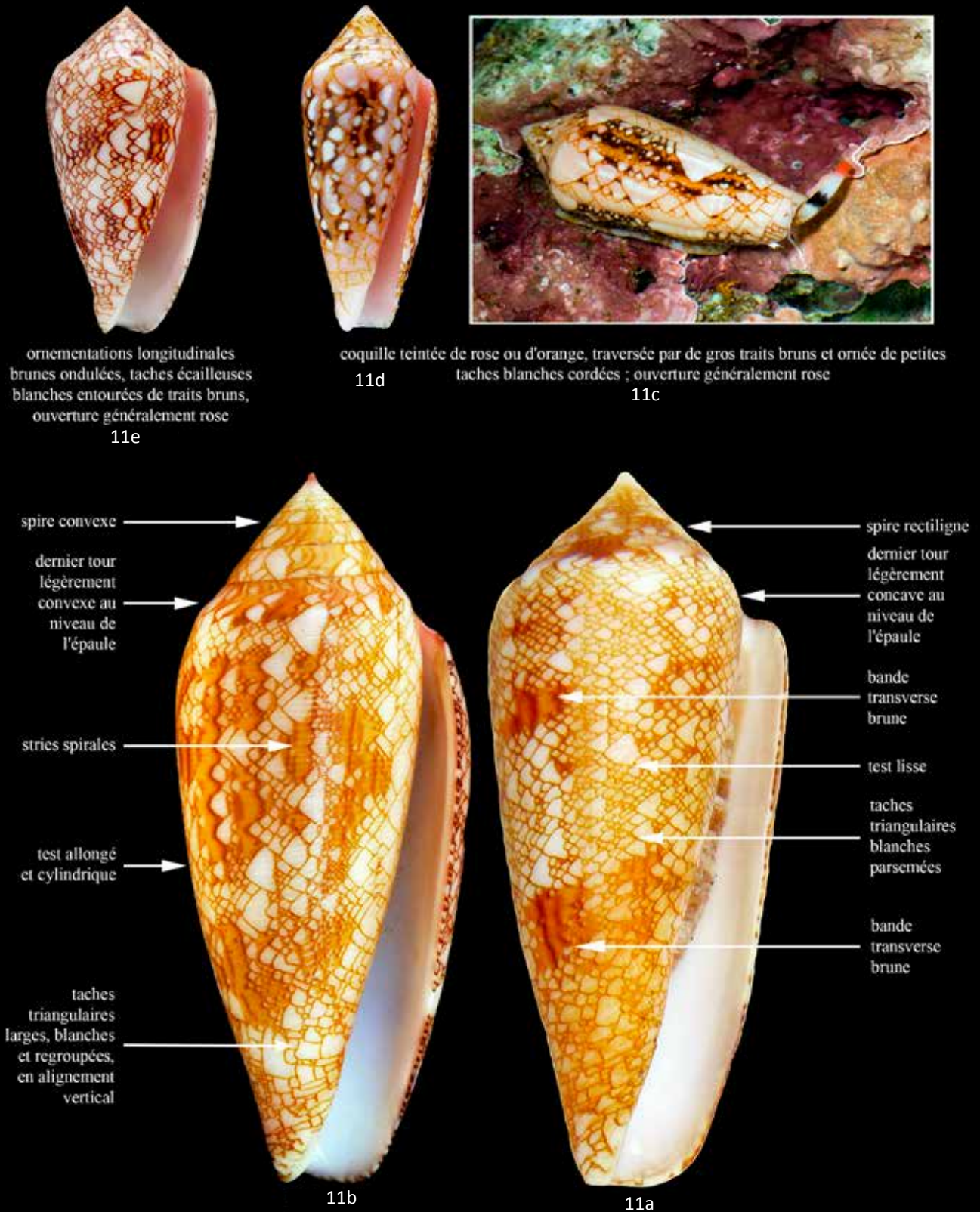


Figure 11a : *Cylinder patmartii* nov. sp., holotype, MNHN-IM-2019-1971, LC 44.6 mm, île de Fatu Hiva, - 45 m, collecté en février 2022. Photographie : Patrick Marti.

Figure 11b : *Cylinder aureus* (Hwass, 1792), CFV, LC 61.1 mm, île de Hao, archipel des Tuamotu, collecté en 2019. Photographie : Fabrice Volle.

Figure 11c : *Cylinder legatus* (Lamarck, 1810), CPM, LC 38 mm, île de Tahiti, archipel de la Société, - 38 m, collecté en avril 2018. Photographie : Patrick Marti.

Figure 11d : *Cylinder legatus* (Lamarck, 1810), LC 32 mm, île de Tahiti, archipel de la Société. Photographie : Philippe Bacchet.

Figure 11e : *Cylinder canonicus* (Hwass, 1792), CPM, LC 57,5 mm, île de Nuku Hiva, - 25 m, collecté en février 2019. Photographie : Patrick Marti.

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ABSTRACT.

A new species closely related to *Phyllonotus pomum* (Gmelin, 1791), *P. bellettii* Petuch, Berschauer & Powell, 2024, has just been described from Guadeloupe. Its validity is revised in this paper. Sequencing of a large sampling of populations collected during the MNHN's KARUBENTHOS 1 campaign in Guadeloupe has been carried out. The results clearly showed the presence of a single species in spite of a great conchological variability. *Phyllonotus belletti* was consequently put in synonymy of *P. pomum*.

KEY WORDS.

Muricidae, *Phyllonotus bellettii*, *P. pomum*, new species, variability, sequencing, phenotypic plasticity, synonymy, Guadeloupe.

INTRODUCTION.

Phyllonotus pomum is widely distributed throughout the Caribbean region. This species is quite commonly found in shallow coastal areas. Its variability has long intrigued us, so much so that a few years ago we seriously considered the coexistence of two species under this taxon (Garrigues & Lamy, 2019). However, molecular biology suggests that these different morphs are conspecific.

MATERIAL AND METHODS.

The specimens studied were collected by diving during the MNHN's KARUBENTHOS 2012 campaign in Guadeloupe and by the two authors. Most of them could be sequenced using the COI gene of the mitochondrial genome, a decisive technique in the separation of different species (Puillandre, 2019).

Abbreviations. MNHN: Muséum national d'Histoire naturelle (recent molluscs section), Paris, France.

BG: Bernard Garrigues collection (Lalande, France)

DL: Dominique Lamy collection (Baie-Mahault, Guadeloupe)

Systematics.

The systematics and identity of *Phyllonotus pomum*, has already been extensively detailed in previous papers (see Vokes, 1990; Merle *et al.*, 2011; Garrigues & Lamy, 2016, 2019; Lamy & Pointier, 2017).

Habitat. *Phyllonotus pomum* is widespread from Florida in the north to the Recife region of Brazil in the south, and throughout the Caribbean area. In Guadeloupe, it occupies two very different types of habitat. The first is located on the Atlantic side and in the Grand and Petit-Cul-de-sac-marin, on algal reef flats dotted pockets of sand. The second one extends along the leeward coast (pl. II, figs 1-3). The environment consists of calm zones between 0 and 20 m depth, covered with black volcanic sand and dotted with large boulders. The mollusc, often buried in the substrate, sometimes leaves only a small dorsal part of its shell protruding.

Dietary regime.

Phyllonotus pomum has a different diet depending on its habitat. Specimens on the leeward coast feed mainly feed on Spondylidae (*Spondylus tenuis* Schreibers, 1793) and dead fish. The reef form of the Grand-cul-de-sac-marin has a more diversified regime: *Spondylus americanus* Hermann, 1781, *Arca zebra* (Swainson, 1833), *Anadara notabilis* (Röding, 1798), *Arca imbricata* Bruguière, 1789, *Modiolus americanus* Leach, 1815 and *Chama macerophylla* Gmelin, 1791.

Shell Morphology. Each of these two habitats is occupied by a particular morph or form which can be distinguished by a number of shell characters. To simplify we will speak of the reef form and the leeward coast form.

Both forms share a protoconch of 2.5 whorls, or 1.7 whorls if the nucleus (embryonic shell) is excluded. This suggests a non-planktotrophic larval development. The microsculpture of this protoconch, starting at the nucleus and extending for around 11/4 whorls, has a glossy appearance with fine spiral cords. It then gradually takes on a matt, granular texture. Hatching probably precedes metamorphosis by a short time, or is only separated by a very short planktonic stage.

Teleoconch has 8 whorls, sometimes 7 in both forms. The reef form (pl. I, figs 1-10) is smaller overall, with an average adult size of around 65 mm, not exceeding 85 mm.

In comparison, the leeward shore form (pl. I, figs 11-18) is significantly larger, with an average size of around 100 mm. The largest specimens in this population exceed 130 mm.

The shoulder is more pronounced in the reef form giving it an almost triangular appearance. In most cases, there is only one large intervarical nodule. Dark spiral bands contrast with the lighter background color. These specimens are often covered with concretions.

Shells from the leeward coast have a more ovoid shape, proportionately less wide, with two to three less prominent intervarical nodules, more squamous sculpture and more developed foliations on the convex part of the whorl. Their color is uniformly brown, rather dark, although varicose veins may be lighter.

The ratio of shell height to diameter is between 1.48 and 1.64 for the reef form, and between 1.64 and 1.80 for the leeward coast form.

The siphonal canal is shorter in the majority of specimens of the reef form than in the other form.

The anal canal of the leeward coast form is more indented, corresponding to an operculum whose adapical part is rather pointed (pl. I, fig. 13c).

Phylogenetic analysis and conclusion. Molecular biology results suggest quite clearly that the two forms belong to the same species, *Phyllonotus pomum*! Thirteen specimens from Guadeloupe belonging to each form were sequenced and all have very similar sequences. These two forms could be described as ecophenotypic. A given genotype has produced different phenotypes in response to different environmental conditions. In other words, *P. pomum* from the reefs of Guadeloupean Atlantic coast, as well as those of the Grand and Petit-Cul-de-sac-marin and the leeward coast, are a single species, displaying particular conchylological characters in response to their environment.

Phyllonotus bellettii, whose type locality is Pointe Noire on the leeward coast of Guadeloupe, at a depth of 7 m, is a typical form of this zone and is synonymous with all *P. pomum* from Guadeloupe.

Phylogenetic tree. This tree is obtained from COI sequences (maximum likelihood tree with 100 bootstraps, created using MEGA software).

The provenance of these specimens, most of which are illustrated on Plate I, should be noted. Specimens IM-2013-19522, 19524, 19525, 19526, 19532 and 19589 come from Ilet Fortune in the Petit-cul-de-sac-marin. Specimens IM-2013-20044, 20946 and 20896 are from Baie-Mahault in the Grand-cul-de-sac-marin. Specimen IM-2013-8250 is from Bouillante on the leeward coast; IM-2013-43970 and 43971 are from the Basse Terre region, also on the leeward coast. Finally, IM-2013-20015 is from Anse Babin near Morne-à-l'eau.

A closer look at these locations reveals that some specimens, typical of the shape of the leeward coast, were collected in the Grand-cul-de-sac-marin (Baie-Mahault, Anse Babin). Although these sites are not far from the Ilet Fajou flats, their sandy-muddy biotope is very different.

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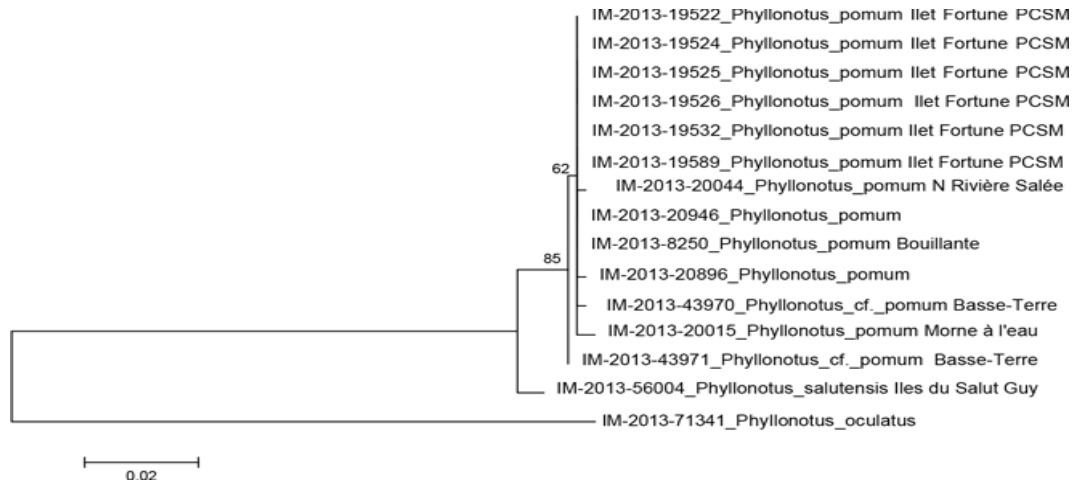


Figure 1: Phylogénétique tree obtained from sequencing COI (tree in maximum likelihood with 100 bootstraps, done with MEGA software)

MNHN	BOLD	GB COI
IM-2013-19522	MUBA946-24	PQ279513
IM-2013-19524	MUBA947-24	PQ279510
IM-2013-19525	MUBA955-24	PQ279507
IM-2013-19526	MUBA948-24	PQ279512
IM-2013-19532	MUBA949-24	PQ279505
IM-2013-19589	MUBA950-24	PQ279504
IM-2013-20015	MUBA951-24	PQ279508
IM-2013-20044	MUBA952-24	PQ279509
IM-2013-20896	MUBA953-24	PQ279511
IM-2013-20946	MUBA954-24	PQ279506
IM-2013-43970	MUBA956-24	PQ279503
IM-2013-43971	MUBA957-24	PQ279502
IM-2013-56004	MUBA740-16	XX422193
IM-2013-71341	MUBA846-18	MX308438
IM-2013-8250	MUBA613-15	XX422200

Figure 2: Genbank numbers of the specimens of *Phyllonotus pomum* studied by the MNHN

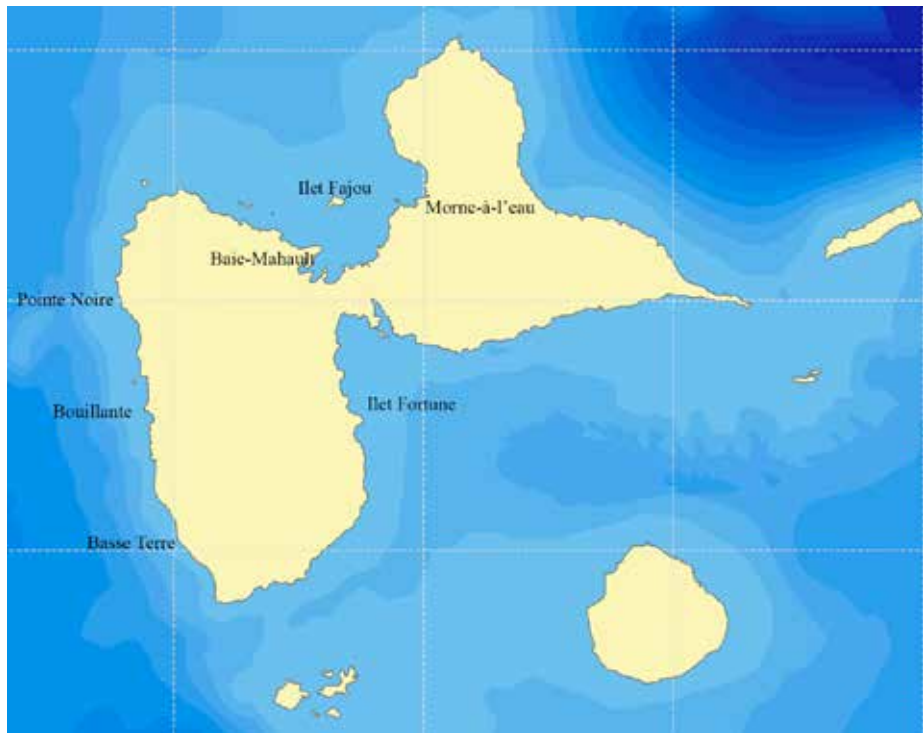


Figure 3: Localisations in Guadeloupe of the *Phyllonotus pomum* studied

Plate 1 *Phyllonotus pomum* Guadeloupe



Living specimenn his habitat, Basse Terre



Female on his egg mass, Basse Terre



Adult specimen in sand, Basse Terre

Plate 2 *Phyllonotus pomum*, Guadeloupe



ADDITIONS TO THE MARINE MALACOFaUNA OF CYPRUS (BY MAY 2024)

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KEYWORDS

Cyprus, Mediterranean Sea, Mollusca, New Records

ABSTRACT

The present work is the continuation of the investigation on the marine mollusks of Cyprus by April 2024. It incorporates 16 species out of which one is a first and another one a second records, respectively, and 10 and 4 are first and second image documented records, respectively. Essential taxonomic characteristics and ecological data, such as distribution, habitat and origin are presented.

INTRODUCTION AND METHODS

The status of the marine molluscs' diversity in the Cypriot waters and the methods of study applied have been recently presented in Varnava *et al.*, 2024. The scientific names of species are based on WoRMS (May 2024). The collection stations are indicated in Fig. 1, while the representatives of the collected specimens are depicted in Fig. 2. All specimens are deposited in the premises of D. Kolokotronis, M. Papatheodoulou and Y. Varnava who give access to the biological material for inspection by appointment.

RESULTS AND DISCUSSION

Abbreviations: Be – beached; C1 – first record for Cyprus; C2 – second record for Cyprus; DC1 – first image documented record for Cyprus; DC2 – second image documented record for Cyprus; JSP - Jubilee Shoals Pissuri; Li – Limnitis; Lio - Liopetri; Par – Paralimni; PURC – previous undocumented report for Cyprus; RB – rocky bottom; Ri-Di SC - Rizokarpaso – Dikarpaz, South Coast; SB – sandy bottom; So – Sotira; Sof – Softades; Xy – Xylophagou.

THE RECORDS

BIVALVIA

Nucula nitidosa Winckworth, 1930 (Fig. 2a), 10 shells, Be, Sof 34°49'N -33°33', PURC Öztürk *et al.* (2004), DC1, description in Gofas *et al.* (2011).

Megaxinus unguiculinus Pallary, 1904 (Fig. 2b), 6 shells, SB, 21-23 m, Par 35°04'N -34°01' and Xy 34°56'N -33°49'E, PURC Di Franco *et al.* (2021), DC1, presented in Huber (2015).

Arcopella balaustina (Linnaeus, 1758) (Fig. 2c), 2 shells, SB, 18 m, Xy 34°56'N -33°49'E, DC1 from synonym Tornaritis (1987), DC2, description in Gofas *et al.* (2011).

GASTROPODA

Gibbula philberti (Récluz, 1843) (Fig. 2d), 10 shells, Be, Sof 34°49'N -33°33' and So 34°59'N -33°57', PURC Öztürk *et al.* (2004), DC1, description in Gofas *et al.* (2011).

Dizoniopsis coppolae (Aradas, 1870) (Fig. 2e), 3 shells, Be, Lio 34°58'N -33°53' and 30 m, RB, JSP 34°38'N -32°43', PURC Öztürk *et al.* (2004), DC1, presentation in Giannuzzi-Savelli *et al.* (1999).

Melanella frielei (H. K. Jordan, 1895) (Fig. 2f), 2 shells, Be, Lio 34°58'N -33°53' and Li 35°10'N -32°44'E, C1, description in Gofas *et al.* (2011).

Parvioris ibizenca (F. Nordsieck, 1968) (Fig. 2g), 10 shells, Be, So 34°59'N -33°57', PURC Öztürk *et al.* (2004), DC1, description in Gofas *et al.* (2011).

Alvania geryonia (Nardo, 1847) (Fig. 2h), 1 shell, Be, Li 35°10'N -32°44'E, PURC Öztürk et al. (2004), DC1, description in Gofas et al. (2011).

Alvania mamillata Risso, 1826 (Fig. 2i), 1 shell, Be, Lio 34°58'N -33°53', PURC Öztürk et al. (2004), DC1, description in Gofas et al. (2011).

Pusillina lineolata (Michaud, 1830) (Fig. 2j), 6 shells, Be, Lio 34°58'N -33°53' and 30 m, JSP 34°38'N -32°43', PURC Öztürk et al. (2004), DC2, description in Scaperrotta et al. (2013).

Trivia levantina Smriglio, Mariottini & Buzzurro, 1998 (Fig. 2k), 1 shell, Be, So 34°58'N -33°55'E, PURC Öztürk et al. (2004), DC1, description in Smriglio et al. (1998).

Teretia teres (Reeve, 1844) (Fig. 2l), 1 shell, Be, Sof 34°49'N -33°33', C1 Öztürk et al. (2004), C2, description in Gofas et al. (2011).

Mitromorpha olivoidea (Cantraine, 1835) (Fig. 2m), 1 shell, Be, Li 35°10'N -32°44'E, PURC from synonym Öztürk et al. (2004), DC1, description in Gofas et al. (2011).

Ringicula auriculata (Ménard de la Groye, 1811) (Fig. 2n), 1 shell, Be, Ri-Di SC 34°38'N -32°43', DC1 Tornaritis (1987), DC2, description in Gofas et al. (2011).

Limacina trochiformis (d'Orbigny, 1835) (Fig. 2o), 1 shell, MB, 30 m, JSP 34°38'N -32°43', PURC Öztürk et al. (2004), DC1, description in Gofas et al. (2011).

Tylodina perversa (Gmelin, 1791) (Fig. 2p), 3 shells, Be, Lio 34°58'N -33°53' and AT 34°58'N -33°56'E, DC1 <https://www.facebook.com/photo?fbid=10159417324221993&set=gm.3516982528561931&idortvany=1519592034967667>, DC2, description in Gofas et al. (2011).

DISCUSSION

The present work incorporates 16 species out of which one is a first and another one a second records, respectively, and 10 and 4 are first and second image documented records, respectively. All presented material are shells collected from the sublittoral zone or the beach. The beached ones, in particular, were present in the thanatocoenoses thus giving an idea of the local biodiversity as the study of thanatocoenoses is an approximate method in the assessments of that (Battisti & Cerfolli, 2016). We are of the opinion that the local biodiversity is in need of farther and thorough revision hence the necessity of cooperation between the two ethnic communities. In essence, the Cypriot molluscan biodiversity is not explored adequately yet, and more species are waiting to see the light of publicity. The clean oligotrophic waters of the East Mediterranean Sea, the variety of substrates, depths and costal length and diversity, the climate change as well as the human activities (e.g. navigation, expansion of the Suez Canal) seem to be in support of our anticipations. The climate change transformation of the Mediterranean Sea, in particular, from temperate to subtropical environment and the human activities that induce a dynamic status to increase the alien biodiversity by means of species introduction from tropical and subtropical seas make ever more crucial for the scientists to turn and focus

their attention on those effects on the molluscs' diversity of the Island. In addition, any new alien record comprises an alert signal for its expansion, distribution and future effects on the whole Mediterranean Sea environment. The work continues on all molluscan families and expands in more areas and depths of the Cypriot Seas.

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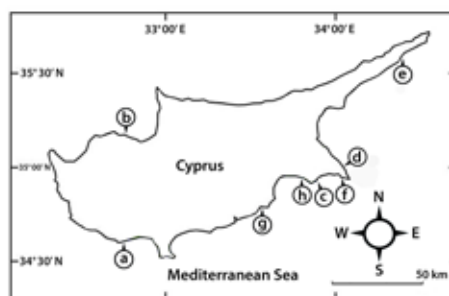


Figure 1: a. Jubilee Shoals Pissuri; b. Limnitis; c. Liopetri; d. Paralimni; e. Rizokarpaso – Dikarpaz, South Coast; f. Sotira; g. Softades; h. Xylophagou



Figure 2: a. *Nucula nitidosa* Winckworth, 1930; b. *Megaxinus unguiculinus* Pallary, 1904; c. *Arcopella balaustina* (Linnaeus, 1758), two different shells; d. *Gibbula philberti* (Récluz, 1843); e. *Dizoniopsis coppolae* (Aradas, 1870); f. *Melanella frielei* (H. K. Jordan, 1895); g. *Parvioris ibizenca* (F. Nordsieck, 1968); h. *Alvania geryonia* (Nardo, 1847); i. *Alvania mamillata* Risso, 1826; j. *Pusillina radiata* (R. A. Philippi, 1836); k. *Trivia levantina* Smriglio, Mariottini & Buzzurro, 1998; l. *Teretia teres* (Reeve, 1844); m. *Mitromorpha olivoidea* (Cantraine, 1835); n. *Ringicula auriculata* (Ménard de la Groye, 1811); o. *Limacina trochiformis* (d'Orbigny, 1835); p. *Tylodina perversa* (Gmelin, 1791). Barr = 1 mm, unless otherwise indicated.



Ransoniella punctata (Linnaeus, 1771)
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KEYWORDS

Biodiversity, Greece, Mediterranean Sea, Mollusca, New Records

ABSTRACT

The Hellenic marine malacofauna is affected by current environmental changes, ongoing research, and the introduction of alien species. Out of the species in this work: *Alveinus miliaceus* (Issel, 1869) and *Pyrgulina maiae* Hornung & Mermod, 1924 are first records for Greece of those Lessepsian migrants, *Parthenina dantarti* (Peñas & Rolán, 2008) is second record, *Finella pupoides* A. Adams, 1860 is third record while *Turbonilla lactea* (Linnaeus, 1758) is first image documented record. Taxonomy is followed by discussion.

INTRODUCTION

Biological invasions are considered as seriously threatening the global biodiversity thus constituting a crucial pervasive factor of its change (Galil 2007; Molnar *et al.*, 2008; Occhipinti-Ambrogi & Galil, 2010). The Mediterranean Sea is a hotspot of biodiversity and endemism (Bianchi & Morri 2000; Coll *et al.*, 2010) and is affected by the so-called “Lessepsian invasion” or “Erythrean invasion” a major biological marine invasion that followed the opening of the Suez Canal in 1869, especially after its recent widening. The Canal is an artificial waterway and also the most significant vector for introductions of non-indigenous species (NIS) to the Mediterranean basin, followed by shipping (Galil 2008; Galil *et al.*, 2016; Zenetos, 2017). The Levantine Sea in the southeastern Mediterranean is primarily affected due to its proximity to the Suez Canal and the prevailing surface currents (Galil, 2008, 2016; Tzomos *et al.*, 2012; Galil *et al.*, 2016). Biological invasions have the potential to radically change the structure and functioning of the recipient communities (Molnar

et al., 2008; Ehrenfeld, 2010; Fanelli *et al.*, 2015), thus causing extensive ecological, social and economic damage apart from the negative affects on the local biodiversity (Galil, 2007; Pyšek & Richardson, 2010; Simberloff *et al.*, 2013), particularly if NIS affect the provision of ecosystem services (Wallentinus & Nyberg, 2007) and/or adversely affect human health (Mazza *et al.*, 2014). Taking into account the above and the rapid range expansions observed for many Lessepsian invaders originally restricted to the Levantine Basin (Galil, 2009; Tzomos *et al.*, 2012; Galil *et al.*, 2016), it is of paramount importance the early reporting of new findings as it essentially aids to the better understanding of the dynamics of the invasion and to the early recognition of potentially invasive species, when mitigation measures are most effective (Crooks, 2005; Simberloff *et al.*, 2013). This invasion is nowadays registered from adjacent countries (e.g. Katsanevakis *et al.*, 2009; Manousis *et al.*, 2010; Çinar *et al.*, 2011; Manousis & Galinou-Mitsoudi, 2013; Ammar, 2018; Manousis *et al.*, 2018; Zenetos *et al.*, 2018; Bariche & Fricke, 2020; Crocetta *et al.*, 2020; Manousis *et al.*, 2020a, b, c; Mbazios *et al.*, 2020; Kontadakis *et al.*, 2021; Manousis *et al.*, 2021; Zaminos *et al.*, 2021; Manousis *et al.*, 2022a, b; Mbazios *et al.*, 2022; Kolokotronis *et al.*, 2023) and some have reached the Central Mediterranean, e.g., Tunisia (e.g. Ounifi-Ben Amor *et al.*, 2016), Italy (e.g. Occhipinti-Ambrogi *et al.*, 2011), and France (e.g. Daniel *et al.*, 2009; Bodilis *et al.*, 2011). Moreover, accelerating climate warming is especially affecting the Eastern Mediterranean Basin (Ozer *et al.*, 2017), causing the decline of indigenous species and potentially more favourable conditions for the establishment of tropical species (Rilov, 2016; Albano *et al.*, 2021). The Hellenic marine molluscan biodiversity has been reviewed two years ago in a systematic catalogue by Manousis (2021) and further updated recently (Manousis, 2023). Perceptive human resources management in the form of citizen science monitoring projects have shown that the Hellenic Seas

are in need of further search (Galinou-Mitsoudi *et al.*, 2023). Considering the above issues, the aim of this study is to further investigate the presence of autochthonous and allochthonous molluscan species in localities of the Hellenic Seas.

METHODS

Sampling, handling and species recognition were conducted according to Manousis & Galinou-Mitsoudi (2013) and Mbazios *et al.* (2020) from May 2023 to July 2024 in locations of the Hellenic Seas. All 1st and 2nd documented records for the Mediterranean and the Hellenic Seas are listed below with collection data. Findings are presented in Figure 1. Specimens are deposited in the premises of P. Ovalis and G. Jaux. Scientists are welcome to access the biological material for inspection by appointment.

RESULTS

Abbreviations: **DG1** – first image documented record for Greece; **G1, G2, G3** – first, second and third record for Greece, respectively; **KLM** – Known Lessepsian Migrant; **MB** – Mixed Bottom; **NLM** – New Lessepsian Migrant; **PDI** – previous dubious identification; **PMS** – previous misidentified specimen; **PURG** – previous undocumented record for Greece

THE RECORDS

Bivalvia

Alveinus miliaceus (Issel, 1869) (Kelliellidae) (Fig. 1a). 4 fresh shells and 4 valves, MB, 8 m, Pefki Beach, Rhodes Island 36°04'N - 28°02'E, G1, NLM.

Gastropoda

Finella pupoides A. Adams, 1860 (Scaliolidae) (Fig. 1b). 3 shells, MB, 12 m, Prasoniki, Rhodes Island 35°52'N - 27°45'E, G1 Manousis (2021), G2 Ovalis & Zenetos, 2024, G3, KLM.

Parthenina dantarti (Peñas & Rolán, 2008) (Pyramidellidae) (Fig. 1c). 3 shells, MB, 8 m, Pefki Beach, Rhodes Island 36°04'N - 28°02'E, G1 Manousis (2021), G2, description in Penas *et al.* (2008) and in Giannuzzi-Savelli *et al.* (2014).

Pyrgulina maiiae Hornung & Mermod, 1924 (Pyramidellidae) (Fig. 1d). 1 shell, MB, 8 m, Pefki Beach, Rhodes Island 36°04'N - 28°02'E, G1, description in Giannuzzi-Savelli *et al.* (2014).

Turbonilla lactea (Linnaeus, 1758) (Pyramidellidae) (Fig. 1e). 1 shell, MB, 7 m, Kalamaki, Messinia, 36°55'11N, 21°55'58E, PURG Koukouras (2010), PDI Tenekides (1989), PMS Manousis (2012), DG1. The species is characterized by its B type protoconch of some 300 µm wide. Description in Giannuzzi-Savelli *et al.* (2014).

DISCUSSION

This work includes collection results of the period from May 2023 to July 2024 and is part of the ongoing study of the marine malacofauna in locations throughout the Hellenic Seas. Out of the 5 image documented species in this work: first record for the Hellenic Seas is *Alveinus miliaceus* (Issel, 1869); second

records are *Finella pupoides* A. Adams, 1860, *Parthenina dantarti* (Peñas & Rolán, 2008) and *Pyrgulina maiiae* Hornung & Mermod, 1924, while *Turbonilla lactea* (Linnaeus, 1758) is a first image documented species for the area. *Alveinus miliaceus*, a known Lessepsian migrant, is a minute bivalve that reaches a shell size of some 2 mm (Oliver & Zuschin 2000), is a poorly known species originally described by Issel (1869) from the Gulf of Suez, and known also from the Bay of Safaga, Egypt, in the northern Red Sea and Oman (Oliver & Zuschin, 2000). It was rather recently re-described by Oliver & Zuschin (2000). Here, we report its first record for the Hellenic Seas. First reported for the Mediterranean Sea from Israel by Steger *et al.* (2018). *Finella pupoides*, a species of variable shape, sculpture and color, has been established in the Mediterranean Sea some decades ago (Bogi & Galil, 2013). Beside of the coasts of Israel and Turkey, it has been reported from Anafi Island, Greece (Manousis, 2021), Saronikos Gulf (Ovalis & Zenetos, 2024), and now from the south of Rhodes Island. All introduced species, independently of the introduction path and their robustness, need significant parameters in order to survive and settle and establish or dominate in their new areas, such as continual favorable environmental similar to the conditions of their origin, and luck of pollution and predators. Nevertheless, the case that all needed appropriate parameters coincide is rare and many alien individuals disappear or remain alive for short periods. Perhaps, this is the reason that first findings are shells or fresh dead specimens. As the climatic change is a fact, the efforts to find new records are valuable for the biodiversity status of any locality.

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Figure legend

Figure 1: a. *Alveinus miliaceus* (Issel, 1869); b. *Finella pupoides* A. Adams, 1860; c. *Parthenina dantarti* (Peñas & Rolán, 2008); d. *Pyrgulina maiae* Hornung & Mermod, 1924; e. *Turbonilla lactea* (Linnaeus, 1758). Barr = 1 mm, unless otherwise indicated.

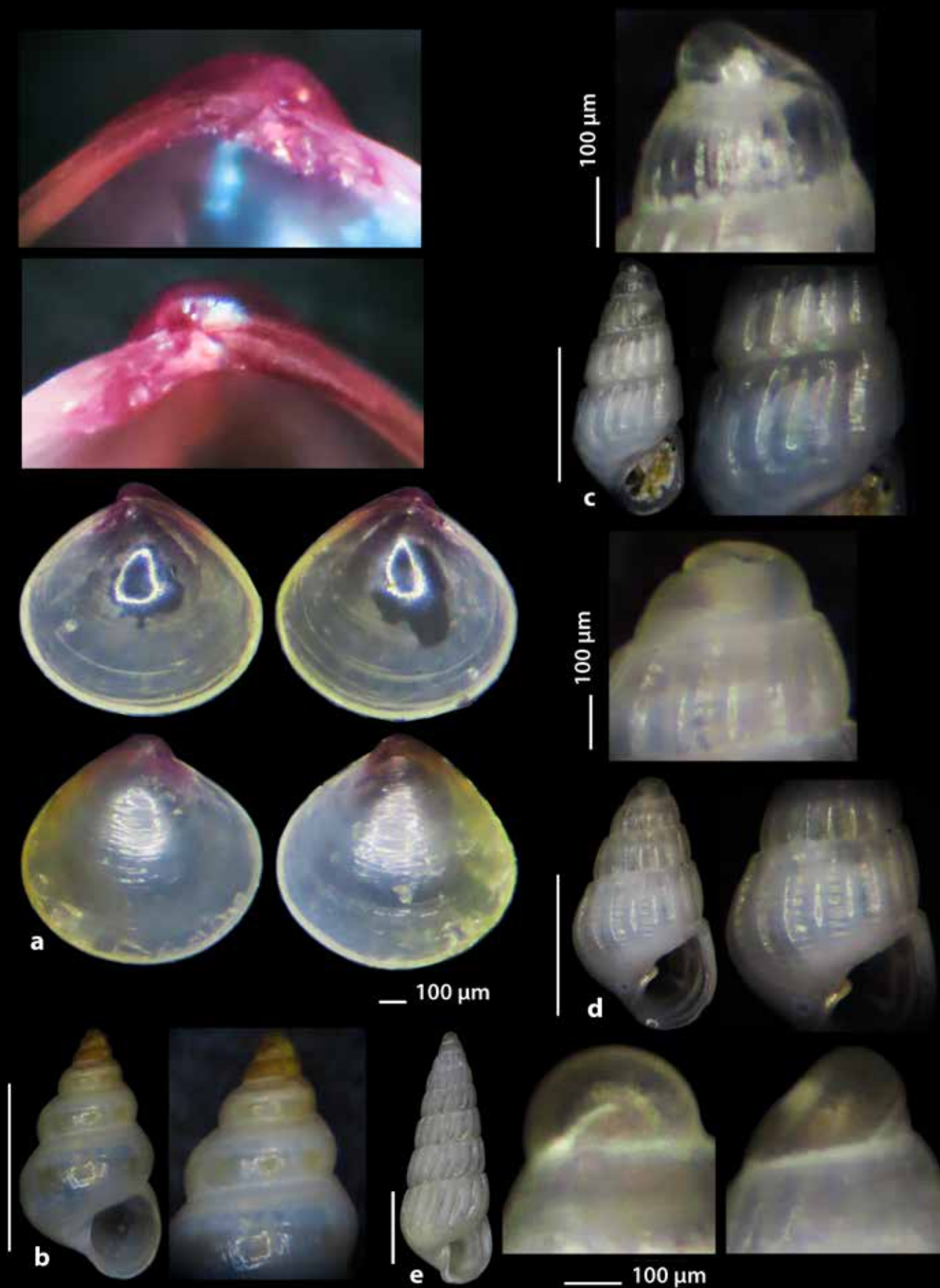


Figure 1: a. *Alveinus miliaceus* (Issel, 1869); b. *Finella pupoides* A. Adams, 1860; c. *Parthenina dantarti* (Peñas & Rolán, 2008); d. *Pyrgulina maiae* Hornung & Mermod, 1924; e. *Turbonilla lactea* (Linnaeus, 1758).

Barr = 1 mm, unless otherwise indicated

THE MARINE MOLLUSCA OF CYPRUS (BY SEPTEMBER 2024): AN UPDATED, SYSTEMATIC CATALOGUE, DOCUMENTED WITH BIBLIOGRAPHIC AND PICTORIAL REFERENCES

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KEYWORDS

Biodiversity, Cyprus, Marine Mollusca, Systematic Catalogue

ABSTRACT

This survey provides with an updated, systematic catalogue documented with bibliographic and pictorial references on the extant marine Mollusca present in the Cypriot Economic Exclusive Zone, by September 2024. It adds 121 species for the area to the previous taxa register. It incorporates 871 species belonging to 195 families and referred to in some 90 publications, and increases the Cypriot malacofauna by 15%. The up-to-date nomenclature is based on the WoRMS Database.

INTRODUCTION

The abrupt decline of the biodiversity of the planet during the second half of the 20th and the beginning of the 21st century causes a crisis that we need to address by understanding its extent and anticipate its consequences by means of an overall knowledge of the biological diversity, ecology and evolutionary history of the organisms. Accordingly, the published data on the marine molluscs' diversity of the Cypriot Seas were based on faunistic publications which referred mainly to relatively large species and, in some cases, included no documenting images. Consequently, smaller species remained underestimated and lacking from the recorded fauna. In addition, the collection efforts were rather obsolete; hence the published literature leads to an inaccurate profile of the present malacofauna and, unavoidably, includes possibly extinct species. At the same time, the relevant faunistic books of Stavrinos (1974), Tornaritis (1987) and Orr (2000) are rather old, thus calling for a revision and renewal. The most recent source of information on the marine molluscs of the area is the checklist by Kolokotronis *et al.* (2022) with 185 families and 755 species, already exiding by some 60 species that of Öztürk *et al.* (2004). Lately, the Cypriot marine malacofauna was enriched with new findings (e.g. Kolokotronis *et al.*, 2023; 2024a, b; Papatheodoulou *et al.*, 2024; Varnava *et al.*, 2024a, b, c) while the nomenclature has chanced considerably the last couple of years. Our general aim is to update the knowledge on the Cypriot marine malacofauna by intensive field work and with this work, in particular, to present the findings by an updated catalogue including documentation references. Apparently, some 680 consultations, in two years of the previous relevant catalogue (Kolokotronis *et al.*, 2022) in the ResearchGate portal is an indication of the necessity of the present step.

MATERIALS AND METHODS

All relevant studies in the last five decades or so on the extant marine Mollusca of the Cypriot Seas were retaken into account to produce this systematic catalogue. For reasons of economy of space, the references are numbered in a nearly absolute alphabetical and chronological order. The nomenclature followed is that of the WoRMS Database (assessed, September 2024). Each species is accompanied by the reference in which is depicted or mentioned. Marked with an asterisk (*) are those references which report but do not depict the relevant species - an unavoidable compromise, as there is no other documentation on that species presence and they cannot be ignored due to uncertainty. Dubious identifications due to lack of detailed presentation, misidentifications, species of questionable establishment or questionable documentation and origin are also appropriately marked.

RESULTS AND DISCUSSION

Abbreviations of localities

AT - Agia Triada;
AA-Es - Agios Amvrosios - Esentepe;
AGA - Agios Georgios Alamanou;
AGP - Agios Georgios Pegia;
AE-Ça - Agios Epikytos - Çatalköy;
Aka - Akamas;
Akr - Akrotiti;
AI-Ak - Ayia Irini - Akdeniz;
Am - Amathus (Lim);
AN - Ayia Napa;
AP-Di - Agios Philon (Rizokarpaso) - Dipkarpaz;
A-T - Akanthou - Tatlisu;
AT - Ayia Thekla;
ATS - Agia Thekla - Sotira;
Bo - Bogazi - Boghaz;
CAA-ZB - Cape Apostolos Andreas - Zafer Burnu;
CA - Cape Arnautis;
CCP - Cyclops Cave, Protaras;
CG - Cape Greco;
Ch - Chelones (Panayia Apakou);
CKo-CKo - Cape Kormakitis - Cap Koruçam;
CKi - Cape Kiti;
CP - Cape Pyla;
Da-Ka - Daylos - Kaplica;
Dh - Dhekelia (Larnaka);
Ep - Episkopi;
ESW - "Elpida" Shipwreck;
Fa - Famagusta;

GB - Governor's Beach;
 G-K - Galinoporni - Kaleburnu;
 JSP - Jubilee Shoals Pissuri;
 KaC - Kakoskali cave;
 K-G - Keryneia - Girne;
 K-K - Kormakitis - Koruçam;
 KY-Ku - Komatou Yalou - Kumyalı;
 KB - Konnos Bay;
 La - Larnaca;
 Lar - Lara;
 Lat - Latsi;
 Lim - Limassol;
 LM - Ladies Mile;
 Lio - Liopetri;
 Li - Sa - Livera - Sadrazamköy;
 Li - Ye - Limnitis - Yeşilırmak;
 MB-GB - Morphou Bay - Güzelyurt Bay;
 MPS - Moni Power Station;
 NC - North Coast;
 N-T - Neta - Taslica;
 N - Nisia;
 O-K - Orga - Kayalar;
 Or - Ormidia;
 Pa - Paphos;
 Par - Paralimni;
 Pe - Pernera;
 PP - Petunda Point;
 Po-Ye - Potamos-Yedidalga;
 Pro - Protaras;
 PL - Psarolimano, Larnaca;
 Ri-Di - Rizokarpaso - Dipkarpaz;
 Ri-Di SC - Rizokarpaso - Dipkarpaz, South Coast;
 Ri-KP - Rizokarpaso - Karpaz Peninsula;
 Sa - Salamis;
 SC - South Coast;
 So - Sotira;
 Sof - Softades;
 SPL - 1 km south of Potamos, Liopetri;
 Un - Unspecified;
 Va - Vasilikos;
 Xy - Xylophagou;
 Y-Y - Yialousa - Yenierenkoy;
 Zy - Zygi

Symbols

! - As drawing;
 ‡ - Dubious identification;
 † - From synonym;
 † (after scientific name) - Fossil;
 ✕ - Misidentification;
 * - No image documentation;
 ? - Questionable placement

The present work includes 51 families of Bivalvia with 212 species, 7 families of Cephalopoda with 23 species, 127 families of Gastropoda with 611 species, 6 families of Polyplacophora with 15 species, and 4 families of Scaphopoda with 10 species. In total, 195 families and 871 species are catalogued and supported by some 90 references. It adds 10 more families and 116 more species for the area in comparison with the previous taxa register (Kolokotronis *et al.* 2022), and raises the marine malacofauna of Cyprus by some 15% and the corresponding families by some 5%.

A review of the catalogue reveals the complete absence or low representation of Bivalve families belonging to the Order Nuculanida such as those of Malletiidae and Neilonellidae, Pristiglomidae and Yoldiidae that include small and/or deep-water species. The situation of poor representation is also similar for the Gastropod family of Skeneidae which is internationally represented by very small to minute species, hence our overall opinion that the Seas of Cyprus are under searched, and our, under confirmation, anticipation that more are yet to come by means of persistent search and appropriate collaborations. As before (Kolokotronis *et al.* 2022), *Dendostrea* cfr. *folium* (Linnaeus, 1758), *Spondylus* cfr. *spinosus* Schreibers, 1793 and *Viriola* cfr. *bayani* Jousseaume, 1884 are included in the catalogue because their identity can only be confirmed by DNA markers analysis. The establishment status of the alien or Lessepsian species in the area changes with each new relevant publication and we anticipate that this catalogue will be in use for some time, therefore, readers are invited to form their own opinion on the matter by taking into account the available literature of the time and the criteria employed by Zenetos *et al.*, (2005) and particularly defined by Crocetta (2012). We hope that in due time we will be again in the position to reupdate this catalogue to catch up with the ever-enriching malacofauna of the area due to more incomers and the finding of as yet unknown local residents.

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Catalogue of species

BIVALVIA	Publications	Locality
SOLEMYIDAE Gray, 1840		
<i>Solemya togata</i> (Poli, 1791)	76*, 84, 87	Xy
NUCULIDAE J. E. Gray, 1824		
<i>Austronucula perminima</i> (Monterosato, 1875)	27	KaC
<i>Ennucula tenuis</i> (Montagu, 1808)	76†*	Un
<i>Nucula nitidosa</i> Winckworth, 1930	12*, 27*, 67, 76*	KaC, Sof
<i>Nucula nucleus</i> (Linnaeus, 1758)	76*, 84	Xy
<i>Nucula sulcata</i> Bronn, 1831	76*, 84	Xy
NUCULANIDAE H. Adams & A. Adams, 1858 (1854)		
<i>Lembulus pella</i> (Linnaeus, 1758)	76†*, 84†, 87	AN, Or
<i>Saccella commutata</i> (R. A. Philippi, 1844)	76*, 84†	AN
ARCIDAE Lamarck, 1809		
<i>Acar clathrata</i> (Defrance, 1816)	27*	KaC
<i>Acar plicata</i> (Dillwyn, 1817)	76*	Un
<i>Anadara corbuloides</i> (Monterosato, 1881)	76*, 84	Un
<i>Anadara gibbosa</i> (Reeve, 1844)	76†*, 84†	Un
<i>Arca noae</i> Linnaeus, 1758	12*, 27*, 75, 76*, 84	KaC
<i>Barbatia barbata</i> (Linnaeus, 1758)	2, 12*, 27*, 75, 76*, 84	Akr, AN, KaC
<i>Bathyarca penctunculoides</i> (Scacchi, 1835)	76*, 89	Or
<i>Tetrarca tetragona</i> (Poli, 1795)	75†, 76†*, 84†	Un
NOETIIDAE Stewart, 1930		
<i>Striarca lactea</i> (Linnaeus, 1758)	12*, 27*, 75, 76*, 84	KaC, Lim, Pa
GLYCYMERIDIDAE Dall, 1908		
<i>Glycymeris bimaculata</i> (Poli, 1795)	75†, 76*, 84†	Un
<i>Glycymeris glycymeris</i> (Linnaeus, 1758)	12*, 27*, 76*, 84	KaC, La, LM, Pa
<i>Glycymeris nummaria</i> (Linnaeus, 1758)	12†*, 75†, 76†*, 84†	Fa, MB-GB
<i>Glycymeris pilosa</i> (Linnaeus, 1767)	84	Un
<i>Glycymeris stellata</i> (Bruguière, 1789)	84*	AN, LM
MYTILIDAE Rafinesque, 1815		
<i>Brachidontes pharaonis</i> (P. Fischer, 1870)	3, 12*, 23, 33*, 62*, 76*	CKo-CKo
<i>Brachidontes variabilis</i> (Krauss, 1848)	75, 84 ‡	Un
<i>Crenella arenaria</i> Monterosato, 1875 ex H. Martin, ms.	75a*, 76*, 77	JSP, KaC
<i>Dacrydium hyalinum</i> (Monterosato, 1875)	77	JSP
<i>Gregariella petagnae</i> (Scacchi, 1832)	76*, 84	Un
<i>Gregariella semigranata</i> (Reeve, 1858)	77	Par
<i>Lioberus ligneus</i> (Reeve, 1858)	1, 65	Lio,Ri-KP
<i>Lithophaga lithophaga</i> (Linnaeus, 1758)	27*, 76*, 84	KaC, Lim, Xy
<i>Modiolula phaseolina</i> (R. A. Philippi, 1844)	76*, 84‡as <i>G. barbatella</i> , 87	Un
<i>Modiolus adriaticus</i> Lamarck, 1819	76*, 84, 88	La, Lim, Xy
<i>Modiolus barbatus</i> (Linnaeus, 1758)	12*, 76*, 84	Xy
<i>Musculus costulatus</i> (Risso, 1826)	12*, 27*, 76*, 84	KaC, Xy
<i>Musculus subpictus</i> (Cantraine, 1835)	12†*, 76†*	Un
<i>Mytilaster minimus</i> (Poli, 1795)	12*, 76*	Un
<i>Mytilus edulis</i> Linnaeus, 1758	12*, 75,	Lim, Xy

<i>Mytilus galloprovincialis</i> Lamarck, 1819	75, 76*, 84†	Xy
<i>Septifer cumingii</i> Récluz, 1849	3, 23, 27*, 62†*, 92	AE-Ça, CKo-CKo, Da-Ka, KaC
PINNIDAE Leach, 1819		
<i>Pinna nobilis</i> Linnaeus, 1758	12*, 75, 76*, 84	LM
<i>Pinna rudis</i> Linnaeus, 1758	94*	AGP, ESW
MARGARITIDAE Blainville, 1824		
<i>Pinctada albina</i> (Lamarck, 1819)	53‡,	Fa, Xy
<i>Pinctada radiata</i> (Leach, 1814)	3†, 12†*, 23†, 27†*, 33†*, 62†*, 75†, 76†*, 84†	AP-Di, KaC
PTERIIDAE Gray, 1847 (1820)		
<i>Pteria hirundo</i> (Linnaeus, 1758)	76*, 84	Un
MALLEIDAE Lamarck 1818		
<i>Malleus regula</i> (Forsskål in Niebuhr, 1775)	3, 10†, 12†*, 23, 62†*, 76*, 84*	A-T, K-G, La
ISOGNOMONIDAE Woodring, 1925		
<i>Isognomon australicus</i> (Reeve, 1858)	1†	AA-Es
<i>Isognomon bicolor</i> (C. B. Adams, 1845)	2, 87	AT
<i>Isognomon legumen</i> (Gmelin, 1791)	2, 65	Lar, Lio
PROPEAMUSSIIDAE Abbott, 1954		
<i>Similipecten similis</i> (Laskey, 1811)	76*	Un
PECTINIDAE Rafinesque, 1815		
<i>Aequipecten opercularis</i> (Linnaeus, 1758)	75†, 76*, 84*	Un
<i>Flexopecten flexuosus</i> (Poli, 1795)	76*	Un
<i>Flexopecten glaber</i> (Linnaeus, 1758)	75†, 76*, 84†	Un
<i>Flexopecten hyalinus</i> (Poli, 1795)	12†*, 27*, 75†, 76†*, 84†	KaC
<i>Karnekampia sulcata</i> (O. F. Müller, 1776)	75†	
<i>Manupecten pesfelis</i> (Linnaeus, 1758)	76*, 84†	Un
<i>Mimachlamys varia</i> (Linnaeus, 1758)	75†, 76*, 84†	Un
<i>Palliolium incomparabile</i> (Risso, 1826)	12*, 76*	Un
<i>Pecten jacobaeus</i> (Linnaeus, 1758)	75†, 75, 76*, 84	Un
<i>Perapecten commutatus</i> (Monterosato, 1875)	75†	
<i>Pseudamussium clavatum</i> (Poli, 1795)	76*, 84†	Un
<i>Pseudamussium peslutrae</i> (Linnaeus, 1771)	76†*, 84*	Un
<i>Talochlamys multistriata</i> (Poli, 1795)	76†*, 84†	Un
<i>Talochlamys pusio</i> (Linnaeus, 1758)	84*	Un
SPONDYLIDAE J. E. Gray, 1826		
<i>Spondylus gaederopus</i> Linnaeus, 1758	12*, 27*, 75, 76*, 84	KaC
<i>Spondylus</i> cfr. <i>spinosus</i> Schreibers, 1793	62*, 89, 92	Lim, Zy
ANOMIIDAE Rafinesque, 1815		
<i>Anomia ephippium</i> Linnaeus, 1758	12*, 27*, 75, 76*, 84	KaC
LIMIDAE d'Orbigny, 1846		
<i>Lima lima</i> (Linnaeus, 1758)	12*, 27*, 75, 76*, 84	AN, KaC, Xy
<i>Limaria hians</i> (Gmelin, 1791)	12*, 75†, 76*, 84‡	LM
<i>Limaria loscombi</i> (G. B. Sowerby I, 1823)	76	Un
<i>Limaria tuberculata</i> (Olivi, 1792)	12†*, 76*, 84‡ as <i>L. hians</i>	LM
<i>Limatula gwyni</i> (Sykes, 1903)	27*, 76*, 77	JSP, KaC
<i>Limatula subauriculata</i> (Montagu, 1808)	76*, 84†	Pro
<i>Limea crassa</i> (Forbes, 1844)	77	JSP

OSTREIDAE Rafinesque, 1815		
<i>Dendostrea</i> cfr. <i>folium</i> (Linnaeus, 1758)	3, 65	Lio
<i>Dendostrea frons</i> (Linnaeus, 1758)	23, 62*, 92	A-T, La
<i>Magallana gigas</i> (Thunberg, 1793)	66	Xy
<i>Ostrea edulis</i> Linnaeus, 1758	27*, 75, 76*, 84	KaC, La, Xy
<i>Ostrea stentina</i> Payraudeau, 1826	76†*	Un
GRYPHAEIDAE Vyalov, 1936		
<i>Neopycnodonte cochlear</i> (Poli, 1795)	76*, 84†	La
LUCINIDAE Fleming, 1828		
<i>Ctena decussata</i> (O. G. Costa, 1829)	12*, 27*, 76*, 84, 89	KaC, Lio, Xy
<i>Divaricella ornatissima</i> (d'Orbigny, 1846)	76†*	Un
<i>Loripes orbiculatus</i> Poli, 1791	12†*, 76†*, 84†	Xy
<i>Loripinus fragilis</i> (R. A. Philippi, 1836)	27*, 76†*, 84, 89	KaC, Xy
<i>Lucinella divaricata</i> (Linnaeus, 1758)	12*, 27*, 76*, 84*, 77	KaC, Par, Xy
<i>Megaxinus transversus</i> (Bronn, 1831) †?	12*, 76†*	Un
<i>Megaxinus unguiculinus</i> Pallary, 1904	27*, 67	KaC, Par
<i>Myrtea spinifera</i> (Montagu, 1803)	76*, 84	PL
THYASIRIDAE Dall, 1900 (1895)		
<i>Axinulus croulinensis</i> (Jeffreys, 1847)	76*	Un
<i>Axinulus cycladius</i> (S. V. Wood, 1848) <i>taxon inquirendum</i>	76†*	Lim, Pa, Xy
<i>Mendicula ferruginosa</i> (Forbes, 1844)	27*	KaC
<i>Thyasira biplicata</i> (R. A. Philippi, 1836)	76*	Un
<i>Thyasira flexuosa</i> (Montagu, 1803)	84†	AN, Xy
<i>Thyasira marwicki</i> Amano, C. Little, K. A. Campbell, R. G. Jenkins & Saether, 2015 †	76*	Un
<i>Thyasira subovata</i> (Jeffreys, 1881)	76†*	Un
UNGULINIDAE Gray, 1854		
<i>Microstagon trigonum</i> (Scacchi, 1835)	12†*, 76†*	Un
CHAMIDAE Lamarck, 1809		
<i>Chama asperella</i> Lamarck, 1819	23, 78*, 93	N-T, NCy
<i>Chama gryphoides</i> Linnaeus, 1758	12*, 27*, 76*, 84	KaC, La
<i>Chama pacifica</i> Broderip, 1835	3, 23, 62*	AP-Di
<i>Pseudochama corbieri</i> (Jonas, 1846)	76*	Un
<i>Pseudochama gryphina</i> (Lamarck, 1819)	12*, 27*, 75, 76*, 53, 84	KaC, La
LASAEIDAE Gray, 1842		
<i>Bornia sebetia</i> (O. G. Costa, 1830)	12*, 76*, 84, 87	Pa, SPL, Xy
<i>Tellimya ferruginosa</i> (Montagu, 1808)	76*	Un
CARDITIDAE Férussac, 1822		
<i>Cardita calyculata</i> (Linnaeus, 1758)	12*, 27*, 75, 76*, 84	KaC, Xy
<i>Cardites antiquatus</i> (Linnaeus, 1758)	12†*, 27*, 76*, 84†	AN, KaC
<i>Centrocardita aculeata</i> (Poli, 1795)	76†*, 84†, 89	AN, Xy
<i>Centrocardita squamigera</i> (Deshayes, 1832)	84†	
<i>Coripia corbis</i> (R. A. Philippi, 1836)	12†*, 66, 76†*, 84†	CG, Lio
<i>Coripia jozinae</i> (vanAartsen, 1985)	12†*, 76†*	Un
<i>Glans trapezia</i> (Linnaeus, 1767)	12*, 27*, 75, 76*, 53!, 84†	AN, KaC, Xy
<i>Pleuromeris marshalli</i> (Marwick, 1924)	84†*	CG
ASTARTIDAE d'Orbigny, 1844 (1840)		
<i>Astarte sulcata</i> (da Costa, 1778)	76*, 84*	Un
<i>Laevastarte fusca fusca</i> (Poli, 1795)	76†*, 84†, 87†	Zy

CARDIIDAE Lamarck, 1809		
<i>Acanthocardia aculeata</i> (Linnaeus, 1758)	75, 76*, 84	Un
<i>Acanthocardia echinata</i> (Linnaeus, 1758)	12*, 76*, 84	Un
<i>Acanthocardia paucicostata</i> (G. B. Sowerby II, 1834)	76*, 84	Un
<i>Acanthocardia tuberculata</i> (Linnaeus, 1758)	12*, 75, 76*, 84	Un
<i>Cerastoderma edule</i> (Linnaeus, 1758)	12*, 76*, 84	La, Lim, Xy
<i>Cerastoderma glaucum</i> (Bruguère, 1789)	76*, 84†*	Xy
<i>Fulvia fragilis</i> (Forsskål in Niebuhr, 1775)	62*, 76*, 92	Y-Y
<i>Laevicardium crassum</i> (Gmelin, 1791)	76*, 84†	GB
<i>Laevicardium oblongum</i> (Gmelin, 1791)	76*, 84	GB
<i>Papillicardium papillosum</i> (Poli, 1791)	12†*, 75†, 76†*, 84†	Xy
<i>Parvicardium carrozzai</i> van Aartsen & Goud, 2001	77	JSP
<i>Parvicardium exiguum</i> (Gmelin, 1791)	12*, 76*, 84†, 87	AN, Xy
<i>Parvicardium pinnulatum</i> (Conrad, 1831)	12†*, 76†*, 84†*	Xy
<i>Parvicardium scabrum</i> (R. A. Philippi, 1844)	76*	Un
<i>Parvicardium scriptum</i> (Bucquoy, Dautzenberg & Dollfus, 1892)	12*, 16, 27*, 76*, 87	ATS, Fa, KaC
<i>Parvicardium trapezium</i> Cecalupo & Quadri, 1996	12, 16, 76*	K-G
MACTRIDAE Lamarck, 1809		
<i>Mactra corallina</i> (Linnaeus, 1758)	84	Un
<i>Mactra stultorum</i> (Linnaeus, 1758)	12*, 75, 76*, 84	La, MB-GB
MESODESMATIDAE Gray, 1840		
<i>Donacilla cornea</i> (Poli, 1791)	76*, 84	Fa
SOLENIIDAE Lamarck, 1809		
<i>Solen marginatus</i> Pulteney, 1799	75, 76*, 84	Fa, Mo
PHARIDAE H. Adams & A. Adams, 1856		
<i>Ensis ensis</i> (Linnaeus, 1758)	75, 76*, 84	La, LM
<i>Ensis magnus</i> Schumacher, 1817	76†*	Un
<i>Ensis minor</i> (Chenu, 1843)	12*, 76*	Un
<i>Ensis siliqua</i> (Linnaeus, 1758)	75, 84	La, LM
<i>Phaxas pellucidus</i> (Pennant, 1777)	76*	Un
TELLINIDAE Blainville, 1814		
<i>Arcopella balaustina</i> (Linnaeus, 1758)	12†*, 67, 76†*, 84†	Par, Xy
<i>Asbjornsenia pygmaea</i> (Lovén, 1846)	76†*	Un
<i>Bosemprella incarnata</i> (Linnaeus, 1758)	75†, 76†*, 84†	AN, LM
<i>Fabulina fabula</i> (Gmelin, 1791)	65, 76†*, 84†	LM, Ri-Di
<i>Gastrana fragilis</i> (Linnaeus, 1758)	76*, 84	Lim, Xy
<i>Macoma balthica</i> (Linnaeus, 1758) cfr.	76*	Un
<i>Macomangulus tenuis</i> (da Costa, 1778)	75†, 32†*, 84†	LM
<i>Moerella distorta</i> (Poli, 1791)	76†*, 89	Zy
<i>Moerella donacina</i> (Linnaeus, 1758)	12†*, 76†*, 84†	LM
<i>Moerella pulchella</i> (Lamarck, 1818)	76†*, 84†	Fa
<i>Oudardia compressa</i> (Brocchi, 1814)	12†*, 76†*	Un
<i>Peronaea planata</i> (Linnaeus, 1758)	75†, 76†*, 84†	Lim
<i>Peronidia albicans</i> (Gmelin, 1791)	12†*, 76†*, 84†	AN, LM
<i>Psammacoma gubernaculum</i> (Hanley, 1844)	62†*, 92	La
<i>Serratina serrata</i> (Brocchi, 1814)	76†*	Un
DONACIDAE J. Fleming, 1828		
<i>Donax semistriatus</i> Poli, 1795	12*, 75, 76*, 84	Un

<i>Donax trunculus</i> Linnaeus, 1758	75, 76*, 84	Un
<i>Donax venustus</i> Poli, 1795	12*, 75, 76*, 84	LM
PSAMMOBIIDAE J. Fleming, 1828		
<i>Gari depressa</i> (Pennant, 1777)	76*, 84†	Fa
SEMELIDAE Stoliczka, 1870		
<i>Abra alba</i> (W. Wood, 1802)	27*, 76*	KaC
<i>Abra longicallus</i> (Scacchi, 1835)	76*	Un
<i>Abra nitida</i> (O. F. Müller, 1776)	12*, 76*	Un
<i>Abra prismatica</i> (Montagu, 1808)	76*	Un
<i>Abra segmentum</i> (Récluz, 1843)	76*	Un
<i>Abra tenuis</i> (Montagu, 1803)	76*, 84	Xy
SOLECURTIDAE d'Orbigny, 1846		
<i>Azorinus chamasolen</i> (da Costa, 1778)	76*	Un
<i>Solecurtus strigilatus</i> (Linnaeus, 1758)	75, 76*, 84	La, LM
KELLIELLIDAE P. Fischer, 1887		
<i>Kelliella miliaris</i> (R. A. Philippi, 1844)	76†	Un
TRAPEZIDAE Lamy, 1920 (1895)		
<i>Coralliophaga lithophagella</i> (Lamarck, 1819)	27*	KaC
GLOSSIDAE J. E. Gray, 1847		
<i>Glossus humanus</i> (Linnaeus, 1758)	75, 76†*, 84	Pa
VENERIDAE Rafinesque, 1815		
<i>Callista chione</i> (Linnaeus, 1758)	75, 76*, 84	Xy
<i>Chamelea gallina</i> (Linnaeus, 1758)	12*, 75, 76*, 84‡	Un
<i>Chamelea striatula</i> (da Costa, 1778)	84‡as <i>C. gallina</i>	
<i>Clausinella fasciata</i> (da Costa, 1778)	75, 76*, 84	Un
<i>Dosinia exoleta</i> (Linnaeus, 1758)	27*, 75, 76*, 84	KaC, LM, Xy
<i>Dosinia lupinus</i> (Linnaeus, 1758)	12*, 37, 75, 76*, 84	LM
<i>Gafrarium pectinatum</i> (Linnaeus, 1758)	62*, 65, 92‡	Lio, O-K
<i>Gafrarium savignyi</i> (Jonas, 1846)	66, 92	Lio, O-K
<i>Globivenus effossa</i> (R. A. Philippi, 1836)	76*	Un
<i>Gouldia minima</i> (Montagu, 1803)	27*, 76*, 84	KaC, LM
<i>Irus irus</i> (Linnaeus, 1758)	12*, 75, 76*, 84	AN, Pa
<i>Lajonkairia lajonkairii</i> (Payraudeau, 1826)	12†*, 27*, 65, 84	An, KaC, Lio, Pa, Xy
<i>Lajonkairia substriata</i> (Montagu, 1808)	76†*, 84	An, Pa, Xy
<i>Mysia undata</i> (Pennant, 1777)	76*, 84*	Pa, Xy
<i>Paratapes textile</i> (Gmelin, 1791)	62†*, 92	Li-Sa
<i>Petricola lithophaga</i> (Retzius, 1788)	76*, 84	La, Lim, Xy
<i>Pitar rudis</i> (Poli, 1795)	76*, 84‡, 87, 77	Lio, Pro
<i>Polititapes aureus</i> (Gmelin, 1791)	75†, 76†*, 53, 84†	Lim, Xy
<i>Polititapes lucens</i> (Locard, 1886)	89	Xy
<i>Polititapes rhomboides</i> (Pennant, 1777)	76†*, 84†	Un
<i>Ruditapes decussatus</i> (Linnaeus, 1758)	75†, 76*, 84†	AN, Lim
<i>Timoclea ovata</i> (Pennant, 1777)	76*, 84*	Fa
<i>Venerupis corrugata</i> (Gmelin, 1791)	12†*, 75†, 76†*, 84†	MPS, Xy
<i>Venerupis geographica</i> (Gmelin, 1791)	84	Lim, MPS, Xy
<i>Venus casina</i> Linnaeus, 1758	12*, 75, 76*, 84	Un
<i>Venus verrucosa</i> Linnaeus, 1758	27*, 75, 76*, 84	KaC, LM

CORBULIDAE Lamarck, 1818		
<i>Varicorbula gibba</i> (Olivi, 1792)	12 ⁺ *, 27*, 76 ⁺ *, 84 ⁺	KaC, Xy
GASTROCHAENIDAE Gray, 1840		
<i>Rocellaria dubia</i> (Pennant, 1777)	76 ⁺ *	Xy
HIATELLIDAE J. E. Gray, 1824		
<i>Hiatella arctica</i> (Linnaeus, 1767)	12*, 76 ⁺ *, 84, 87	La, Lio
PHOLADIDAE Lamarck, 1809		
<i>Barnea candida</i> (Linnaeus, 1758)	76*, 84	Xy
<i>Pholas dactylus</i> Linnaeus, 1758	76*, 84	Un
TEREDINIDAE Rafinesque, 1815		
<i>Lyrodus pedicellatus</i> (Quatrefages, 1849)	76*	Un
<i>Nototeredo norvagica</i> (Spengler, 1792)	76*	Un
<i>Teredo navalis</i> Linnaeus, 1758	12*, 76*, 84 ⁺	Un
<i>Teredo utriculus</i> Gmelin, 1791 <i>nomen dubium</i>	12*, 76*, 84 ⁺	Un
THRACIIDAE Stoliczka, 1870 (1839)		
<i>Thracia phaseolina</i> (Lamarck, 1818)	12 ⁺ *, 76 ⁺ *, 84 ⁺ , 87	CG, LM
<i>Thracia pubescens</i> (Pulteney, 1799)	76*, 84	GB
CLAVAGELLIDAE d'Orbigny, 1843		
<i>Bryopa aperta</i> (G. B. Sowerby I, 1823)	84 ⁺ * as <i>B. melitensis</i>	Xy
<i>Bryopa melitensis</i> (Broderip, 1834)	76 ⁺ *, 84 ⁺	Xy
PANDORIDAE Rafinesque, 1815		
<i>Pandora inaequalis</i> (Linnaeus, 1758)	76*, 84, 87	LM
LYONSIIDAE P. Fischer, 1887		
<i>Lyonsia norvegica</i> (Gmelin, 1791)	76*	Un
CUSPIDARIIDAE Dall, 1886		
<i>Cardiomya costellata</i> (Deshayes, 1835)	76*, 84*, 87	PL
<i>Cuspidaria cuspidata</i> (Olivi, 1792)	76*, 84	Un
<i>Cuspidaria microrrhina</i> Dall, 1886	76 ⁺ *, 84 ⁺ *	Un
VERTICORDIIDAE Stoliczka, 1870		
<i>Haliris trapezoidea</i> (G. Seguenza, 1876)	16 ⁺	Un
CEPHALOPODA		
SEPIIDAE Leach, 1817		
<i>Rhomboscion elegans</i> (Blainville, 1827)	26 ⁺ *, 76 ⁺ *, 84 ⁺	Co
<i>Rhomboscion orbignyanum</i> (A. Férussac, 1826)	76 ⁺ *	Un
<i>Sepia officinalis</i> Linnaeus, 1758	12*, 26*, 75, 76*, 84	Co
SEPIOLIDAE Leach, 1817		
<i>Euprymna scolopes</i> S. S. Berry, 1913	48	Pro
<i>Sepietta neglecta</i> Naef, 1916	76*	Un
<i>Sepietta oweniana</i> (d'Orbigny, 1841)	76*	Un
<i>Sepiola robusta</i> Naef, 1912	76*	Un
<i>Sepiola rondeletii</i> Leach, 1817	26*, 76*	Co
<i>Sepiola steenstrupiana</i> Levy, 1912	76*	Un
LOLIGINIDAE Lesueur, 1821		
<i>Alloteuthis media</i> (Linnaeus, 1758)	76*	Un
<i>Loligo forbesii</i> Steenstrup, 1856	76*	Un

<i>Loligo vulgaris</i> Lamarck, 1798	26*, 76*, 84	Co
<i>Sepioteuthis lessoniana</i> d'Orbigny, 1826	66, 86	Or, Pa
OMMASTREPHIDAE Steenstrup, 1857		
<i>Illex coindetii</i> (Vérany, 1839)	76*	Un
<i>Todarodes sagittatus</i> (Lamarck, 1798)	26†*, 76*	Co
<i>Todaropsis eblanae</i> (Ball, 1841)	76*	Un
ELEDONIDAE Rochebrune, 1884		
<i>Eledone moschata</i> (Lamarck, 1798)	76*, 84*	Un
OCTOPODIDAE d'Orbigny, 1840		
<i>Callistoctopus macropus</i> (Risso, 1826)	26†*, 49, 76*, 84*	Co
<i>Macrotritopus defilippi</i> (Vérany, 1851)	18	AN
<i>Octopus salutii</i> Vérany, 1836	76*	Un
<i>Octopus vulgaris</i> Cuvier, 1797	12*, 26†*, 75, 76*, 84	Co
<i>Scaevurgus uniccirrhus</i> (Delle Chiaje [in Férussac & d'Orbigny], 1841)	76*	Un
ARGONAUTIDAE Cantraine, 1841		
<i>Argonauta argo</i> Linnaeus, 1758	12*, 26*, 75, 76*, 84	Ra, Un
GASTROPODA		
NACELLIDAE Thiele, 1891		
<i>Cellana radiata</i> (Born, 1778)	75	Un
PATELLIDAE Rafinesque, 1815		
<i>Patella aspera</i> Röding, 1798	84	Un
<i>Patella caerulea</i> Linnaeus, 1758	12*, 26†*, 27*, 74, 75, 76*, 84	AN, KaC, Pa
<i>Patella rustica</i> Linnaeus, 1758	12*, 75, 76*, 84†	Un
<i>Patella ulyssiponensis</i> Gmelin, 1791	12*, 75, 76*	Un
LOTTIIDAE Gray, 1840		
<i>Tectura virginea</i> (O. F. Müller, 1776)	12*, 26†*, 27*, 76*, 84	KaC, O-K
LEPETIDAE Gray, 1850		
<i>Propilidium exiguum</i> (W. Thompson, 1844)	76†*	Un
SEGUENZIOIDEA Verrill, 1884		
<i>Akritogyra conspicua</i> (Monterosato, 1880)	27*	KaC
FISSURELLIDAE Fleming, 1822		
<i>Diodora dorsata</i> (Monterosato, 1878)	12*, 76*, 87	Lio
<i>Diodora gibberula</i> (Lamarck, 1822)	12*, 27*, 73, 76*, 84	KaC
<i>Diodora graeca</i> (Linnaeus, 1758)	76*, 84, 87	Xy
<i>Diodora italica</i> (Defrance, 1820)	26*, 37, 76*, 84	AN, Fa, MB-GB, NC, Xy
<i>Diodora producta</i> (Monterosato, 1880)	65	Ri-Di
<i>Emarginula adriatica</i> O. G. Costa, 1830	27*, 65	KaC, Lio
<i>Emarginula crebrisculpta</i> Coen, 1939 uncertain	12*	Un
<i>Emarginula huzardii</i> Payraudeau, 1826	12†*, 27*, 76*, 84, 87	AN, KaC, Xy
<i>Emarginula octaviana</i> Coen, 1939	12*, 26†*, 27*, 76*, 84†	AN, Fa, KaC
<i>Emarginula punctulum</i> Piani, 1980	87	Lio
<i>Emarginula rosea</i> Bell, 1824		
<i>Emarginula sicala</i> J. E. Gray, 1825	12*, 26†*, 27*, 75, 76*, 84	AN, KaC, Pro
<i>Fissurella nubecula</i> (Linnaeus, 1758)	12*, 75, 76*, 84	Pa, Xy

SCISSURELLIDAE Gray, 1847		
<i>Anatoma aspera</i> (R. A. Philippi, 1844)	89	Or
<i>Anatoma crispata</i> (J. Fleming, 1828)	76*	Un
<i>Scissurella costata</i> d'Orbigny, 1824	12*, 27*, 76*, 77	JSP, KaC
<i>Sinezona cingulata</i> (O. G. Costa, 1861)	27*, 29*†, 76*	Lim, KaC
<i>Sinezona semicostata</i> Burnay & Rolán, 1990	77	JSP
HALIOTIDAE Rafinesque, 1815		
<i>Haliotis tuberculata</i> Linnaeus, 1758	12*, 26+*, 27*, 75, 76*, 84	KaC
ADDISONIIDAE Dall, 1882		
<i>Addisonia excentrica</i> (Tiberi, 1855)	76+*, 84‡	Un
TROCHIDAE Rafinesque, 1815		
<i>Clanculus corallinus</i> (Gmelin, 1791)	12*, 26+*, 27*, 76*, 84	AN, Fa, KaC, NC, Pa, Xy
<i>Clanculus cruciatus</i> (Linnaeus, 1758)	12*, 27*, 75, 76*, 84	KaC, Pa, Xy
<i>Clanculus jussieui</i> (Payraudeau, 1826)	12*, 27*, 73, 76*, 84	KaC, Pa, Xy
<i>Clelandella miliaris</i> (Brocchi, 1814)	76*	Un
<i>Gibbula ardens</i> (Salis Marschlin, 1793)	12*, 27*, 75, 76*, 84	KaC, Pa, Xy
<i>Gibbula fanulum</i> (Gmelin, 1791)	27*, 75, 76*, 84	KaC, Xy
<i>Gibbula guttadauri</i> (R. A. Philippi, 1836)	76*, 84	La
<i>Gibbula magus</i> (Linnaeus, 1758)	76*, 84	Un
<i>Gibbula philberti</i> (Récluz, 1843)	12*, 27*, 76*	KaC, Sof
<i>Gibbula turbinoides</i> (Deshayes, 1835)	12*, 27*, 76*, 84†, 87	KaC, Lio, Pa
<i>Jujubinus corallinus</i> Monterosato, 1884 <i>taxon inquirendum</i>	84	Un
<i>Jujubinus exasperatus</i> (Pennant, 1777)	12*, 26+*, 27*, 75, 76*, 84	KaC, Pa, Xy
<i>Jujubinus gravinae</i> (Dautzenberg, 1881)	27*	KaC
<i>Jujubinus striatus</i> (Linnaeus, 1758)	27*, 75, 76*, 84	KaC, Pa, Xy
<i>Phorcus articulatus</i> (Lamarck, 1822)	12†*, 26†*, 75†, 76†*, 84†	Un
<i>Phorcus mutabilis</i> (R. A. Philippi, 1851)	59, 76†*, 84†	Lim
<i>Phorcus richardi</i> (Payraudeau, 1826)	12†*, 76†*, 84†	Lim, Pa
<i>Phorcus turbinatus</i> (Born, 1778)	12†*, 26†*, 75†, 76†*, 84†	Co
<i>Steromphala adansonii</i> (Payraudeau, 1826)	75†, 76†*, 84†	Co
<i>Steromphala albida</i> (Gmelin, 1791)	27*, 87	KaC, Lio
<i>Steromphala divaricata</i> (Linnaeus, 1758)	12†*, 75†, 76†*, 84†	Lim, Xy
<i>Steromphala leucophaea</i> (R. A. Philippi, 1836)	76†*, 84†	Xy
<i>Steromphala nebulosa</i> (R. A. Philippi, 1839)	12†*, 76†*, 84†, 87	Lio, Xy
<i>Steromphala racketti</i> (Payraudeau, 1826)	87	Xy
<i>Steromphala rarilineata</i> (Michaud, 1829)	12†*, 75†, 76†*, 84†	Lim, Pa
<i>Steromphala umbilicaris</i> (Linnaeus, 1758)	12†*, 27*, 77	KaC, Lio
<i>Steromphala varia</i> (Linnaeus, 1758)	12†*, 26†*, 76†*, 84†	Xy
<i>Trochus erithreus</i> Brocchi, 1821	16, 58, 62*, 70, 76*, 84	La
CALLIOSTOMATIDAE Thiele, 1924		
<i>Calliostoma conulus</i> (Linnaeus, 1758)	12*, 27*, 75, 76*, 84	KaC
<i>Calliostoma granulatum</i> (Born, 1778)	26†*, 27*, 76*, 84	KaC, SC
<i>Calliostoma laugeri</i> (Payraudeau, 1826)	12*, 14, 26†*, 27*, 32c, 75, 76*, 77†, 84	AI-Ak, JSP, KaC, Lim, Xy

<i>Calliostoma zizyphinum</i> (Linnaeus, 1758)	12 ⁺ *, 26 ⁺ *, 76*, 84	Fa
SKENEIDAE W. Clark, 1851		
<i>Skeneoides exilissima</i> (R. A. Philippi, 1844)	27*	KaC
COLLONIIDAE Cossmann, 1917		
<i>Cantrainea peloritana</i> (Cantraine, 1835)	76*	Un
<i>Glabracollonia carmelae</i> (Oliverio & Buzzurro, 1994)	16, 76*, 72, 75 ⁺ , 84 ⁺ as sp.	La, Or
<i>Homalopoma sanguineum</i> (Linnaeus, 1758)	12*, 26 ⁺ *, 27*, 75, 84	AN, KaC, Pa, Pro, Sa, Xy
TURBINIDAE Rafinesque, 1815		
<i>Bolma rugosa</i> (Linnaeus, 1767)	12*, 26 ⁺ *, 27*, 75, 76*, 84 ⁺	KaC, La, Pa
PHASIANELLIDAE Swainson, 1840		
<i>Tricolia pullus</i> (Linnaeus, 1758)	12*, 26 ⁺ *, 27*, 75, 76*, 84	AN, KaC, Pa, Xy
<i>Tricolia speciosa</i> (Megerle von Mühlfeld, 1824)	12*, 26 ⁺ *, 27*, 75, 76*, 84	KaC, Pa, Xy
<i>Tricolia tenuis</i> (Michaud, 1829)	12*, 75, 76*, 84 ⁺	AN, Pa, Xy
NERITIDAE Rafinesque, 1815		
<i>Smaragdia souverbiana</i> (Montrouzier, 1863)	62*, 76*	Un
<i>Smaragdia viridis</i> (Linnaeus, 1758)	12*, 26 ⁺ *, 27*, 75, 76*, 84	AN, KaC, Pa, Pro, Xy
CERITHIIDAE Fleming, 1822		
<i>Bittium latreillii</i> (Payraudeau, 1826)	12*, 27*, 75, 76*, 84	KaC
<i>Bittium reticulatum</i> (da Costa, 1778)	11 ⁺ , 12, 26 ⁺ *, 27*, 75, 76*, 84	KaC, K-G
<i>Cerithidium diplax</i> (R. B. Watson, 1886)	11 ⁺ , 12, 76 ⁺ *	K-G
<i>Cerithidium perparvulum</i> (R. B. Watson, 1886)	16, 62*	Un
<i>Cerithidium submammillatum</i> (De Rayneval & Ponzi, 1854)	27*	KaC
<i>Cerithium alucastrum</i> (Brocchi, 1814)	76*, 84*	Xy
<i>Cerithium lividulum</i> Risso, 1826	27*, 76*, 89	KaC, Lio
<i>Cerithium nesioticum</i> Pilsbry & Vanatta, 1906	16, 31, 62*, 76*	AN
<i>Cerithium protractum</i> Bivona Ant., 1838	16, 31, 76 ⁺ *	Un
<i>Cerithium renovatum</i> Monterosato, 1884	27*, 65, 89	KaC, Lio, Xy
<i>Cerithium scabridum</i> R. A. Philippi, 1848	3, 12*, 23, 27*, 62*, 76 ⁺ *, 80	AP-Di, KaC, Lim
<i>Cerithium vulgatum</i> Bruguière, 1792	12*, 26 ⁺ *, 27*, 31, 75, 76*, 84	Fa, KaC, Pa
<i>Rhinoclavis kochi</i> (R. A. Philippi, 1848)	12*, 23, 62*, 75, 76*, 84 ⁺	AN, N-T
SCALIOLIDAE Jousseume, 1912		
<i>Finella pupoides</i> A. Adams, 1860	12*, 62*, 76*	Un
PLANAXIDAE Gray, 1850		
<i>Fossarus ambiguus</i> (Linnaeus, 1758)	12*, 76 ⁺ *, 87	SPL
POTAMIDIDAE H. Adams & A. Adams, 1854		
<i>Pirenella conica</i> (Blainville, 1829)	76*, 84	Xy
SILICULARIIDAE Anton, 1838		
<i>Petalopoma elisabettae</i> Schiaparelli, 2002	27*, 77	JSP, KaC
<i>Tenagodus obtusus</i> (Schumacher, 1817)	76*, 84	Un
TURRITELLIDAE Lovén, 1847		
<i>Archimediella triplicata</i> (Brocchi, 1814) ⁺ ?	26 ⁺ *, 84 ⁺	Ep, NC
<i>Turritellinella tricarinata</i> (Brocchi, 1814)	26 ⁺ *, 75 ⁺ , 76 ⁺ *, 83 ⁺ , 84 ⁺	Ep, Fa, La, Lim, Pa, Xy

<i>Turritella turbona</i> Monterosato, 1877	12*, 27*, 75†, 76*, 84†	KaC
TRIPHORIDAE Gray, 1847		
<i>Cheirodonta pallescens</i> (Jeffreys, 1867)	12*, 76*	Un
<i>Euthymella colzumensis</i> (Jousseaume, 1898)	2	Ri-Di
<i>Marshallora adversa</i> (Montagu, 1803)	12*, 27*, 76*, 87	KaC, Xy
<i>Metaxia bacillum</i> (Issel, 1869)	11, 12*, 30, 62*, 76*	Un
<i>Metaxia metaxa</i> (Delle Chiaje, 1828)	76*	Un
<i>Monophorus perversus</i> (Linnaeus, 1758)	12*, 26†*, 27*, 75, 76*, 83†, 84††	KaC, MB-GB, NC, Pa, Pro, Xy
<i>Similiphora similior</i> (Bouchet & Guillemot, 1978)	12*, 27*, 76*	KaC
<i>Viriola</i> cfr. <i>bayani</i> Jousseaume, 1884	65, 87	CCP, Lio
CERITHIOPSIDAE H. Adams & A. Adams, 1853		
<i>Cerithiopsis diadema</i> Monterosato, 1874	76*	Un
<i>Cerithiopsis fayalensis</i> R. B. Watson, 1880	12*, 76*	Un
<i>Cerithiopsis minima</i> (Brusina, 1865)	12*, 76*	Un
<i>Cerithiopsis pulvis</i> (Issel, 1869)	12*, 23, 62*, 76*	CKo-CKo
<i>Cerithiopsis scalaris</i> Locard, 1891	76	Un
<i>Cerithiopsis tenthrenois</i> (Melvill, 1896)	62*, 76*	Un
<i>Cerithiopsis tubercularis</i> (Montagu, 1803)	12*, 27*, 76†*, 84†	KaC
<i>Costulopsis nana</i> Jeffreys, 1867	12†*, 76†*	Un
<i>Dizoniopsis bilineata</i> (M. Hörnes, 1848) †?	12*, 76*	Un
<i>Dizoniopsis coppolae</i> (Aradas, 1870)	12*, 27*, 76*	KaC, Lio
<i>Joculator problematicus</i> Albano & Steger, 2021	2	KB
<i>Krachia tiara</i> (Monterosato, 1874)	12†*, 76*, 77	JSP
<i>Seila trilineata</i> (R. A. Philippi, 1836)	76*, 84, 88	Lim, Pa
EPITONIIDAE Berry, 1910		
<i>Cycloscala hyalina</i> (G. B. Sowerby II, 1844)	10, 12*, 62*, 76*, 87	K-G, Lio
<i>Epitonium algerianum</i> (Weinkauff, 1866)	12*, 76*	Un
<i>Epitonium clathratulum</i> (Kanmacher, 1798)	75†, 76*, 84†	Xy
<i>Epitonium clathrus</i> (Linnaeus, 1758)	12†*, 76†*, 84†	Un
<i>Epitonium turtonis</i> (W. Turton, 1819)	12†*, 75, 76†*, 84†	La, LM
<i>Gyroscala commutata</i> (Monterosato, 1877)	12†*, 26†*, 75†, 76†*, 83†, 84†	AN, Ch, Lim, MB-GB, Xy
<i>Janthina exigua</i> Lamarck, 1816	16, 75	Un
<i>Janthina globosa</i> Swainson, 1822	12†*, 26†*, 76†*, 83†	Fa, MB-GB
<i>Janthina janthina</i> (Linnaeus, 1758)	12*, 26†*, 27*, 75, 76*, 83†, 84	Fa, KaC, MB-GB
<i>Janthina pallida</i> W. Thompson, 1840	89	Akr
<i>Opalia coronata</i> (R. A. Philippi & Scacchi, 1840)	12†*, 76†*	Un
EULIMIDAE R. A. Philippi, 1853		
<i>Crinophtheiros comatulicula</i> (Graff, 1875)	76*	Un
<i>Curveulima devians</i> (Monterosato, 1884)	76†*	Un
<i>Ersilia mediterranea</i> (Monterosato, 1869)	27*, 76*, 77	JSP, KaC
<i>Eulima bilineata</i> Alder, 1848	76*	Un
<i>Eulima ephamilla</i> R. B. Watson, 1883	84‡	Un
<i>Eulima glabra</i> (da Costa, 1778)	65, 26†*, 76†*, 83, 84†	Lio, NC
<i>Hemiliostraca clandestina</i> (Mifsud & Ovalis, 2019)	2	Akr
<i>Melanella boscii</i> (Payraudeau, 1826)	12*, 76*, 84‡ as <i>E. ephamilla</i> , 89	Akr, Pa, Xy

<i>Melanella frielei</i> (H. K. Jordan, 1895)	67	Lio
<i>Melanella levantina</i> (Oliverio, Buzzurro & Villa, 1994)	12†*, 76†*, 84‡as <i>E. distorta</i> , 77	JSP, Pa, Xy
<i>Melanella monterosatoi</i> (Monterosato, 1890)	12*, 76*	Un
<i>Melanella polita</i> (Linnaeus, 1758)	65, 75, 76*	Lio
<i>Melanella translucens</i> (Monterosato, 1890)	88	Lio, Xy
<i>Nanobalcis nana</i> (Monterosato, 1878)	76*	Un
<i>Parvioris ibizenca</i> (F. Nordsieck, 1968)	12†*, 27*, 67, 76*	KaC, So
<i>Sabinella bonifaciae</i> (F. Nordsieck, 1974)	76†*	Un
<i>Sticteulima jeffreysiana</i> (Brusina, 1869)	12*, 27*, 76*, 77	JSP, KaC
<i>Sticteulima lentiginosa</i> (A. Adams, 1861)	62*, 76*	Un
<i>Vitreolina antiflexa</i> (Monterosato, 1884)	27*,	KaC
<i>Vitreolina arcuata</i> (C. B. Adams, 1850)	26†*	NC
<i>Vitreolina curva</i> (Monterosato, 1874)	76*	Un
<i>Vitreolina incurva</i> (Bucquoy, Dautzenberg & Dollfus, 1883)	12*, 76*	Un
<i>Vitreolina perminima</i> (Jeffreys, 1883)	76*	Un
<i>Vitreolina philippi</i> (de Rayneval & Ponzi, 1854)	22, 27*, 76*, 84‡	KaC
LITTORINIDAE Children, 1834		
<i>Echinolittorina punctata</i> (Gmelin, 1791)	76†, 89	A-T
<i>Melarhaphe neritoides</i> (Linnaeus, 1758)	12†*, 26†*, 75†, 76*, 83†, 84†	AN, Pa, Pro, Xy
CINGULOPSIDAE Fretter & Patil, 1958		
<i>Eatonina fulgida</i> (J. Adams, 1797)	12*, 27*, 76†*	KaC
<i>Eatonina pumila</i> (Monterosato, 1884)	87	SPL
RISSOIDAE Gray, 1847		
<i>Alvania amatii</i> Oliverio, 1986	31, 76*	Fa
<i>Alvania aspera</i> (R. A. Philippi, 1844) cfr.	12*, 76*, 84, 89	Akr
<i>Alvania beanii</i> (Hanley in Thorpe, 1844)	27*, 76*, 84†, 77	JSP, KaC, Pa
<i>Alvania cancellata</i> (da Costa, 1778)	76*, 84‡as <i>A. hirta</i>	Un
<i>Alvania cimex</i> (Linnaeus, 1758)	26*, 75, 76*, 83, 84	AN, Fa, NC, Pa
<i>Alvania clarae</i> Nofroni & Pizzini, 1991	16,	
<i>Alvania colossophilus</i> Oberling, 1970	12*, 16, 27*, 31, 76*, 88	AN, KaC
<i>Alvania datchaensis</i> Amati & Oliverio, 1987	12*, 27*, 76*, 69	KaC
<i>Alvania discors</i> (T. Brown, 1818)	12*, 76*, 84†, 89	AN, Xy
<i>Alvania dorbignyi</i> (Audouin, 1826)	8, 12*, 62*, 76*	Un
<i>Alvania fractospira</i> Oberling, 1970	27*, 31, 76*	KaC
<i>Alvania geryonia</i> (Nardo, 1847)	67, 76*	Li
<i>Alvania hallgassi</i> Amati & Oliverio, 1985	12*, 76*	Un
<i>Alvania hirta</i> (Monterosato, 1884)	84‡	Un
<i>Alvania hispidula</i> (Monterosato, 1884)	76*	Un
<i>Alvania lactea</i> (Michaud, 1830)	76*	Un
<i>Alvania lanciae</i> (Calcara, 1845)	12*, 76*, 84	Pa, Xy
<i>Alvania lineata</i> Risso, 1826	12*, 27*, 76*, 84	KaC, Pa, Xy
<i>Alvania lucinae</i> Oberling, 1970	89	Xy
<i>Alvania mamillata</i> Risso, 1826	12*, 27*, 67, 76*	KaC, Lio
<i>Alvania marmarisensis</i> Bitlis & Öztürk, 2017	77	JSP
<i>Alvania monterosatoi</i> (P. Fischer, 1877) †?	12*	Un
<i>Alvania oliverioi</i> Buzzurro, 2003	16	Un
<i>Alvania peloritana</i> Aradas & Benoit, 1874	88	Lio

<i>Alvania rominae</i> Amati, Trono & Oliverio, 2020	89	Lio
<i>Alvania rudis</i> (R. A. Philippi, 1844)	12*, 76*	Un
<i>Alvania scabra</i> (R. A. Philippi, 1844)	12*, 27*, 76*	KaC
<i>Alvania sculptilis</i> (Monterosato, 1877)	12*, 76*	Un
<i>Alvania subcrenulata</i> (Bucquoy, Dautzenberg & Dollfus, 1884)	27*, 88, 87	Li-Ye, Lio, KaC
<i>Alvania testae</i> (Aradas & Maggiore, 1844)	76	Un
<i>Alvania weinkauffi</i> Weinkauff, 1868	12†*, 76†*	Un
<i>Cingula trifasciata</i> (J. Adams, 1800)	76*, 84†	Xy
<i>Crisilla beniamina</i> (Monterosato, 1884)	11, 12†*, 76†*	K-G
<i>Crisilla chiarellii</i> (Cecalupo & Quadri, 1995)	11, 12†*, 31, 76†*	K-G
<i>Manzonina crassa</i> (Kannmacher, 1798)	12*, 27*, 76*, 77	JSP, KaC
<i>Obtusella intersecta</i> (S. V. Wood, 1857)	77	JSP
<i>Obtusella macilenta</i> (Monterosato, 1880)	12*, 27*, 76*	KaC
<i>Peringiella elegans</i> (Locard, 1891)	12*, 76*	Un
<i>Pusillina inconspicua</i> (Alder, 1844)	12*, 27*, 76*	KaC
<i>Pusillina lineolata</i> (Michaud, 1830)	12*, 67, 76*, 84†	Lio, Pa
<i>Pusillina marginata</i> (Michaud, 1830)	12*, 27*, 76*, 77	JSP, KaC
<i>Pusillina munda</i> (Monterosato, 1884)	12*, 27*, 76*	KaC
<i>Pusillina philippi</i> (Aradas & Maggiore, 1844)	12*, 27*, 31, 76*	AN, KaC
<i>Pusillina radiata</i> (R. A. Philippi, 1836)	12*, 27*, 76*, 89	KaC, Xy
<i>Rissoa angustior</i> (Monterosato, 1917) uncertain > taxon inquirendum	12*, 16, 31, 76*	K-G
<i>Rissoa auriformis</i> Pallary, 1904	12*, 16, 31, 76*	Un
<i>Rissoa auriscalpium</i> (Linnaeus, 1758)	27*, 76*, 84	AN, KaC, Pa
<i>Rissoa decorata</i> R. A. Philippi, 1846	12*, 76*	Un
<i>Rissoa guerinii</i> Récluz, 1843	12*, 76*, 84, 87	AN, CCP, Pa
<i>Rissoa italiensis</i> Verduin, 1985	12*, 76*	Un
<i>Rissoa lia</i> (Monterosato, 1884)	12*, 27*, 76*, 84†‡	KaC, Pa, Xy
<i>Rissoa membranacea</i> (J. Adams, 1800)	12†*, 76†*, 89	Akr
<i>Rissoa monodonta</i> R. A. Philippi, 1836	12*, 76*, 84	Xy
<i>Rissoa parva</i> (da Costa, 1778)	27*	KaC
<i>Rissoa rodhensis</i> Verduin, 1985	12*, 27*, 31, 36, 76*	KaC, K-G
<i>Rissoa scurra</i> (Monterosato, 1917)	12*, 27*, 76*, 87, 89	Akr, KaC, Lio
<i>Rissoa similis</i> Scacchi, 1836	12*, 27*, 76*, 84†, 87	KaC, SPL, Xy
<i>Rissoa splendida</i> Eichwald, 1830	76	Un
<i>Rissoa variabilis</i> (Megerle von Mühlfeld, 1824)	12*, 27*, 76*, 84, 87	AN, KaC, Lio,
<i>Rissoa ventricosa</i> Desmarest, 1814	12*, 76*, 83, 84	Xy
<i>Rissoa violacea</i> Desmarest, 1814	12*, 76*, 84, 87	SPL
<i>Setia amabilis</i> (Locard, 1886)	89	Akr
<i>Setia ambigua</i> (Brugnone, 1873)	12*, 76*	Un
<i>Setia fusca</i> (R. A. Philippi, 1841)	12*, 76*	Un
<i>Setia slikorum</i> (Verduin, 1984)	12*, 76*	Un
<i>Setia turriculata</i> Monterosato, 1884	27*, 76*, 87	KaC, SPL
RISSOINIDAE Stimpson, 1865		
<i>Rissoina bertholleti</i> Issel, 1869	62*, 65, 76*, 87	AT, Lio
<i>Rissoina bruguieri</i> (Payraudeau, 1826)	12*, 27*, 76*, 84	KaC, Xy
ANABATHRIDAE Keen, 1971		
<i>Pisinnia glabrata</i> (Megerle von Mühlfeld, 1824)	76*	Un

ASSIMINEIDAE H. Adams & A. Adams, 1856		
<i>Paludinella sicana</i> (Brugnone, 1876)	12*, 76*	Un
BARLEEIIDAE Gray, 1857		
<i>Barleeia unifasciata</i> (Montagu, 1803)	27*, 76*, 84†	KaC, Pa, Xy
CAECIDAE Gray, 1850		
<i>Caecum armoricum</i> de Folin, 1869	27*	KaC
<i>Caecum auriculatum</i> de Folin, 1868	12*, 27*, 76*, 77	JSP, KaC
<i>Caecum clarkii</i> P. P. Carpenter, 1859	27*, 77	JSP, KaC
<i>Caecum subannulatum</i> de Folin, 1870	27*, 77	JSP, KaC
<i>Caecum trachea</i> (Montagu, 1803)	12*, 27*, 76*, 77	JSP, KaC
HYDROBIIDAE Stimpson, 1865		
<i>Hydrobia acuta</i> (Draparnaud, 1805)	76	Un
TATEIDAE Thiele, 1925		
<i>Potamopyrgus antipodarum</i> (J. E. Gray, 1843)	84†*	Un
TORNIDAE Sacco, 1896		
<i>Tornus subcarinatus</i> (Montagu, 1803)	12*, 27*, 76*, 84, 87	KaC, SPL, Xy
VITRINELLIDAE Bush, 1897		
<i>Circulus striatus</i> (R. A. Philippi, 1836)	76*	Un
TRUNCATELLIDAE Gray, 1840		
<i>Truncatella subcylindrica</i> (Linnaeus, 1767)	12*, 26*, 27*, 75, 76*, 84	AN, KaC, MB-GB, NC, Pa, Xy
VERMETIDAE Rafinesque, 1815		
<i>Dendropoma anguliferum</i> (Monterosato, 1878)	12*, 27*, 76*	KaC
<i>Dendropoma cristatum</i> (Biondi-Giunti, 1859)	12†*, 76†*, 84†	Un
<i>Petalconchus glomeratus</i> (Linnaeus, 1758)	12*, 26†*, 76*, 83†	Fa
<i>Thylacodes arenarius</i> (Linnaeus, 1758)	76†*, 83†, 84†	Fa
<i>Thylaeodus rugulosus</i> (Monterosato, 1878)	12†, 27*, 76†*	KaC, K-G
<i>Thylaeodus semisurrectus</i> (Bivona e Bernardi, 1832) cfr.	76†*	Un
<i>Vermetus granulatus</i> (Gravenhorst, 1831) cfr.	76*, 84	Un
<i>Vermetus triquetrus</i> Bivona e Bernardi, 1832	12*, 26†*, 76*, 83†	Un
STROMBIDAE Rafinesque, 1815		
<i>Conomurex decorus</i> (Röding, 1798)	10†, 75†, 84†	AA-Es
<i>Conomurex persicus</i> (Swainson, 1821)	3, 10†, 12*, 16, 23, 26†*, 28, 31, 33*, 62†*, 76†*, 84†	AA-Es, Aka, AN, CP, KaC, Y-Y
APORRHAIIDAE Gray, 1850		
<i>Aporrhais pespelecani</i> (Linnaeus, 1758)	26†*, 75, 76*, 83, 84	Fa, La, MB-GB
CALYPTRAEIDAE Lamarck, 1809		
<i>Calyptrea chinensis</i> (Linnaeus, 1758)	26†*	KaC
<i>Crepidula unguiformis</i> Lamarck, 1822	76*, 84, 87	Lim, LM
CAPULIDAE Fleming, 1822		
<i>Capulus ungaricus</i> (Linnaeus, 1758)	26†*, 76*, 84, 87	Akr, Al-Ak
VELUTINIDAE Gray, 1840		
<i>Lamellaria latens</i> (O. F. Müller, 1776)	76*, 84	Lim
<i>Marsenia perspicua</i> (Linnaeus, 1758)	12†*, 26†*, 76†*, 84†, 89	KaC, Lio, LM, Xy
TRIVIIDAE Troschel, 1863		
<i>Trivia arctica</i> (Pulteney, 1799)	76*	Un
<i>Trivia candidula</i> (Gaskoin, 1836)		

<i>Trivia levantina</i> Smriglio, Mariottini & Buzzurro, 1998	67, 76*	So
<i>Trivia mediterranea</i> (Risso, 1826)	12†*, 26*, 75†, 76†*, 83, 84†	AN, Fa, NC, Pa
<i>Trivia monacha</i> (da Costa, 1778)	75, 84‡	AN, Pa
CYPRAEIDAE Rafinesque, 1815		
<i>Erronea caurica</i> (Linnaeus, 1758)	31, 75†,	AN, Lim
<i>Luria lurida</i> (Linnaeus, 1758)	12*, 26†*, 26†*, 37†, 75†, 76*, 83†, 84†	Aka, AN, Fa, KaC, NC, XY
<i>Naria spurca</i> (Linnaeus, 1758)	12†*, 26*, 26†*, 37†, 75†, 76†*, 83†, 84†	Fa, KaC, NC
<i>Purpuradusta gracilis notata</i> (Gill, 1858)	62*, 75†, 92	Xy
<i>Zonaria pyrum</i> (Gmelin, 1791)	75†	
NATICIDAE Guilding, 1834		
<i>Euspira guilleminii</i> (Payraudeau, 1826)	12*, 26†*, 76*, 89	Lim, Zy
<i>Euspira nitida</i> (Donovan, 1804)	12*, 26†*, 76†*, 83†, 84†	Pro
<i>Naticarius hebraeus</i> (Martyn, 1786)	12†*, 26†*, 75†, 76†*, 83†, 84	Lim, LM, MB-GB, Pa, Xy
<i>Naticarius stercusmuscarum</i> (Gmelin, 1791)	12†*, 26†*, 31, 44, 75†, 76†* 83†, 84†	CA, Fa, Lim, LM, Po
<i>Neverita josephina</i> Risso, 1826	12*, 26†*, 75†, 76*, 83†, 84	AI-Ak, AN, Fa
<i>Notocochlis dillwynii</i> (Payraudeau, 1826)	12†*, 27*, 75†, 76†*, 84†	AN, KaC, Xy
<i>Payraudeautia intricata</i> (Donovan, 1804)	12†*, 52†, 75†, 76†*, 84†	Lim, Pro
<i>Tectonatica rizzae</i> (R. A. Philippi, 1844)	12†*, 76*, 84†	Un
<i>Tectonatica sagraiana</i> (A. d'Orbigny, 1842)	76†*, 84†	La
TONNIDAE Suter, 1913		
<i>Tonna galea</i> (Linnaeus, 1758)	12*, 26†*, 65, 75, 76*, 83, 84	Fa, Lio, SC,
CYMATIIDAE Iredale, 1913		
<i>Monoplex corrugatus</i> (Lamarck, 1816)	26†*, 75†, 76†*, 83†, 84†	Bo
<i>Monoplex parthenopeus</i> (Salis Marschlin, 1793)	76†*, 84†	Un
RANELLIDAE Gray, 1854		
<i>Cabestana cutacea</i> (Linnaeus, 1767)	76*, 84†	CG
<i>Charonia lampas</i> (Linnaeus, 1758)	16, 75, 76*, 84†	Un
<i>Charonia sequezae</i> (Aradas & Benoit, 1871)	84	Aka
<i>Charonia variegata</i> (Lamarck, 1816)	12†*, 26*, 31, 75, 76*, 83	NC
<i>Ranella olearium</i> (Linnaeus, 1758)	75, 76†*, 84†	La
CASSIDAE Latreille, 1825		
<i>Galeodea echinophora</i> (Linnaeus, 1758)	26†*, 75†, 76*, 83†, 84†	Fa, SC
<i>Galeodea rugosa</i> (Linnaeus, 1771)	76*, 84†*	Un
<i>Semicassis granulata</i> (Born, 1778)	12†*, 26†*, 75†, 76†*, 83†, 84†	KaC
<i>Semicassis saburon</i> (Bruguère, 1792)	26†*, 76†*	Fa, MB-GB, NC
PTEROTRACHEIDAE Rafinesque, 1814		
<i>Firoloida desmarestia</i> Lesueur, 1817	76*	Un
<i>Pterotrachea hippocampus</i> R. A. Philippi, 1836	13, 76*	Un
ATLANTIDAE Rang, 1829		
<i>Atlanta brunnea</i> J. E. Gray, 1850	27*	KaC
<i>Atlanta peronii</i> Lesueur, 1817	27*, 76*	KaC
<i>Atlanta selvagensis</i> de Vera & Seapy, 2006	27*	KaC

<i>Proatlanta souleyeti</i> (E. A. Smith, 1888)	27*, 76, 77	JSP, KaC
MURICIDAE Rafinesque, 1815		
<i>Aspella anceps</i> (Lamarck, 1822)	12*, 76*, 84	Un
<i>Babelomurex benoiti</i> (Tiberi, 1855)	75	Un
<i>Babelomurex cariniferus</i> (G. B. Sowerby II, 1834)	76†*, 84†	Xy
<i>Bolinus brandaris</i> (Linnaeus, 1758)	26†*, 75†, 76*, 83†, 84†	Al-Ak, Fa, La
<i>Coralliophila meyendorffii</i> (Calcara, 1845)	12*, 27*, 75, 76*, 84	KaC
<i>Dermomurex scalaroides</i> (Blainville, 1829)	12*, 76*, 87, 77	JSP, PL
<i>Ergalatax junionae</i> Houart, 2008	3, 21, 23, 62*	AN, Bo, G-K, La, Pro, A-T
<i>Ergalatax martensi</i> (Schepman, 1892)	76†*	Un
<i>Gracilipurpura craticulata</i> (Bucquoy & Dautzenberg, 1882)	76†*, 84†	Un
<i>Hexaplex trunculus</i> (Linnaeus, 1758)	12*, 26†*, 27*, 75†, 76†*, 83†, 84†	Al-Ak, AN, Fa, KaC
<i>Hirtomurex squamosus</i> (Bivona e Bernardi, 1838)	75†, 76†*, 84†	Un
<i>Indothais lacera</i> (Born, 1778)	62†*, 76†*	Un
<i>Murexsul aradasii</i> (Monterosato, 1883)	12*, 75†, 76†*, 84†	Xy
<i>Murexsul cevikeri</i> (Houart, 2000)	76*	Un
<i>Muricopsis cristata</i> (Brocchi, 1814)	12*, 26†*, 27*, 75, 76*, 83†, 84	Fa, KaC, NC
<i>Ocenebra baetica</i> (Reeve, 1845)	12†*, 76†*	Un
<i>Ocenebra edwardsii</i> (Payraudeau, 1826)	12†*, 75†, 76†*, 84†	Lim, Pa
<i>Ocenebra erinaceus</i> (Linnaeus, 1758)	75, 76*, 84†	An, Xy
<i>Ocenebra hispidula</i> (Pallary, 1904)	12†*, 76†*	Un
<i>Ocenebrina aciculata</i> (Lamarck, 1822)	75, 76*, 83†, 84*	AN, Pro
<i>Ocenebrina aegeensis</i> Aissaoui, Barco & Oliverio, 2017	87	Or
<i>Stramonita haemastoma</i> (Linnaeus, 1767)	12*, 26†*, 75†, 76*, 83†, 84†	Al-Ak, La
<i>Typhinellus labiatus</i> (de Cristofori & Jan, 1832)	12†*, 27*, 76†*, 84†	KaC, La
GRANULINIDAE G. A. Covert & H. K. Covert, 1995		
<i>Granulina boucheti</i> Gofas, 1992	76	Un
<i>Granulina marginata</i> (Bivona e Bernardi, 1832)	12*, 26†*, 76*, 84†*, 87	KaC, SPL
<i>Granulina occulta</i> (Monterosato, 1869)	12*, 76*	Un
MARGINELLIDAE Fleming, 1828		
<i>Volvarina mitrella</i> (Risso, 1826)	12*, 26†*, 75, 76†*, 84†	AN, KaC, Pa
CYSTISCIDAE Stimpson, 1865		
<i>Gibberula miliaria</i> (Linnaeus, 1758)	12*, 26†*, 76*, 84‡	AN, Fa, NC
<i>Gibberula philippii</i> (Monterosato, 1878)	12*, 26†*, 76*, 84‡ as <i>G. miliaria</i> , 87	KaC, Lio
MITRIDAE Swainson, 1829		
<i>Episcomitra cornicula</i> (Linnaeus, 1758)	12†*, 26*, 27*, 75†, 76†*, 83†, 84†	AN, Co, KaC, Pa, Xy
<i>Episcomitra zonata</i> (Marryat, 1818)	75†, 76†*, 84†	AN
COSTELLARIIDAE MacDonald, 1860		
<i>Pusia ebenus</i> (Lamarck, 1811)	12†*, 26*, 27*, 75†, 76†*, 83†, 84†	KaC, NC, Pe
<i>Pusia granum</i> (Forbes, 1844)	12†, 27*, 75†, 76†*, 84†	AN, KaC, K-G, Pa, Xy
<i>Pusia savignyi</i> (Payraudeau, 1826)	76†*, 84*	Un
<i>Pusia tricolor</i> (Gmelin, 1791)	12†*, 76†*, 83, 84	AN, Pa, Xy
<i>Vexillum hypatiae</i> (Pallary, 1912)	12	K-G

CHAUVETIIDAE Kantor, Fedosov, Kosyan, Puillandre, Sorokin, Kano, R. Clark & Bouchet, 2021		
<i>Chauvetia affinis</i> (Monterosato, 1889)	27*, 88	Lim, KaC
<i>Chauvetia brunnea</i> (Donovan, 1804)	65	Lio
<i>Chauvetia candidissima</i> (R. A. Philippi, 1836)	76*	Un
<i>Chauvetia recondita</i> (Brugnone, 1873)	12*, 76†*	Un
<i>Chauvetia turritellata</i> (Deshayes, 1835)	27*	KaC
TUDICLIDAE Cossmann, 1901		
<i>Euthria cornea</i> (Linnaeus, 1758)	12†*, 26*, 27*, 75†, 76†*, 83†, 84	CG, Fa, KaC, MB-GB, NC, Pa
COLUBRARIIDAE Dall, 1904		
<i>Cumia intertexta</i> (Helbling, 1779)	12†*, 27†*, 53†, 75†, 76†*, 84†	AGP, AN, KaC, Xy
NASSARIIDAE Iredale, 1916		
<i>Nassarius circumcinctus</i> (A. Adams, 1852)	76	Un
<i>Nassarius conoidalis</i> (Deshayes, 1833)	83†‡	
<i>Tritia corniculum</i> (Olivi, 1792)	12†*, 26*, 75†, 76†*, 84†	Fa, Lim
<i>Tritia corrugata</i> (Brocchi, 1814)	12†*, 65†, 75†, 76†*, 84†	Lim, Lio, MPS
<i>Tritia gibbosula</i> (Linnaeus, 1758)	12†*, 16, 26†*, 73, 75†, 76†*, 83†, 84†	Fa, MB-GB, Pa
<i>Tritia incrassata</i> (Strøm, 1768)	12†*, 75†, 76†*, 83†, 84†	Pa, Xy
<i>Tritia lima</i> (Dillwyn, 1817)	75†, 76†*, 84†*	Xy
<i>Tritia lousi</i> (Pallary, 1912)	12†*, 16, 57, 76†*, 88	Lim
<i>Tritia mutabilis</i> (Linnaeus, 1758)	12†*, 26*, 75†, 76†*, 83†, 84†	Un
<i>Tritia neritea</i> (Linnaeus, 1758)	12†*, 26*, 75†, 76†*, 83†, 84†	Fa, Pa, Xy
<i>Tritia pellucida</i> (Risso, 1826)	84†	Xy
<i>Tritia reticulata</i> (Linnaeus, 1758)	12†*, 26*, 76†*, 83†, 84†	Fa, Pa, Xy
<i>Tritia turulosa</i> (Risso, 1826)	16	
PISANIIDAE Gray, 1857		
<i>Aplus coccineus</i> (Monterosato, 1884)	76†*, 88, 84†	An, Xy
<i>Aplus dorbignyi</i> (Payraudeau, 1826)	12†*, 26†*, 27*, 75†, 76†*, 83†, 84†	KaC
<i>Aplus scacchianus</i> (R. A. Philippi, 1844)	12†*, 27*, 75†, 76†*, 84†	AN, KaC
<i>Enginella leucozona</i> (R. A. Philippi, 1844)	12†*, 27*, 75†, 76†*, 84†	KaC
<i>Pisania striata</i> (Gmelin, 1791)	12*, 26†*, 27*, 75, 76*, 83†, 84†	KaC
COLUMBELLIDAE Swainson, 1840		
<i>Columbella rustica</i> (Linnaeus, 1758)	12*, 26†*, 27*, 75, 76*, 83, 84	KaC
<i>Mitrella bruggeni</i> van Aartsen, Menkhorst & E. Gittenberger, 1984	76*	Un
<i>Mitrella coccinea</i> (R. A. Philippi, 1836)	27*	KaC
<i>Mitrella gervillii</i> (Payraudeau, 1826)	12*, 76*, 84†	Pa
<i>Mitrella minor</i> (Scacchi, 1836)	12*, 76*	Un
<i>Mitrella scripta</i> (Linnaeus, 1758)	12*, 26†*, 27*, 75, 76*, 83†, 84†	KaC, NC
<i>Mitrella svelta</i> Kobelt, 1889	27*	KaC
<i>Zafra savignyi</i> (Moazzo, 1939)	12†*, 33†*, 62*	Un
<i>Zafra selasphora</i> (Melvill & Standen, 1901)	33†*, 62*	Un

FASCIOLARIIDAE Gray, 1853		
<i>Aptyxis syracusana</i> (Linnaeus, 1758)	12 ⁺ *, 26*, 75 ⁺ , 76 ⁺ *, 83 ⁺ , 84 ⁺	Xy
<i>Fusinus rudis</i> (R. A. Philippi, 1844) †?	12*, 76 ⁺ *	AN, Xy
<i>Pseudofusus patriciae</i> (Russo & Olivieri, 2013)	88	Or
<i>Pseudofusus pulchellus</i> (R. A. Philippi, 1840)	75 ⁺ , 84 ⁺	AN, Pa, Xy
<i>Pseudofusus rostratus</i> (Olivi, 1792)	12 ⁺ *, 26 ⁺ *, 75 ⁺ , 76 ⁺ *, 83 ⁺	Un
<i>Tarantinaea lignaria</i> (Linnaeus, 1758)	12 ⁺ *, 26*, 75 ⁺ , 76 ⁺ *, 83 ⁺ , 84 ⁺	NC
CANCELLARIIDAE Forbes & Hanley, 1851		
<i>Bivetiella cancellata</i> (Linnaeus, 1767)	76 ⁺ *, 84 ⁺	Un
CLATHURELLIDAE H. Adams & A. Adams, 1858		
<i>Pleurotomoides gracilis</i> (Montagu, 1803)	75 ⁺ ‡, 76 ⁺ *, 87	AN
CONIDAE Fleming, 1822		
<i>Conus ventricosus</i> Gmelin, 1791	12 ⁺ *, 26 ⁺ *, 27*, 75, 76 ⁺ *, 83 ⁺ , 84 ⁺	Co, KaC
DRILLIIDAE Olsson, 1964		
<i>Crassopleura maravignae</i> (Bivona, 1838)	76 ⁺ *, 84 ⁺ *, 88	La
HORAICLAVIDAE Bouchet, Kantor, Sysoev & Puillandre, 2011		
<i>Haedropleura secalina</i> (R. A. Philippi, 1844)	12*	Un
<i>Haedropleura septangularis</i> (Montagu, 1803)	27*, 76*, 84 ⁺ ‡	KaC
MANGELIIDAE P. Fischer, 1883		
<i>Bela cycladensis</i> (Reeve, 1845) cfr.	76*	Un
<i>Bela menkhorsti</i> van Aartsen, 1988	12*, 76*	Un
<i>Bela minoica</i> Bogi, Giannuzzi Savelli & Pusateri, 2021	6, 65	AN, Lio
<i>Bela nebula</i> (Montagu, 1803)	12*, 76*	Un
<i>Bela ornata</i> (Locard, 1891)	12*, 75, 76*	Un
<i>Bela plicatilis</i> (Risso, 1826) cfr.	6, 89	Akr
<i>Bela taprurenensis</i> (Pallary, 1904)	76*	Un
<i>Bela zonata</i> (Locard, 1891)	12 ⁺ *, 76 ⁺ *	Un
<i>Lyromangelia taeniata</i> (Deshayes, 1835)	12 ⁺ *, 27 ⁺ *, 76 ⁺ *, 82 ⁺ , 83 ⁺	KaC
<i>Mangelia callosa</i> (F. Nordsieck, 1977)	12 ⁺ *, 13 ⁺ a, 65, 76 ⁺ *, 82	AN, Lim, Lio
<i>Mangelia pallaryi</i> (F. Nordsieck, 1977) cfr.	76*, 77	Sof
<i>Mangelia sandrii</i> (Brusina, 1865)	12 ⁺ *, 76*	Un
<i>Mangelia striolata</i> Risso, 1826	12 ⁺ *, 76 ⁺ *, 84 ⁺ , 89	Pro, Xy
<i>Mangelia thapsiae</i> (Oberling, 1970)	12 ⁺ , 16, 27*, 76 ⁺ *, 82	KaC, K-G
<i>Mangelia unifasciata</i> (Deshayes, 1835)	12 ⁺ *, 26 ⁺ *, 76*, 83 ⁺ , 84 ⁺ ‡	Fa
<i>Pseudomangelia sicula</i> (Reeve, 1846)	12 ⁺ *, 76 ⁺ *, 89	Xy
<i>Pseudomangelia vauquelini</i> (Payraudeau, 1826)	12 ⁺ *, 76 ⁺ *, 84 ⁺ , 89	Lio, Xy
<i>Smithiella costulata</i> (Risso, 1826)	12 ⁺ *, 76 ⁺ *	Un
<i>Villiersiella attenuata</i> (Montagu, 1803)	76 ⁺ *, 87	Lio
MITROMORPHIDAE T. L. Casey, 1904		
<i>Mitromorpha columbellaria</i> (Scacchi, 1836)	27*	KaC
<i>Mitromorpha karpathensis</i> (F. Nordsieck, 1969)	27*	KaC
<i>Mitromorpha olivoidea</i> (Cantraine, 1835)	12 ⁺ *, 67, 76 ⁺ *, 89	Li, Pro

RAPHITOMIDAE Bellardi, 1875		
<i>Clathromangelia granum</i> (R. A. Philippi, 1844)	27*, 65, 76*, 84*	AN, KaC, Lio, Xy
<i>Clathromangelia quadrillum</i> (Dujardin, 1837) †?	12*	Un
<i>Cyrellia linearis</i> (Montagu, 1803)	76†*, 84†	Un
<i>Gymnobela subaraneosa</i> (Dautzenberg & H. Fischer, 1896)	76	Un
<i>Leufroyia concinna</i> (Scacchi, 1836)	76†*	Un
<i>Leufroyia leufroyi</i> (Michaud, 1828)	76†*	Un
<i>Raphitoma alternans</i> (Monterosato, 1884)	76*	Un
<i>Raphitoma atropurpurea</i> (Locard & Caziot, 1900) cfr.	76*, 83†‡, 84‡	Xy
<i>Raphitoma bicolor</i> (Risso, 1826)	27*, 76*, 84‡	AN, KaC, Xy
<i>Raphitoma contigua</i> (Monterosato, 1884)	88	Pro
<i>Raphitoma cordieri</i> (Payraudeau, 1826)	76*	Un
<i>Raphitoma echinata</i> (Brocchi, 1814) †	75, 76*, 84‡	AN, Xy
<i>Raphitoma farolita</i> F. Nordsieck, 1977	65, 88	Lio, Pro
<i>Raphitoma histrix</i> Bellardi, 1847 †	84*	AN
<i>Raphitoma horrida</i> (Monterosato, 1884)	65, 84‡ as <i>R. echinata</i>	Lio
<i>Raphitoma laviae</i> (R. A. Philippi, 1844)	12†*, 76*	Un
<i>Raphitoma lineolata</i> (Bucquoy, Dautzenberg & Dollfus, 1883)	12*	Un
<i>Raphitoma papillosa</i> (Pallary, 1904)	76*	Un
<i>Raphitoma philberti</i> (Michaud, 1829)	12*, 76*	Un
<i>Raphitoma pruinosa</i> (Pallary, 1906)	76*	Un
<i>Raphitoma pupoides</i> (Monterosato, 1884)	12*, 76*	Un
<i>Raphitoma purpurea</i> (Montagu, 1803)	12†*, 83†‡	Fa
<i>Taranis moerchii</i> (Malm, 1861)	76*	Un
<i>Teretia teres</i> (Reeve, 1844)	67, 76	Sof
MATHILDIDAE Dall, 1889		
<i>Mathilda gemmulata</i> O. Semper, 1865	12*, 76*, 84†‡ as <i>M. elegantissima</i> , 88	Lim, Pa
RISSEOELLIDAE Gray, 1850		
<i>Rissoella angeli</i> Manousis, 2021	77	JSP
<i>Rissoella inflata</i> (Monterosato, 1880)	12*, 76*	Un
<i>Rissoella opalina</i> (Jeffreys, 1848)	32	Pro
CORNIROSTRIDAE Ponder, 1980		
<i>Tomura depressa</i> (Granata Grillo, 1877)	27*, 77	JSP, KaC
OMALOGYRIDAE G.O. Sars, 1878		
<i>Ammonicera fischeriana</i> (Monterosato, 1869)	12*, 27*, 76*, 77	JSP, KaC
<i>Omalogyra atomus</i> (R. A. Philippi, 1841)	27*, 76*, 88	Lio, KaC
<i>Omalogyra disculus</i> Palazzi, 1988	77, 87	JSP, SPL
<i>Omalogyra simplex</i> (O. G. Costa, 1861)	27*, 77	JSP, KaC
HYALOGYRINIDAE Warén & Bouchet, 1993		
<i>Hyalogyra zibrowii</i> Warén, 1997	27	KaC
PYRAMIDELLIDAE Gray, 1840		
<i>Auristomia barashi</i> (Bogi & Galil, 2000)	89	Akr
<i>Auristomia erjaveciana</i> (Brusina, 1869)	12†*, 76†*	Un
<i>Auristomia nofronii</i> (Buzzurro, 2002)	32, 76†*	K-G, Un
<i>Brachystomia angusta</i> (Jeffreys, 1867)	27*	KaC
<i>Brachystomia carrozzai</i> (van Aartsen, 1987)	12†*, 76†*, 77	JSP
<i>Brachystomia eulimoides</i> (Hanley, 1844)	27*	KaC

<i>Brachystomia scalaris</i> (MacGillivray, 1843)	12†*, 27*, 76†*	KaC
<i>Careliopsis modesta</i> (de Folin, 1870)	77	JSP
<i>Carinorbis clathrata</i> (R. A. Philippi, 1844)	27*†, 77, 84†*	AN, KaC, Sof
<i>Chrysallida decorata</i> (R. A. Philippi, 1849)	76*	Un
<i>Cingulina isseli</i> (Tryon, 1886)	23, 62*	CKo-CKo
<i>Eulimella acicula</i> (R. A. Philippi, 1836)	12*, 27*, 76*, 77	JSP, KaC
<i>Eulimella bogii</i> van Aartsen, 1994	76	Un
<i>Eulimella cerullii</i> (Cossmann, 1916)	76*	Un
<i>Eulimella ventricosa</i> (Forbes, 1844)	76*	Un
<i>Euparthenia bulinea</i> (R. T. Lowe, 1841)	4†*, 12†*, 76*, 84†, 89	AGA, Akr, Pa, Xy
<i>Euparthenia humboldti</i> (Risso, 1826)	12†*, 27*, 76*, 89	KaC, Lio
<i>Folinella excavata</i> (R. A. Philippi, 1836)	4†*, 12†*, 27*, 76*, 77, 84†*, 87	AN, JSP, KaC, Xy
<i>Folinella ghisottii</i> van Aartsen, 1984	76	Un
<i>Liostomia clavula</i> (Lovén, 1846)	76*	Un
<i>Macrodomostomia bismichaelis</i> (Sacco, 1892) †?	4†*, 84††*	Xy
<i>Megastomia conoidea</i> (Brocchi, 1814)	4†*, 12†*, 27*, 32†, 76*, 84†, 88	AN, KaC, Lio, Pro, Xy
<i>Megastomia conspicua</i> (Alder, 1850)	27*	KaC
<i>Megastomia crovatoi</i> Nofroni, Renda, Vannozzi, 2022	25, 71*, 76*	CKo-CKo, Po-Ye
<i>Odostomella bicincta</i> (Tiberi, 1868)	77, 87	AT, JSP
<i>Odostomella doliolum</i> (R. A. Philippi, 1844)	4†*, 12†*, 27*, 76*, 77, 84†*	AN, JSP, KaC
<i>Odostomia acuta</i> Jeffreys, 1848	27*	KaC
<i>Odostomia clavulina</i> P. Fischer, 1877 cfr.	12*	Un
<i>Odostomia kromi</i> van Aartsen, Menkhorst & E. Gittenberger, 1984	12*, 27*, 76†*	KaC
<i>Odostomia lukisii</i> Jeffreys, 1859	12*, 27*, 76*	KaC
<i>Odostomia plicata</i> (Montagu, 1803)	4*, 12†*, 27*, 76*	Fa, KaC
<i>Odostomia striolata</i> Forbes & Hanley, 1850	12*, 76*, 77	JSP
<i>Odostomia turriculata</i> Monterosato, 1869	12*, 76*	Un
<i>Odostomia turrita</i> Hanley, 1844	76*	Un
<i>Odostomia unidentata</i> (Montagu, 1803)	76*	Un
<i>Ondina crystallina</i> Locard, 1891	12*, 76*	Un
<i>Ondina vitrea</i> (Brusina, 1866)	12*, 27*, 76*	KaC
<i>Ondina warreni</i> (W. Thompson, 1845)	12*, 76*	Un
<i>Oscilla galilae</i> Bogi, Karhan & Yokeş, 2012	2, 32	KB, Pro
<i>Parthenina alesii</i> Micali, Nofroni & Perna, 2012	32	CKi, K-G
<i>Parthenina angulosa</i> (Monterosato, 1889)	77	JSP
<i>Parthenina clathrata</i> (Jeffreys, 1848)	76†*	Un
<i>Parthenina decussata</i> (Montagu, 1803)	76†*	Un
<i>Parthenina emaciata</i> (Brusina, 1866)	27*, 76†*	KaC
<i>Parthenina indistincta</i> (Montagu, 1808)	4†*, 12†*, 76†*, 77	Fa, JSP
<i>Parthenina interstincta</i> (J. Adams, 1797)	4†*, 12†*, 27*, 76†*	Akr, KaC
<i>Parthenina juliae</i> (de Folin, 1872)	76†	Un
<i>Parthenina limitum</i> (Brusina, 1876)	12†*, 76†*	Un
<i>Parthenina monozona</i> (Brusina, 1869)	27*, 76†*	KaC
<i>Parthenina penchynati</i> (Bucquoy, Dautzenberg & Dollfus, 1883)	27*	KaC
<i>Parthenina suturalis</i> (R. A. Philippi, 1844)	12†*, 16, 27*, 76†*, 77	JSP, KaC
<i>Parthenina terebellum</i> (R. A. Philippi, 1844)	12†*, 76†*	Un

<i>Pyrgiscus rufus</i> (R. A. Philippi, 1836)	4†*, 12†*, 27*, 76†*, 84†, 87	Akr, AN, KaC, Xy
<i>Pyrgostylus striatulus</i> (Linnaeus, 1758)	4†, 12†*, 27*, 76†*	AN, KaC
<i>Pyrgulina microtuber</i> Peñas, Rolán & Sabelli, 2020	2	KB
<i>Pyrgulina nana</i> Hornung & Mermod, 1924	2	KB
<i>Pyrgulina pupaeformis</i> (Souverbie, 1865)	62†*	Un
<i>Syrnola fasciata</i> Jickeli, 1882	62*, 76*	Un
<i>Syrnola lendix</i> (A. Adams, 1863)	32	-
<i>Trabecula jeffreysiana</i> Monterosato, 1884	76†*	Un
<i>Tragula fenestrata</i> (Jeffreys, 1848)	76, 77	JSP
<i>Turbonilla acuta</i> (Donovan, 1804)	4†*, 12†*, 75†, 76*	Fa
<i>Turbonilla acutissima</i> Monterosato, 1884	27*, 76	KaC
<i>Turbonilla cangeyrani</i> Ovalis & Mifsud, 2017	2	Akr
<i>Turbonilla edgarii</i> (Melville, 1896)	12*, 62*, 76*	Un
<i>Turbonilla gradata</i> Bucquoy, Dautzenberg & Dollfus, 1883	12*, 76*	Un
<i>Turbonilla lactea</i> (Linnaeus, 1758)	4*, 26†*, 76*, 83, 84‡	AN, Fa, Xy
<i>Turbonilla pumila</i> G. Seguenza, 1876	12*, 27*, 76*	KaC
<i>Turbonilla pusilla</i> (R. A. Philippi, 1844)	4*, 27*, 76*, 87, 77	Fa, JSP, KaC, Xy
<i>Turbonilla rosewateri</i> Corgan & van Aartsen, 1993 cfr.	76*	Un
<i>Turbonilla sinuosa</i> (Jeffreys, 1884)	76*	Un
<i>Turbonilla syrtensis</i> van Aartsen, 1981 cfr.	16, 76*	Un
AMATHINIDAE Ponder, 1987		
<i>Amathina tricarinata</i> (Linnaeus, 1767)	20, 63*, 65	Lio
<i>Carinorbis clathrata</i> (R. A. Philippi, 1844)	4†*	AN
<i>Leucotina natalensis</i> E. A. Smith, 1910	62*	Un
<i>Monotygmma amoena</i> (A. Adams, 1853)	12*, 76†*	Un
CIMIDAE Warén, 1993		
<i>Cima cylindrica</i> (Jeffreys, 1856)	27*, 32, 76*, 77	JSP, KaC, Pro
<i>Cima minima</i> (Jeffreys, 1858)	27*, 32, 77	JSP, KaC, Pro
<i>Graphis albida</i> (Kanmacher, 1798)	27*, 89	Akr, KaC
MURCHISONELLIDAE Casey, 1904		
<i>Ebala</i> aff. <i>communis</i> Bandel, 2005	77	JSP
<i>Ebala gradata</i> (Monterosato, 1878)	27*, 32	KaC
<i>Ebala nitidissima</i> (Montagu, 1803)	76†*, 87, 77	JSP, SPL
<i>Ebala pointeli</i> (de Folin, 1868)	12†*, 27*, 76†*, 77, 89	JSP, KaC, Lio
<i>Ebala striatula</i> (Jeffreys, 1856)	76†*	Un
<i>Murchisonella mediterranea</i> Peñas & Rolán, 2013	32	Pro
ACTEONIDAE d'Orbigny, 1843		
<i>Acteon tornatilis</i> (Linnaeus, 1758)	4*, 26†*, 76*, 84	Al-Ak
<i>Crenilabium exile</i> (Jeffreys, 1870)	76*	Un
PLAKOBRANCHIDAE Gray, 1840		
<i>Elysia marginata-grandifolia</i> (Martín-Hervás et al., 2023)	15	Lat
<i>Elysia timida</i> (Risso, 1818)	4*, 65, 76*	K-G, Lio
<i>Thuridilla hopei</i> (Vérany, 1853)	4*, 76*	K-G
MNESTIIDAE Oskars, Bouchet & Malaquias, 2015		
<i>Mnestia girardi</i> (Audouin, 1826)	12†*, 16, 23†, 62†*, 65, 76†*	CKo-CKo, Lio
NOTODIAPHANIDAE Thiele, 1931, <i>taxon inquirendum</i>		
<i>Notodiaphana atlantica</i> Ortea, Moro & Espinosa, 2013	54, 76†*, 81,	K-G, Pro

RETUSIDAE Thiele, 1925		
<i>Pyrunculus fourierii</i> (Audouin, 1826)	12†*, 62*, 76*, 87	CCP
<i>Retusa crebrisculpta</i> (Monterosato, 1884)	12†*, 76†*	Un
<i>Retusa crossei</i> (Bucquoy, Dautzenberg & Dollfus, 1886)	4†*, 12†*, 76†*	Fa
<i>Retusa laevisculpta</i> (Granata Grillo, 1877)	12†*, 76†*	Un
<i>Retusa leptoneilema</i> (Brusina, 1866)	12*, 76*	Un
<i>Retusa mammillata</i> (R. A. Philippi, 1836)	12*, 76*	Un
<i>Retusa minutissima</i> (Monterosato, 1878)	16	Un
<i>Retusa tornata</i> (R. B. Watson, 1883)	77	Par
<i>Retusa truncatula</i> (Bruguière, 1792)	4, 12*, 27*, 76*, 84	Bo, Fa, KaC, Xy
<i>Retusa umbilicata</i> (Montagu, 1803)	4*, 12†*, 26*, 76†*, 77, 84*, 87	ESW, Fa, Li-Ye, Xy
<i>Retusa</i> sp.	77	JSP
RHIZORIDAE Dell, 1952		
<i>Volvulella acuminata</i> (Bruguière, 1792)	4†*, 12*, 76*, 84†*	Xy
TORNATINIDAE P. Fischer, 1883		
<i>Acteocina mucronata</i> (R. A. Philippi, 1849)	10, 12*, 23, 62*, 76†*	CKo-Cko, K-G
RINGICULIDAE R. A. Philippi, 1853		
<i>Ringicula auriculata</i> (Ménard de la Groye, 1811)	4*, 12*, 67, 76*, 84	AN, CG, Ri-Di SC
<i>Ringicula conformis</i> Monterosato, 1877	4*, 12*cfr., 76*, 84*	AN, CG, GB, Fa, Pa, Pro
BULLIDAE Gray, 1827		
<i>Bulla arabica</i> Malaquias & D. Reid, 2008	78*, 35‡as <i>B. ampulla</i> , 89	Pro
<i>Bulla striata</i> Bruguière, 1792	4*, 12*, 26†*, 27*, 76†*, 83, 84†	Al-Ak, AN, Bo, Fa, KaC, Lim
HAMINOEIDAE Pilsbry, 1895		
<i>Haminoea hydatis</i> (Linnaeus, 1758)	4*, 12*, 26*, 27*, 76*, 83, 84	Akr, Fa, KaC, Xy
<i>Haminoea navicula</i> (daCosta, 1778)	4*, 12*, 27*, 76*, 84	Akr, KaC, LM
<i>Haminoea orbignyana</i> (A. Férussac, 1822)	12*, 76*	Un
<i>Lamprohaminoea ovalis</i> (Pease, 1868)	3†, 33†*, 90†	CG, N, Pro
<i>Liloea mongii</i> (Audouin, 1826) cfr.	12†, 33*	K-G
<i>Roxaniella jeffreysi</i> (Weinkauff, 1866)	12†*, 27*, 76†*, 88	Lio, KaC
<i>Weinkauffia macandrewii</i> (E. A. Smith, 1872)	24, 27*, 65, 68†, 84‡ as <i>C. cylindracea</i>	Lio, KaC, Pro
<i>Weinkauffia turgidula</i> (Forbes, 1844)	12*, 76*	Un
PHILINIDAE Gray, 1850		
<i>Philine catena</i> (Montagu, 1803)	4*, 27*, 76*, 87	Akr, Fa, KaC, Li-Ye
<i>Philine punctata</i> (J. Adams, 1800)	4*, 76*	Fa
<i>Philine quadripartita</i> Ascanius, 1772	4†, 12†*, 26†*, 76†*, 83†, 84†	AGA, Akr, Al-Ak, Fa, Xy
SCAPHANDRIDAE G. O. Sars, 1878		
<i>Scaphander lignarius</i> (Linnaeus, 1758)	76*	Un
AGLAJIDAE Pilsbry, 1895		
<i>Biuve fulvipunctata</i> (Baba, 1938)	27*, 53, 85†	AGA, KaC, Lim, Pro
<i>Melanochlamys algirae</i> (A. Adams, 1850)	12†*, 76†*	Un
<i>Philineopsis depicta</i> (Renier, 1807)	4†, 76*	Fa
ALACUPPIDAE Oskars, Bouchet & Malaquias, 2015		
<i>Roxania utriculus</i> (Brocchi, 1814)	4, 76*	Fa

GASTROPTERIDAE Swainson, 1840		
<i>Gastropteron rubrum</i> (Rafinesque, 1814)	76†*	Un
CYLICHNIDAE H. Adams & A. Adams, 1854		
<i>Cylichna cylindracea</i> (Pennant, 1777)	4*, 76*, 84‡	Akr, Pa
COLINATYDIDAE Oskars, Bouchet & Malaquias, 2015		
<i>Colinatys alayoi</i> (Espinosa & Ortea, 2004) cfr.	53	Un
CAVOLINIIDAE Gray, 1850 (1815)		
<i>Cavolinia gibbosa</i> (A. d'Orbigny, 1835)	76*	Un
<i>Cavolinia inflexa</i> (Lesueur, 1813)	27*	KaC
<i>Cavolinia tridentata</i> (Forsskål, 1775)	4*, 12*, 13, 27*, 76*, 84	KaC
CLIIDAE Jeffreys, 1869		
<i>Clio cuspidata</i> (Bosc, 1801)	4*, 76*, 84*	Un
<i>Clio pyramidata</i> Linnaeus, 1767	4*, 12*, 27*, 76*, 84*	Fa, KaC
CRESEIDAE Rampal, 1973		
<i>Creseis acicula</i> (Rang, 1828)	27*, 76*	KaC
<i>Styliola subula</i> (Quoy & Gaimard, 1827)	4*, 12*, 27*, 76*, 77	JSP, KaC, KY-Ku
HYALOCYLIDAE A. W. Janssen, 2020		
<i>Hyalocylis striata</i> (Rang, 1828)	76*	Un
LIMACINIDAE Gray, 1840		
<i>Heliconoides inflatus</i> (A. d'Orbigny, 1835)	27*, 76†*, 89	JSP, KaC
<i>Limacina trochiformis</i> (A. d'Orbigny, 1835)	13*, 27*, 67, 76*	JSP, KaC
OXYNOIDAE Stoliczka, 1885		
<i>Lobiger serradifalci</i> (Calcara, 1840)	4*, 76*	Fa
<i>Oxynoe olivacea</i> Rafinesque, 1814	4, 76*, 84*	Akr, Fa
VOLVATELLIDAE Pilsbry, 1895		
<i>Ascobulla fragilis</i> (Jeffreys, 1856)	76*	Un
UMBRACULIDAE Dall, 1889		
<i>Umbraculum umbraculum</i> ([Lightfoot], 1786)	4†, 26†*, 45, 76*, 83†, 84†	AN, K-K, Pa, Pro, Y-Y
TYLODINIDAE Gray, 1847		
<i>Tylodina perversa</i> (Gmelin, 1791)	12*, 27*, 60, 67, 76*	Akr, KaC, Lio
PLEUROBRANCHAEIDAE Pilsbry, 1896		
<i>Pleurobranchaea meckeli</i> (Blainville, 1825)	4*, 26*, 76*	Akr, Fa, NC
PLEUROBRANCHIDAE Gray, 1827		
<i>Berthella ocellata</i> (Delle Chiaje, 1830)	4*, 76*	CKi
<i>Berthella plumula</i> (Montagu, 1803)	27*	KaC
<i>Berthellina citrina</i> (Rüppell & Leuckart, 1828)	5, 12*, 41, 76†*	CG
<i>Berthellina engeli</i> Gardiner, 1936, <i>taxon inquirendum</i>	4*, 76*	AGA, CKi
<i>Pleurobranchus membranaceus</i> (Montagu, 1816)	4†*, 76*, 84	Fa
<i>Pleurobranchus testudinarius</i> Cantraine, 1835	4†*, 76*	Fa
AKERIDAE Mazzarelli, 1891		
<i>Akera bullata</i> O. F. Müller, 1776	4*, 76*, 84*	Fa, Y-Y
APLYSIIDAE Lamarck, 1809		
<i>Aplysia dactylomela</i> Rang, 1828	23, 39, 62*	O-K, Pro
<i>Aplysia depilans</i> Gmelin, 1791	4*, 26*, 75, 76*, 84	Pa
<i>Aplysia fasciata</i> Poiret, 1789	4*, 12*, 27*, 46, 76*, 55, 84*	AGA, KaC

<i>Aplysia parvula</i> Mörch, 1863	4*, 76*	CAA-ZB, Fa
<i>Aplysia punctata</i> (Cuvier, 1803)	4*, 76*	AGA, Fa
<i>Bursatella leachii</i> Blainville, 1817	9, 78*	
<i>Notarchus punctatus</i> R. A. Philippi, 1836	4*, 27*, 76*	CAA-ZB, Fa, KaC
<i>Petalifera gravieri</i> (Vayssière, 1906)	12*, 76*, 87	SPL
<i>Petalifera petalifera</i> Rang, 1828)	27*	KaC
CHROMODORIDIDAE Bergh, 1892		
<i>Felimare picta</i> (R. A. Philippi, 1836)	4†*, 43	AN, NC
<i>Felimare tricolor</i> (Cantraine, 1835)	76†*	Un
<i>Felimare Villafranca</i> (Risso, 1818)	4†*, 76†*	NC
<i>Goniobranchus annulatus</i> (Eliot, 1904)	47, 53, 85	Dh, La, Pro
<i>Hypselodoris infucata</i> (Rüppell & Leuckart, 1830)	53, 85	Am, Lim
DISCODORIDIDAE Bergh, 1891		
<i>Peltodoris atromaculata</i> Bergh, 1880	4, 26†*, 38, 76†*	Akr, CP, Dh, NC, Pro
<i>Tayuva lilacina</i> (A. Gould, 1852)	78*	
AEGIRIDAE P. Fischer, 1883		
<i>Aegires punctilucens</i> (A. d'Orbigny, 1837)	4, 76*	Fa
DENDRODORIDIDAE O'Donoghue, 1924		
<i>Dendrodoris areolata</i> (Alder & Hancock, 1864)	4	PP, Va
<i>Dendrodoris grandiflora</i> (Rapp, 1827)	4*, 75, 76*	AGA, Bo
<i>Doriopsilla areolata</i> Bergh, 1880	76*	Un
TETHYDIDAE Rafinesque, 1815		
<i>Melibe viridis</i> (Kelaart, 1858)	62*, 53, 85	Am, Lim
<i>Tethys fimbria</i> Linnaeus, 1767	4*, 76*	Fa
POLYCERIDAE Alder & Hancock, 1845		
<i>Kaloplocamus ramosus</i> (Cantraine, 1835)	4*, 76*	Fa
<i>Plocamopherus ocellatus</i> Rüppell & Leuckart, 1828	18, 34*	AN, Pro
JANOLIDAE Pruvot-Fol, 1933		
<i>Antiopella cristata</i> (Delle Chiaje, 1841)	4†*, 76†*	Fa, K-G
AEOLIDIIDAE Gray, 1827		
<i>Spurilla neapolitana</i> (Delle Chiaje, 1841)	50	Pro
FACELINIDAE Bergh, 1889		
<i>Cratena peregrina</i> (Gmelin, 1791)	42, 56	La
FLABELLINIDAE Bergh, 1889		
<i>Coryphellina rubrolineata</i> O'Donoghue, 1929	53†, 85	La
<i>Edmundsella pedata</i> (Montagu, 1816)	51	La
<i>Flabellina affinis</i> (Gmelin, 1791)	4*, 40, 32*, 84†	La, NC
SIPHONARIIDAE Gray, 1827		
<i>Siphonaria crenata</i> Blainville, 1827	33*, 65	Ri-Di
<i>Siphonaria pectinata</i> (Linnaeus, 1758)	76*, 84†*	Pa
<i>Williamia gussoni</i> (O. G. Costa, 1829)	12*, 27*, 76*, 84‡ as <i>A. lateralis</i> , 87	KaC, SPL
TRIMUSCULIDAE J. Q. Burch, 1945 (1840)		
<i>Trimusculus mammillaris</i> (Linnaeus, 1758)	12*, 26†*, 76*, 83†, 84	AN, Pa, Pro
ELLOBIIDAE L. Pfeiffer, 1854		
<i>Leucophytia bidentata</i> (Montagu, 1808)	65†, 76†*	Lio
<i>Myosotella myosotis</i> (Draparnaud, 1801)	12†*, 26†*, 76†*, 83, 84†	Pro, Xy
<i>Ovatella firminii</i> (Payraudeau, 1826)	12*, 76*, 87	ATS

POLYPLACOPHORA		
ACANTHOCHITONIDAE Pilsbry, 1893		
<i>Acanthochitona crinita</i> (Pennant, 1777)	12*, 76*	Un
<i>Acanthochitona fascicularis</i> (Linnaeus, 1767)	12*, 26+*, 27*, 76*	KaC, K-K
<i>Acanthochitona oblonga</i> Leloup, 1981	7*, 19	Un
CALLOCHITONIDAE Plate, 1901		
<i>Callochiton septemvalvis</i> (Montagu, 1803)	12*, 26+*, 27*, 76+*	KaC, NC
CHITONIDAE Rafinesque, 1815		
<i>Rhyssoplax corallina</i> (Risso, 1826)	12+*, 27*, 76+*	KaC
<i>Rhyssoplax olivacea</i> Spengler, 1797	12+*, 26+*, 27*, 75+, 76+*, 84+	KaC
<i>Rhyssoplax phaseolina</i> (Monterosato, 1879)	12+*, 76+*	Un
ISCHNOCHITONIDAE Dall, 1889		
<i>Ischnochiton rissoi</i> (Payraudeau, 1826)	12*, 27*, 76+*	KaC
LEPTOCHITONIDAE Dall, 1889		
<i>Lepidopleurus cajetanus</i> (Poli, 1791)	12*, 27*, 75, 76*, 84	KaC
<i>Leptochiton bedullii</i> Dell'Angelo & Palazzi, 1986	27*, 76+*	KaC
<i>Leptochiton boettgeri</i> (Šulc, 1934) †	12*	Un
<i>Leptochiton cancellatus</i> (G. B. Sowerby II, 1840)	76+*, 84+	Un
TONICELLIDAE Simroth, 1894		
<i>Lepidochitona cinerea</i> (Linnaeus, 1767)	27*	KaC
<i>Lepidochitona furtiva</i> (Monterosato, 1879)	27*	KaC
<i>Lepidochitona monterosatoi</i> Kaas & VanBelle, 1981	27*	KaC
SCAPHOPODA		
DENTALIIDAE Gray, 1847		
<i>Antalis dentalis</i> (Linnaeus, 1758)	12+*, 75+, 76*, 83+, 84+	Fa
<i>Antalis entalis</i> (Linnaeus, 1758)	12+*, 76*	Un
<i>Antalis inaequicostata</i> (Dautzenberg, 1891)	75+	
<i>Antalis novemcostata</i> (Lamarck, 1818)	89	Zy
<i>Antalis panorma</i> (Chenu, 1843)	12+*, 76*, 84+*	Un
<i>Antalis rossati</i> (Caprotti, 1966)	65, 76*	Lio
<i>Antalis vulgaris</i> (daCosta, 1778)	12+*, 26+*, 27*, 75+, 76*, 83+, 84+	Co, KaC
FUSTIARIIDAE Steiner, 1991		
<i>Fustiaria rubescens</i> (Deshayes, 1825)	26+*, 27*, 75, 76*, 83+, 84+	KaC
ENTALINIDAE Chistikov, 1979		
<i>Entalina tetragona</i> (Brocchi, 1814)	76*	Un
GADILIDAE Stoliczka, 1868		
<i>Dischides politus</i> (S. V. Wood, 1842)	76*	Un

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